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# The wasps, bees and ants (Insecta: Vespida = Hymenoptera) from the Insect Limestone (Late Eocene) of the Isle of Wight, UK

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**ABSTRACT:** The types and undescribed material of the hymenopteran fossils of the Insect Bed of the Bembridge Marls from the Isle of Wight (UK) are critically revised and studied. A total of 1460 fossils are recorded and attributed to 20 families: Gasteruptionidae s.l. (1); Proctotrupidae (3); Diapriidae (24); Cynipidae (7); Figitidae (6); Pteromalidae (1); Agaonidae (3); Scelionidae (12); Platygasteridae (2); Ichneumonidae (32); Braconidae (75); Bethylinidae (3); Crabronidae (2); Sphecidae (1); Apidae (2); Scolidae (1); Tiphidae (2); Vespidae (4); and Formicidae (1220). Described as new are 51 species, 13 genera, two tribes and two subfamilies. Minimum number of species recorded (either as described species or representing higher taxa with no described species in the assemblage) is 118. The composition of the hymenopteran assemblage is most similar to that of Baltic amber and indicative of a well forested territory, as well as of a humid, equable (aseasonal but not very hot) climate, more typically equable than in the Baltic amber source area, judging from the absence of Aphidinae and scarcity of aphids.

**KEY WORDS:** Bembridge Marls, Eocene, fossil insects, new combinations, new taxa



Material for the present study comprises the hymenopteran fossils preserved in concretions or tabular bands of very fine-grained micrite, known as Insect Limestone. The unit where these concretions/bands occur is known as the Insect Bed, which lies towards the base of the Bembridge Marls Member (Solent Group: Bouldnor Formation). The Bembridge Marls are dated as the latest Eocene (Hooker *et al.* 2009). The deposits outcrop in several sites along the north coast of the Isle of Wight near the south coast of Great Britain.

Most of the specimens studied are kept at the Natural History Museum (NHMUK). The majority of them belong to the collections of A.Court Smith (purchased 1877, 1883), P.B. Brodie (purchased 1898) and R.W. Hooley (purchased 1924). They are labelled 'Gurnard Bay' or 'Gurnet Bay' (which is an old name for Gurnard Bay). However, Smith collected specimens all the way from West Cowes to Newtown River on the northwest side of the Isle of Wight (Jarzemkowski 1980). Most of the specimens probably came from Thorness Bay (Jarzemkowski 1976). Brodie and Hooley acquired parts of Smith's collection, so parts and counterparts of individual insects have turned up in all three collections. The parts and counterparts often have different numbers, because they were registered

at different times. An additional collection was recently discovered at the Sedgwick Museum, Cambridge, by A.J. Ross. This collection has also yielded counterparts of specimens at the NHMUK which indicates that this is another part of the Smith collection. A label with '1883' on it suggests that the Sedgwick Museum acquired this collection in 1883, the same year in which the NHMUK purchased specimens from Smith.

A small collection of the fossils studied, referred to as USNM, is kept at the National Museum of Natural History, Smithsonian Institution, Washington, DC, USA. These are mostly the type material described by Cockerell (1915). They come from the same deposits.

Hymenopterans are the second most common insect order (after dipterans) in the Bembridge assemblage. Even if their high participation depends in part on the hyper-abundance of two species of a single ant genus (*Oecophylla* Smith, 1860 which is responsible for at least 40 per cent of the hymenopteran fossils), the order diversity is impressive, with its 20 families, over 100 genera and more than 120 species recorded conservatively after this non-exhaustive study of the available fossil material.

**Table 1** Current taxonomic position of the previously described species of the Bembridge fossil hymenopterans

Taxa as described	Current taxonomic position
<i>Acourtia perplexa</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: Townesitinae: <i>Acourtia perplexa</i> Cockerell, 1921
<i>Andricus vectensis</i> Cockerell, 1921, Cynipidae	not revised
<i>Aneurhynchus conservatus</i> Cockerell, 1915, Diapriidae	not studied
<i>Bathystomus peritus</i> Cockerell, 1921, Braconidae	Braconidae: Opiinae: <i>Biosteres peritus</i> (Cockerell, 1921)
<i>Calyptoides veterinus</i> Cockerell, 1921, Braconidae	Braconidae: Helconinae: <i>Calyptoides veterinus</i> Cockerell, 1921
<i>Camponotus (Colobopsis) brodiei</i> Donisthorpe, 1920, Formicidae	Formicidae incertae sedis: “ <i>Camponotus</i> ” <i>brodiei</i> Donisthorpe, 1920:
<i>Coleocentrus gurnetensis</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: subfam. indet.: “ <i>Coleocentrus</i> ” <i>gurnetensis</i> Cockerell, 1921
<i>Cremastus? arcuatus</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: Orthocentrinae, Microleptini: <i>Eusterinx arcuatus</i> (Cockerell, 1921)
<i>Dacnusites reductus</i> Cockerell, 1921, Braconidae	Braconidae: Microgastrinae: <i>Dacnusites reductus</i> Cockerell, 1921
<i>Dacnusites sepultus</i> Cockerell, 1921, Braconidae	Braconidae: Microgastrinae: <i>Dacnusites sepultus</i> Cockerell, 1921
<i>Diospiloides hooleyi</i> Cockerell, 1921, Braconidae	Braconidae: Orgilinae: <i>Microtypus hooleyi</i> (Cockerell, 1921)
<i>Diospilus acourti</i> Cockerell, 1921, Braconidae	Braconidae: Helconinae: <i>Diospilus acourti</i> Cockerell, 1921
<i>Dolichoderus anglicus</i> Cockerell, 1915, Formicidae	Formicidae: Aneuretinae: <i>Britaneuretus anglicus</i> (Cockerell, 1915)
<i>Dolichoderus britannicus</i> Cockerell, 1915, Formicidae	Formicidae: Dolichoderinae: <i>Emplastus britannicus</i> (Cockerell, 1915)
<i>Dolichoderus gurnetensis</i> Donisthorpe, 1920, Formicidae	Formicidae: Dolichoderinae: <i>Emplastus gurnetensis</i> (Donisthorpe, 1920)
<i>Dolichoderus ovigerus</i> Cockerell, 1915, Formicidae	Formicidae: Dolichoderinae: <i>Emplastus britannicus</i> (Cockerell, 1915)
<i>Dolichoderus vectensis</i> Donisthorpe, 1920, Formicidae	Formicidae: Dolichoderinae: <i>Dolichoderus vectensis</i> Donisthorpe, 1920
<i>Emplastus emeryi</i> Donisthorpe, 1920, Formicidae	Formicidae: Dolichoderinae: <i>Emplastus britannicus</i> (Cockerell, 1915)
<i>Euponera crawleyi</i> Donisthorpe, 1920, Formicidae	Formicidae: Ponerinae: <i>Ponerites crawleyi</i> (Donisthorpe, 1920)
<i>Holomeristus? vectensis</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: Orthocentrinae, Microleptini: <i>Eusterinx vectensis</i> (Cockerell, 1921)
<i>Ichneumon acourti</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: Cryptinae: “ <i>Hemiteles</i> ” <i>acourti</i> (Cockerell, 1921)
<i>Ipobracon? micrarche</i> Cockerell, 1921, Braconidae	Braconidae: Braconinae: <i>Bracon micrarche</i> (Cockerell, 1921)
<i>Ipobracon? vectensis</i> Cockerell, 1921, Braconidae	Braconidae: Braconinae: <i>Cyanopterus vectensis</i> (Cockerell, 1921)
<i>Itopectis saxosus</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: Pimplinae: “ <i>Itopectis</i> ” <i>saxosa</i> (Cockerell, 1921)
<i>Lampronota disrupta</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: Townesitinae: <i>Marjorietta disrupta</i> (Cockerell, 1921)
<i>Leptothorax gurnetensis</i> Cockerell, 1915, Formicidae	Formicidae: Formicinae: <i>Leucotaphus gurnetensis</i> (Cockerell, 1915)
<i>Leucotaphus cockerelli</i> Donisthorpe, 1920, Formicidae	Formicidae: Formicinae: <i>Camponotus cockerelli</i> (Donisthorpe, 1920)
<i>Leucotaphus permanus</i> Cockerell, 1927, Formicidae	not studied (type lost)
<i>Lithapechtis fumosus</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: subfam. indet.: <i>Lithapechtis fumosus</i> Cockerell, 1921
<i>Lithobelyta reducta</i> Cockerell, 1921, Diapriidae	Chalcidoidea: <i>Lithobelyta reducta</i> Cockerell, 1921
<i>Macroteleia veterina</i> Cockerell, 1921, Scelionidae	Scelionidae: Scelioninae: <i>Calliscelio veterinus</i> (Cockerell, 1921)
<i>Mesitius? rectinervis</i> Cockerell, 1921, Bethyridae	not revised
<i>Miota? strigata</i> Cockerell, 1921, Diapriidae	not revised
<i>Oecophylla atavina</i> Cockerell 1915, Formicidae	Formicidae: Formicinae: <i>Oecophylla atavina</i> Cockerell 1915
<i>Oecophylla megarche</i> Cockerell, 1915, Formicidae	Formicidae: Formicinae: <i>Oecophylla megarche</i> Cockerell, 1915
<i>Oecophylla perdita</i> Cockerell, 1915, Formicidae	Formicidae: Formicinae: <i>Oecophylla atavina</i> Cockerell 1915
<i>Phanomeris? colenutti</i> Cockerell, 1921, Chalcidoidea	not revised
<i>Philoponites clarus</i> Cockerell, 1915, Sphecidae	Tiphidae: Tiphinae: <i>Philoponites clarus</i> Cockerell, 1915
<i>Polybia oblita</i> Cockerell, 1921, Vespidae	Vespidae: Polistinae: <i>Protopolistes oblitus</i> (Cockerell, 1921)
<i>Polybia? anglica</i> Cockerell, 1921, Vespidae	Vespidae: Polistinae: <i>Palaeopolybia anglica</i> (Cockerell, 1921)
<i>Polyclistus anglicus</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: Metopiinae: <i>Hypsicera anglica</i> (Cockerell, 1921)
<i>Polysphincta? atavina</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: subfam. & gen. indet.: “ <i>Polysphincta</i> ” <i>atavina</i> Cockerell, 1921
<i>Ponera hypolitha</i> Cockerell, 1915, Formicidae	Formicidae: Dolichoderinae: <i>Emplastus hypolithus</i> (Cockerell, 1915)
<i>Ponera minuta</i> Donisthorpe, 1920, Formicidae	Agaonidae: “ <i>Ponera</i> ” <i>minuta</i> Donisthorpe, 1920
<i>Pteromalus? vectensis</i> Cockerell, 1921, Pteromalidae	not revised
<i>Rhodites vetus</i> Cockerell, 1921, Cynipidae	not revised
<i>Sceliphron? brevior</i> Cockerell, 1921, Sphecidae	Sphecidae: Protosceliphriini: <i>Protosceliphron brevior</i> (Cockerell, 1921)
<i>Sigalphus cervicalis</i> Cockerell, 1921, Braconidae	Braconidae: Helconinae: <i>Taphaeus cervicalis</i> (Cockerell, 1921)
<i>Stilpnus oligocenus</i> Cockerell, 1921, Ichneumonidae	Ichneumonidae: Cryptinae: “ <i>Stilpnus</i> ” <i>oligocenus</i>
<i>Syntaphus wheeleri</i> Donisthorpe, 1920, Formicidae	Braconidae: Cheloninae: <i>Ascogaster (Syntaphus) wheeleri</i> (Donisthorpe, 1920)
<i>Vectevania vetula</i> Cockerell, 1922, (Evaniidae)	Gasteruptionidae: Aulacinae?: <i>Vectevania vetula</i> Cockerell, 1922
<i>Xenarcha? distracta</i> Cockerell, 1921 Braconidae	Braconidae: Doryctinae: <i>Ontsira distracta</i> (Cockerell, 1921)
<i>Zygota? filicornis</i> Cockerell, 1921, Diapriidae	not revised

The taxonomic history of the Bembridge hymenopteran fossils is short, in the sense that all descriptions predating the present project were made during a short interval of twelve years (Cockerell 1915–1927; Donisthorpe 1920) (Table 1).

In the framework of the present project, the following hymenopteran taxa recorded from the Bembridge Marls are thoroughly studied: aculeate wasps (except Chrysidoidea) by

Dlussky and Prefilieva (ants) and Antropov (all others); Ichneumonoidea by Belokobylskij (Braconidae) and Khalaim (Ichneumonidae); Gasteruptionidae by Rasnitsyn; and Proctotrupidae by Kolyada. Diverse Proctotrupomorpha were being studied by the late Mikhail Kozlov, whose untimely death on 11 September 2006 has left this important group without thorough description (except for Proctotrupidae). For the groups

left unattended, there are preliminary identifications made by Kozlov and completed in part by Kolyada and Compton. Because of different styles in description, and particularly in morphological terminology, including symbols denoting various structures employed in different hymenopteran taxa, unification throughout has not been attempted.

The general compilation and editing of manuscripts by other participants, as well as this introduction, are by Rasnitsyn. The closing Discussion is written by Rasnitsyn, based on the results by other participants in the project.

## 1. Overview of the hymenopteran taxa found in the Bembridge Marls

Symphyta were not found. The absence of Tenthredinoidea is indicative of a warm climate (possibly warmer than in both Baltic amber and Florissant).

The only record of Evaniomorpha is *Vectevania vetula* Cockerell, 1922 (Gasteruptiidae: Aulacinae), a supposed parasite of xylophilous insect larvae, indicative of forested territory.

Proctotrupomorpha are moderately rich but unfortunately poorly studied. Proctotrupeoidea are known from Proctotrupidae (three fossils, described *Oxyserphus kozlovi* Kolyada, sp. nov. and *Mischoserphus* sp., both in the tribe Cryptoserphini, parasitising curculionoid beetles and dipteran larvae, respectively, mainly in mesic forests; see below) and Diapriidae (24 fossils). Earlier described Diapriidae are *Zygota? filicornis* Cockerell, 1921a and *Miota? strigata* Cockerell, 1921a; additional identifications (by Masner and Kolyada in 2006) are Ambositrininae? gen. sp., *Basalys* sp., *Belyta?* sp., *Lyteba* (?) sp., *Pantoclis* (?) sp., Spilomicrini gen. sp. and *Trichopria* sp. Diapriidae are parasites of dipteran larvae preferring mesic environments; Ambositrininae are mainly Gondwanan in their distribution (Australasian and Neotropical with one Afrotropical species known in southern Africa and Madagascar), with one species in North America, and known also in Baltic amber, with one Baltic species hardly distinguishable from an extant African one (Masner 1969).

Cynipoidea are known from 9–11 Cynipidae fossils, including *Andricus vectensis* Cockerell, 1921a and *Rhodites vetus* Cockerell, 1921a, and 6–8 Figitidae. Cockerell's generic identifications are not reliable enough to be used for further inferences.

Platygastridae are found with both living families Platygastridae (two fossils, including *Inostemma* sp., a genus of gall midge parasites, identified by both Kozlov and Masner) and Scelionidae. The latter comprise 12 fossils, including "*Macroteleia*" *veterna* Cockerell, 1921a = *Caloteleia*, a widespread genus of orthopteran parasites; an undescribed genus (determined by Kozlov) near *Apegus* Foerster, 1856, a Palaearctic and Oriental genus whose relatives parasitise tettigoniid eggs on trees; *Gryon* sp., a widespread genus attacking heteropteran eggs; Scelionini gen. sp., parasites on acridid eggs; Baeinae gen. sp. a group of spider egg parasites, and *Calliscelio* sp. (all determined by Masner). Scelionidae are a more xerophilous group than Diapriidae, and the modest participation of this family compared to the latter indicates generally mesic past environments.

Chalcidoidea fossils (15–16 specimens) remain poorly known. Three species are described, one originally as an ant *Ponera minuta* Donisthorpe, 1920, another as a diapriid *Lithobelyta reducta* Cockerell, 1921a, and only the third as a chalcidoid *Pteromalus? vectensis* Cockerell, 1921a. The latter might be a pteromalid, whilst the former, along with two other fossils, are redescribed below as representatives of the family of fig pollinators, Agaonidae (first identified by Kozlov). The family identity of *Lithobelyta reducta* needs clarification.

The superfamily Ichneumonoidea is particularly rich in the Bembridge Marls, yielding more than one hundred fossils (minimum 32 Ichneumonidae and 75 Braconidae) of more than 60 species (minimum 25 and 36, respectively). Detailed information about these insects is presented below.

Aculeate hymenopterans other than ants are not very diverse. This is particularly true for Chrysidoidea, represented by only three fossils of the family Bethyridae, which remain mostly unstudied. One described is *Mesitius? rectinervis* Cockerell, 1921a.

The higher wasps and bees (Aculeata s. str.) are fully revised here (see below). Except for ants, they are only moderately diverse. Apoid wasps and bees are known from two bees (one Apidae: Apinae, another unidentifiable Megachilidae) and three wasps (one Sphecidae and two Crabronidae, the latter indicative of a forested landscape). Vespoidea (s.l.) wasps are recorded from three families: Tiphidae (two fossils), Scoliidae (one) and Vespidae (four; one of these shows (sub)tropical affinities). See below for details.

Ants (Formicidae) are the most abundant and most indicative group in the Bembridge hymenopteran assemblage and the third most diverse one (some 1220 fossils, 20 species and ten genera). They comprise both thermophilous and temperate species, whose co-existence implies most convincingly the equable climate, and the high proportion of dendrobiotic or otherwise forest-dwelling ants infers a well forested source area. The ant assemblage is undoubtedly more related to the Baltic amber fauna than to Florissant, but this similarity might be geographically rather than geochronologically driven (for details see below).

**Institutional repository abbreviations.** CAMSM, Sedgwick Museum of Earth Sciences, University of Cambridge; IWCMS, Isle of Wight County Museum Service; NHMUK, Department of Palaeontology, Natural History Museum, London; USNM, Department of Paleobiology, National Museum of Natural History, Smithsonian Institution, Washington DC.

## 2. Material and methods

The fossils were studied using the usual observation under a stereomicroscope, except that the application of any liquid was avoided because of salt crystals in the rock matrix, often within the fossil itself. Instead of liquid, polarising illumination was used to enhance visibility of the fossilised organic material when preserved. A.V. Antropov used Carl Zeiss Stemi SV 6 and Leica MZ 9.5 stereomicroscopes for study of the fossils, and Nikon Coolpix 885 / Nikon Coolpix 4500 digital cameras to make photographs, which were subsequently improved when necessary using Adobe Photoshop 7.0 software. The line drawings were prepared using CorelDRAW 7 software. All measurements were made with the help of an ocular-micrometer. G. M. Dlussky and K. S. Perfilieva used the Olympus C-4000 Zoom digital camera, and enlarged prints were hand-traced with a pen with the visual control of the specimen under a Leica S6E stereomicroscope. The resulting draft drawing was scanned with ScanExpress 6000 PS and improved finally using the program CorelDRAW 8. V.A. Kolyada used the Wuzhou XTL-3400E stereomicroscope and a Canon PowerShot S50 camera, with photographs automounted with Helicon Focus software and adjusted with Adobe Photoshop 6.0; line drawings were made with Adobe Illustrator 10 software. A. P. Rasnitsyn examined the fossils using LOMO MPS-2 and Leica MZ 9.5 microscopes and prepared photographs using a Nikon Coolpix4500 camera automounted with Helicon Focus software, and the line drawings were prepared using CorelDRAW 11 software.

### 3. Systematic palaeontology

#### 3.1. Superfamily Evanioidea Latreille, 1802

[By Alexandr P. Rasnitsyn.]

*Vectevania vetula* Cockerell, 1922 has been very briefly described based on the unique holotype collected in the Bembridge Marls. Cockerell (1922) ascribed the fossil to the family Evaniidae, which at that time was equivalent to the current superfamily Evanioidea, and compared it with *Hyptiogaster*, which is now attributed to the family Gasteruptiidae s.str. and subfamily Hyptiogastrinae (Jennings & Austin 2000). This fossil remains the only Cenozoic record of Gasteruptiidae s.str. (Nel *et al.* 2004) and so deserves close attention.

The unique holotype (obverse part only) NHMUK In.20535 is kept at the Natural History Museum, London. As usual in the Bembridge Marls, the fossil (Text-fig. 1A) is perfectly three-dimensional, with the body cavity empty, and a split within the body more or less along the wing plane. The counterpart is lost, and the only part at hand shows the internal ventral surface of the insect seen from above. The head cavity is only slightly open dorsally, showing no details. The antennae, legs and metasomal apex are not preserved, nor is practically all of the dorsal surface, except for the 1st metasomal acrosegment seen from above and sidewise, and a small subterminal tergal surface of the metasoma. This restricts the availability of taxonomically meaningful structures mainly to wing venation.

Family Gasteruptiidae Ashmead, 1900

Subfamily Aulacinae Hedicke, 1939

Genus *Vectevania* Cockerell, 1922

**Type species.** *V. vetula* Cockerell, 1922 (by monotypy).

**Diagnosis.** Propleura short. Forewing not folded longitudinally as preserved, with cell 1mcu reversely triangular (RS+M meeting basal vein at its lower corner, 1m-cu short but distinct), 2r-m spectral, 3r-m apparently lost but leaving signs of its former position in slight angulation of RS and M, 2m-cu tubular. Hind wing with C tubular. Metasomal sterna flat, indicating depressed metasoma, with apparent metasomal segment 1 triangular with sides slightly convex (fore part not forming petiole). No trace of possible fusion of two primary sterna discernible.

**Remark.** Taxonomic position of the genus deserves an extensive discussion which is not appropriate herein (cf. Rasnitsyn 2013).

*Vectevania vetula* Cockerell, 1922

Text-fig. 1

1922 *Vectevania* Cockerell, p. 34, fig. 1.

1992 *Vectevania* Carpenter, p. 474.

**Holotype.** NHMUK In.20535, Bembridge Marls, NW Isle of Wight, UK, Smith Coll.; ventral body interior and left wing pair, no appendages; sex unknown.

**Description.** Head apparently large, transverse. Pronotum as preserved arching, not armed. Propleurae short, at most slightly extending beyond fore pronotal contour. Mesopectus with internal surface smooth with few and ordinary features (discrimen, pleural sutures, and bend delimiting anterior (coxae-faced) surface). Forewing with pterostigma large semi-circular. Basal vein composed almost entirely of long 1RS slightly arching in lower half; 1M rudimentary, aligned with 1RS+M thus making 1mcu cell reversal triangular (with narrow end directed basally), 2RS+M short (about as long as 1m-cu), with RS and M diverging almost symmetrically at its distal end. Cross-vein 2r-rs c.0.7 times as long as pterostigma

width, meeting pterostigma at its midlength, slightly inclined posterodistally, meeting RS at about its length basal of 2r-m, 2r-m spectral, meeting M slightly basal of 2m-cu. Cell 2rm subtriangular, almost twice as long as wide. Cell 3r-m, as defined by angulations of RS and M at junctions with former 3r-m, subquadrate. M getting thinner beyond former 3r-m, but apparently persist tubular and coloured. Cell 2m-cu receiving Cu at its midlength basally, 2m-cu c. half as long as 2m-cu maximum height. Crossvein cu-a mirroring 1RS in meeting Cu close to M+Cu fork and being inclined posterodistally at similar angle. Anal vein thick (apparently thicker than R), slightly sinuate. Hind wing with C and R tubular and the rest veins spectral and of usual form. Metasoma elongate tear-shaped, with sterna flat, six in number, implying either no sternal fusion or male sex in case of traceless fusion of sterna 1 and 2. Metasomal 1st acrotergite small and not much modified in side view, 1st apparent sternum beyond acrosternite of subequal length and width, longer than two following sterna combined; following sterna short but of irregular length, possibly being telescoped irregularly. No distinct surface sculpture or colouration of ventral body integuments preserved. Forewing length c. 4.7 mm, body length might be c.5–6 mm (body parts are too incomplete to be worth measuring).

#### 3.2. Superfamily Proctotrupoidea Latreille, 1802

[By Victor A. Kolyada.]

Family Proctotrupidae Latreille, 1802

Tribe Cryptoserphini Kozlov, 1970

Genus *Oxyserphus* Masner, 1961

*Oxyserphus kozlovi* Kolyada, sp. nov.

Plate 1, fig. 1; Text-fig. 2

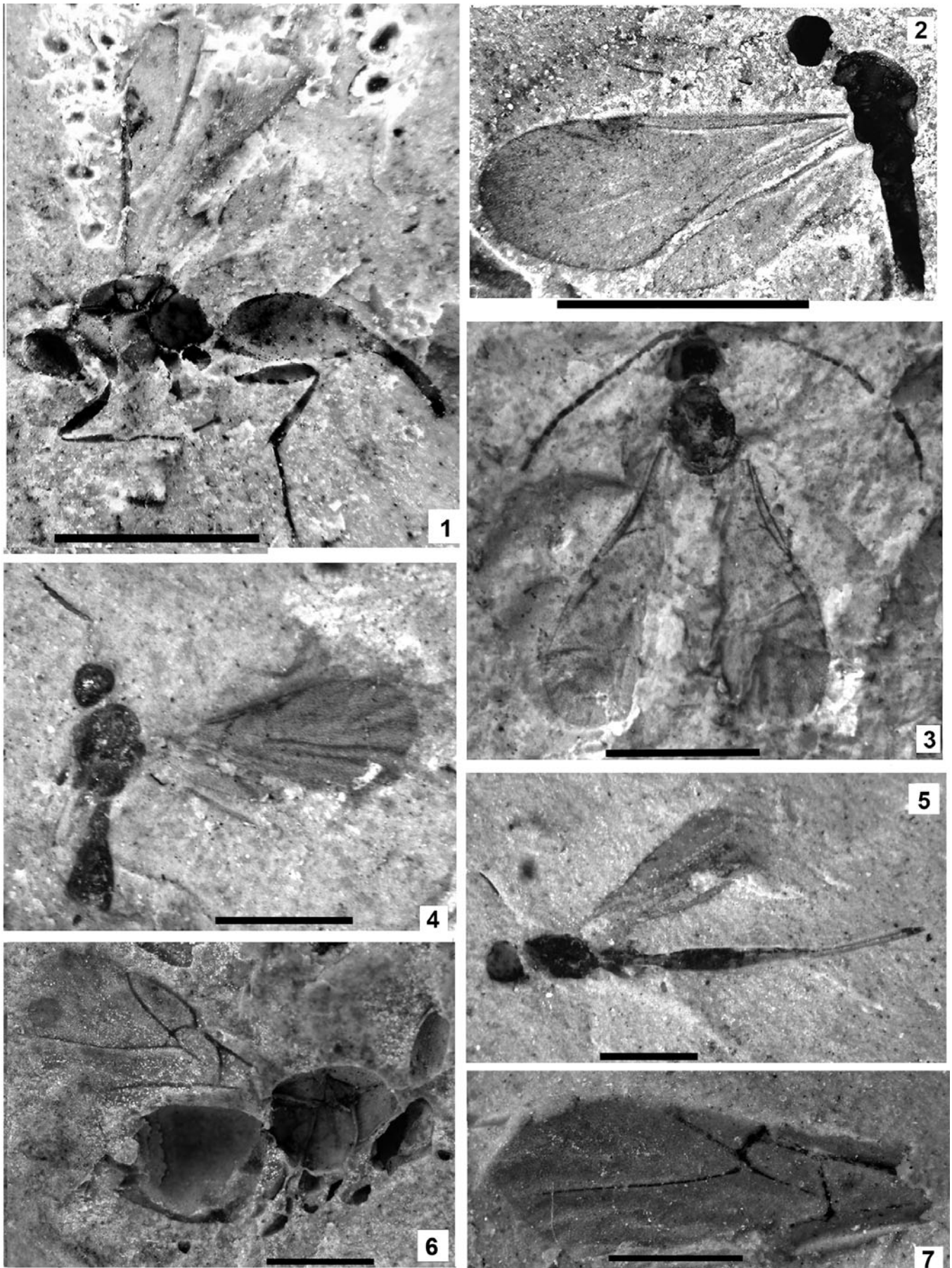
**Etymology.** In memory of Mikhail A. Kozlov.

**Holotype.** NHMUK I.9769, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; female, side view of insect lacking mid leg and most of antenna.

**Diagnosis.** The new species is similar to *O. capitatus* Townes, in Townes & Townes 1981 in ovipositor form which differs in details: the latter species has sheaths shorter (1.0 as long as hind femur), and additionally, unlike the new species, it has the ultimate female flagellomere distinctly incrassate. Incomplete preservation of the holotype makes the diagnosis tentative.

**Description.** Body slender, length 2.9 mm, forewing 1.35 mm. Head slightly transversal, 1.6 times as high as wide. Apical antennal segments of usual thickness, c. 2.5 (penultimate) – 3 times (ultimate) as long as wide, ultimate 1.2 times as long as penultimate. Mesosoma of usual form, not depressed. Horizontal groove across mesopleurum complete. Propodeum with strong reticulation. Forewing with pterostigma of usual form, 1.9 times as long as wide. Vertical part of pterostigma twice as long as wide, with radial vein originating from it. Radial vein slightly arched, joining costal vein at 40°. Costal section of radial cell 0.6 times as long as depth of pterostigma. Costal vein not extending beyond marginal cell. Hind tibia with spur not reaching midlength of basitarsus. Metasoma without a stalk. Ovipositor sheath somewhat widened toward widely rounded apex, 7.5 times as long as wide, c. 1.2 times as long as hind tibia.

**Remarks.** *Oxyserphus* is known to be widespread over Australia and New Zealand (Townes & Townes 1981); however, there are unpublished records from most of the Oriental region, including SE Asia up to Japan, and also for Central America. Apparently only one fifth of the world fauna is

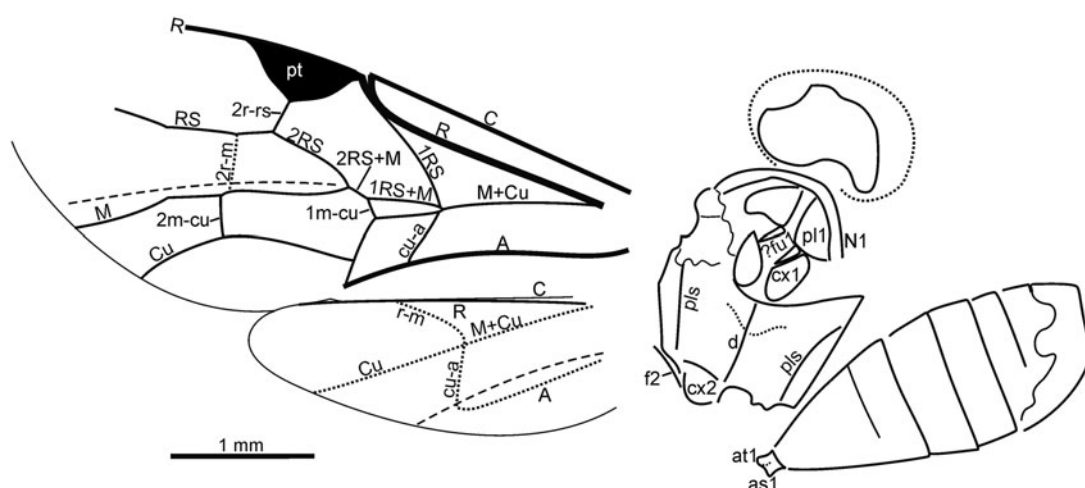


**Plate 1** (1) *Oxyserphus kozlovi* Kolyada, sp. nov., NHMUK I.9769, holotype. (2) *Mischoserphus* sp., specimen NHMUK I.9142. (3) *Zygota? filicornis* Cockerell, NHMUK I.9269, holotype. (4) *Miota? strigata* Cockerell, NHMUK I.9312, holotype. (5) *Macroteleia veterna* Cockerell, NHMUK In.17262, holotype. (6) *Andricus vectensis* Cockerell, NHMUK I.8923, holotype. (7) *Rhodites vetus* Cockerell, NHMUK In.24341, holotype. Scale bars = 1 mm.

(a)



(b)



**Text-figure 1** *Vectevania vetula* Cockerell, holotype NHMUK In.20535, Bembridge Marls, Isle of Wight, England: (A) photograph of the fossil. Ruler units equal to 1 mm; (B) line drawing. For wings, spectral veins are shown by dots, membrane folds by dashes; for body, dot lines indicate the very tentative head contour and the bend between lower and anterior faces of mesopectus. The vein nomenclature is standard. Other abbreviations: at1, as1 = acrotergite and acrosternite of the 1st metasomal segment; f2 = margin of mid femur; ?fu1 = supposedly parts of prothoracic furca; d = discrimen (line of invagination of the mesothoracic sternum); f2 = margin of mid femur; ?fu1 = supposedly parts of prothoracic furca; N1 = fore margin of pronotum; pls = pleural suture; cx1, cx2, pl1 = entrances into cavities of, respectively, fore and mid coxae and of propleura.

described so far. The genus is thermophilous, preferring humid and shadowed forests. Known to be larval endoparasite of Curculionidae and Anthribidae (Townes & Townes 1981).

Genus *Mischoserphus* Townes, in Townes & Townes 1981

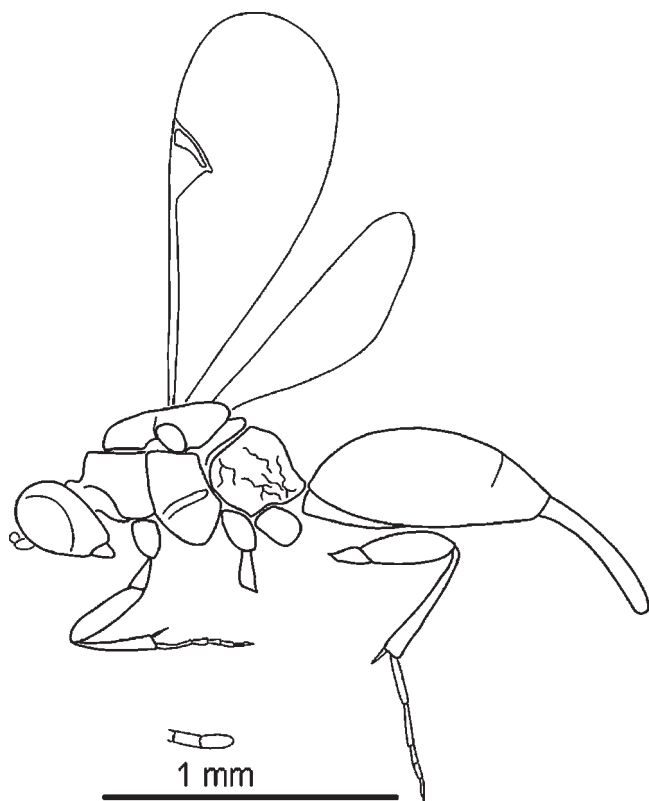
*Mischoserphus* sp.

Plate 1, fig. 2; Text-fig. 3

**Material.** NHMUK I.9142, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; partial body with no characters worth to study preserved, and complete wings.

**Description.** Forewing length 1.47 mm. Costal vein extending beyond apex marginal cell for 0.9 times of costal length of marginal cell.

**Remarks.** The generic position of the fossil is due to its costal vein extending beyond the apex of the marginal cell, which is characteristic of *Mischoserphus*, supported by the slender body and small, round head. Comparison with other congeners is impossible based on the characters available. *Mischoserphus* is a widespread genus parasitising fungus gnats (Mycetophilidae).



**Text-figure 2** *Oxyserphus kozlovi* Kolyada, sp. nov., NHMUK I.9769, holotype.

### 3.3. Superfamily Diaprioidea Haliday, 1833

Family Diapriidae Haliday, 1833  
Genus *Zygota* Foerster, 1856  
*Zygota? filicornis* Cockerell, 1921  
Plate 1, fig. 3

1921a *Zygota? filicornis* Cockerell, p. 23, fig. 29.

**Holotype.** NHMUK I.9269, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie.

**Remark.** The type was not revised. The genus is limited to the cool part of the Holarctic, ranging from tundra to mixed forests, occurring further southward on mountains only. Host unknown.

Genus *Miota* Foerster, 1856  
*Miota? strigata* Cockerell, 1921  
Plate 1, fig. 4

1921a *Miota? strigata* Cockerell, p. 22, fig. 28.

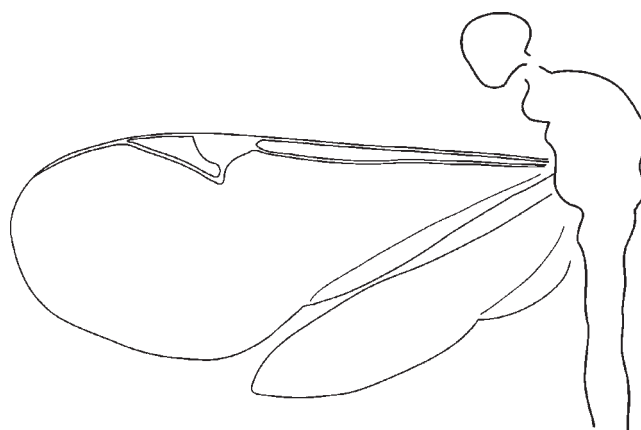
**Holotype.** NHMUK I.9312, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie.

**Remark.** The type was not revised. The genus is distributed worldwide, prefers temperate, mesic, forested environments and parasitises fungus gnats.

### 3.4. Superfamily Platygastroidea Haliday, 1833

Family Scelionidae Haliday, 1839  
Genus *Calotelea* Westwood, in Hope, 1837  
*Calotelea vetera* (Cockerell, 1921), comb. nov.  
Plate 1, fig. 5

1921a *Macroteleia vetera* Cockerell, p. 21, fig. 26.



**Text-figure 3** *Mischoserphus* sp., NHMUK I.9142.

**Holotype.** NHMUK In.17262, Bembridge Marls, NW Isle of Wight, UK, Smith Collection.

**Remark.** Generic attribution is by L. Masner (pers. comm., 2006) based on examination of the type. The genus populates mainly tropics and subtropics, the only host record is *Aeshna* dragonfly.

### 3.5. Superfamily Cynipoidea Latreille, 1802

[By Alexandr P. Rasnitsyn.]

Family Cynipidae Latreille, 1802  
Genus *Andricus* Hartig, 1840  
*Andricus vectensis* Cockerell, 1921  
Plate 1, fig. 6

1921a *Andricus vectensis* Cockerell, p. 23, fig. 30.

**Holotype.** NHMUK I.8923, Bembridge Marls, NW Isle of Wight, UK, Collection P. B. Brodie.

**Remark.** The type has not been revised, so its taxonomic position is not confirmed.

Genus *Rhodites* Hartig, 1840  
*Rhodites vetus* Cockerell, 1921  
Plate 1, fig. 7

1921a *Rhodites vetus* Cockerell, p. 24, fig. 31.

**Holotype.** NHMUK In.24341, Bembridge Marls, NW Isle of Wight, UK, Hooley Collection.

**Remark.** The type has not been revised, so its taxonomic position is not confirmed.

### 3.6. Superfamily Chalcidoidea Latreille, 1817; Family Agaonidae Walker, 1846

[By Stephen G. Compton.]

Donisthorpe (1920) described *Ponera minuta* as a new species of ant (Formicidae) based on a specimen housed in the NHMUK. Two further specimens are present in the collection of the Sedgwick Museum, Cambridge. They display characters that were unclear in the original specimen; in particular, a strongly reduced wing venation and the presence of an exerted ovipositor. Clearly they are not a species of ant. The forewing venation is typical of a chalcid (Hymenoptera, Chalcidoidea) type, but with a pigmented basal vein. Additional features are characteristic of adult female fig wasps that enter figs (the inflorescences of *Ficus* species, Moraceae) to oviposit. These features include a long flattened head, an enlarged fore femur with a reflexed short stout tibia, and the form of the ovipositor.

Fig wasps form an artificial assemblage of several unrelated chalcidoid lineages of gallers, parasitoids and inquiline (Rasplus *et al.* 1998). The best known group are the pollinating fig wasps (Agaonidae *sensu* Rasplus *et al.* 1998) that are partners in a highly co-evolved mutualism with their plant hosts. Figs are shaped like a hollow ball, lined on the inside by hundreds or thousands of tiny flowers. Adult female agaonids enter the figs through a narrow bract-lined ostiole, losing their wings and part of their antennae on entry. Once inside, they lay their eggs down the styles of the female flowers, while at the same time pollinating other female flowers (Weiblen 2002). The length of their ovipositor closely corresponds to the average length of these styles (Nefdt & Compton 1996).

Agaonidae are one of several lineages with females that can enter figs to oviposit (in addition there are fig wasps with much longer ovipositors that oviposit into the flowers from the outside of the fig). All such females have flattened heads and strong, spiny legs, an example of convergence in response to the difficulties associated with fig entry (van Noort & Compton 1996). Bouček (1997) proposed three synapomorphies for female Agaonidae: a unique ridged or toothed mandibular appendage, unusual antennae, and a head with channels to accommodate the scapes of the antennae during fig entry. Unfortunately, none of these features are clearly visible in the three available specimens. The antennae are absent and the tops of the heads are obscured. Flattened mandibles with several transverse ridges are nonetheless present, and are similar in appearance to those of some agaonids. Furthermore, what appears to be a ridged appendage, extending backwards below the head is also present. More conclusively, thoracic pollen pockets, unique to some Agaonidae, are present. The specimens can therefore be placed in Agaonidae, rather than one of the other groups of fig wasps, a conclusion supported by their non-thickened marginal vein, wing shape and the slightly curved exserted ovipositor.

Genus *Archaeagaon* Compton, gen. nov.

**Etymology.** After *archaios*, the Greek for ancient, and genus *Agaon*. Gender neuter.

**Type species.** *Ponera minuta* Donisthorpe, 1920.

**Diagnosis.** *Archaeagaon* can be distinguished from other genera of Agaonidae by the presence of a strongly pigmented basal vein.

*Archaeagaon minutum* (Donisthorpe) 1920, comb. nov.

Plate 2, figs 1–4

1920 *Ponera minuta* Donisthorpe, p. 85, plate 5, figure 4.

**Holotype.** NHMUK I.9734, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; adult female in side view.

**Other material.** CAMSM X.50140.47a (TN 98) and X.50140.97c (TN 152), Bembridge Marls, Isle of Wight, UK; adult females.

**Diagnosis.** As for genus.

**Description.** Length of body approximately 1.7 mm, mesosoma and metasoma approximately equal in length. Head elongate and flattened, about 2.5 times as long as high. Eyes situated towards rear of head, clearly shorter than the genae.

Thorax with pollen pockets (Compton *et al.* 2010). Fore femur enlarged, elongate and broadened, length about four times maximum width. Tibia short and broad, much shorter than the narrow tarsus. Forewing broad, less than twice as

long as wide, with relatively sparse, evenly-spaced setation, reaching proximally to (at least) the basal vein. Marginal, postmarginal and stigmal veins about equal in length, about half the length of the sub-marginal vein. Stigmal vein curved, with stigma not strongly demarcated. Basal vein pigmented, straight for most of its length, meeting the submarginal vein at a relatively acute angle. Ovipositor exserted, slightly down-curved, with sheaths about 0.33 mm in length. Antennae, mid and hind legs not visible in the three available specimens, hind wing venation also unclear. Male unknown.

**3.6.1. Discussion.** Fossil agaonids are well known in mid-Miocene amber from the Dominican Republic in the Caribbean, which dates from about 20 Ma, but they have not been recorded in amber from elsewhere. Three species were recently described from Dominican amber in the extant genera *Tetrapus* Mayr and *Pegoscapus* Cameron, reflecting their essentially modern appearance (Peñalver *et al.* 2006). Brues (1910) also described a supposed Tertiary agaonid, *Tetrapus mayri*, from the Florissant Formation in Colorado, USA. This specimen has not been examined recently, but on the basis of the original drawing and a photograph (Weiblen 2002) it is clearly not a species of *Tetrapus*, nor an agaonid. It may belong to Torymidae. *Archaeagaon minutum* is therefore the earliest confirmed species of fig wasp.

Based on molecular evidence, the association between fig trees and their pollinator fig wasps is believed to date back at least 60 million years (Machado *et al.* 2001; Rønsted *et al.* 2005), and so had been in place for tens of millions of years before the appearance of *Archaeagaon minutum*. The physical structure of this fig wasp reflects the anatomical adaptations for entry into figs that can be seen in modern species, with a similar size, with head, mandibles and legs that are adapted for entry via its tight bract-lined ostiole, and with an ovipositor of similar length. This suggests that the key anatomical features that underpin the fig tree–fig wasp relationship were fully in place during the Eocene, and perhaps well before; a finding consistent with the presence of fruits that are recognisably from figs in Eocene England (Collinson 1989). Modern species of fig trees display both monoecious and functionally dioecious breeding systems, with the former assumed to be ancestral.

Fig wasps are largely tropical or sub-tropical in distribution, though one species (*Blastophaga psenes* (L)) is present in Mediterranean Europe. The habitat associations of modern species are extremely varied, from rainforests to deserts, so their presence in the Isle of Wight deposits provides little information about likely conditions at the time, other than that they were warmer than today. Fig wasps are also exceptionally good dispersers, travelling distances of tens or even hundreds of kilometres between host plants (Ahmed *et al.* 2009), so their host plants could have been some distance away from where they were deposited. A fossil fig leaf has nonetheless been described from the Insect Bed, though its identification is considered doubtful (Compton *et al.* 2010).

### 3.7. Family ? Pteromalidae Dalman, 1820

[By Alexandr P. Rasnitsyn]

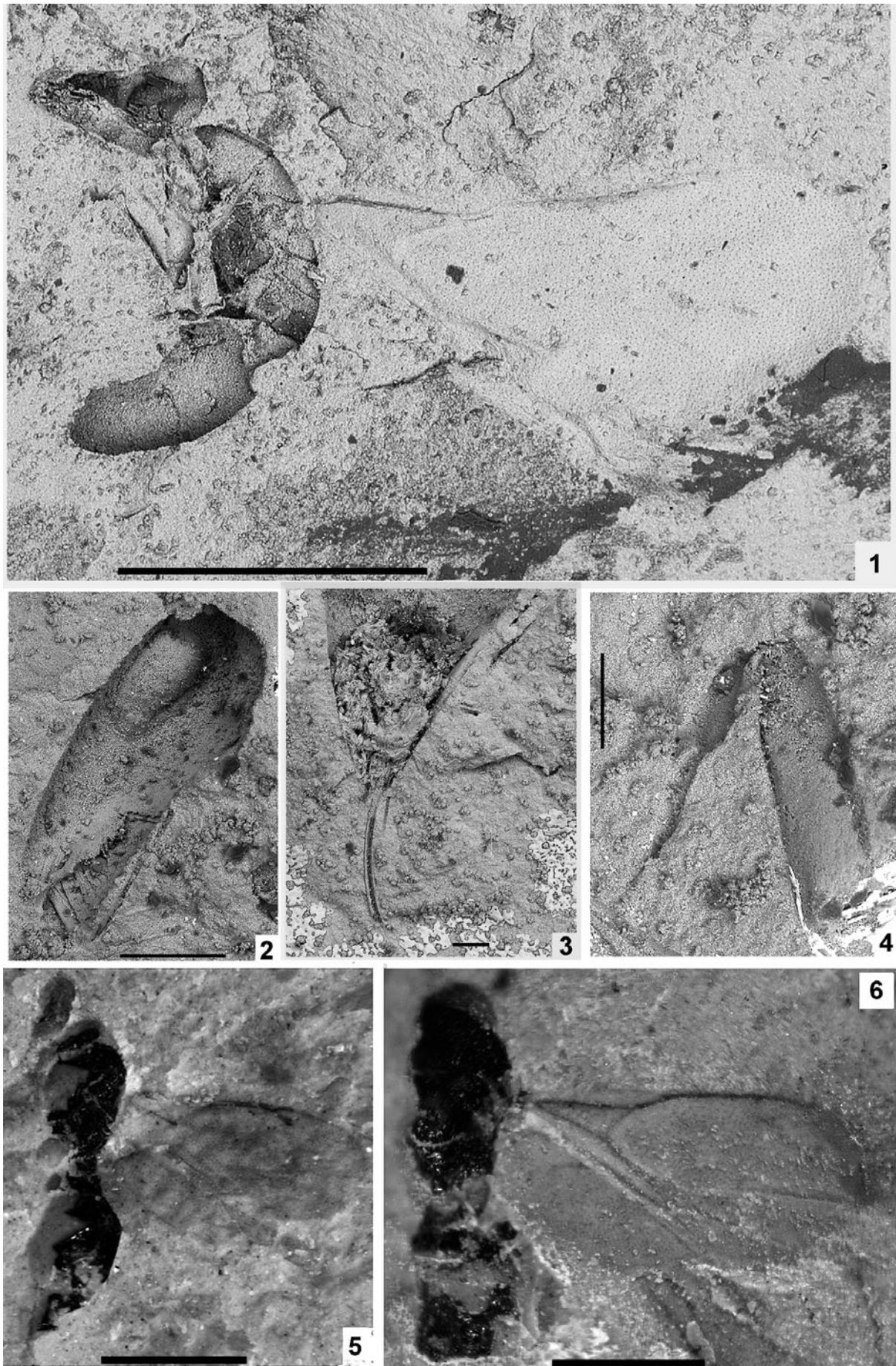
Genus ? *Pteromalus* Swederus, 1795

*Pteromalus? vectensis* Cockerell, 1921

Plate 2, fig. 5

1921a *Pteromalus? vectensis* Cockerell, p. 25, fig. 32.

**Holotype.** NHMUK I.9472, Bembridge Marls, NW Isle of Wight, UK, Collection P. B. Brodie.



**Plate 2** (1–4) *Archaeagaon minutum* (Donisthorpe): (1) NHMUK I.9734, holotype; (2) CAMSM X.50140.97c (TN 152), head; (3) CAMSM X.50140.47a (TN 98), metasomal apex with ovipositor; (4) CAMSM X.50140.97c (TN 152), fore leg. (5) *Pteromalus? vectensis* Cockerell, NHMUK I.9472, holotype. (6) *Lithobelyta reducta* Cockerell, NHMUK In.17091, holotype. Scale bars = 1 mm.

**Remark.** The fossil was not revised, its taxonomic position needs to be proven.

Family indet  
Genus *Lithobelyta* Cockerell, 1921

**Type species.** *L. reducta* Cockerell, 1921.

**Other species.** None.

**Remark.** Taxonomic position is obscure (originally described as Diapriidae, identified as Chalcididea by M. A. Kozlov, in litt., 2004).

*Lithobelyta reducta* Cockerell, 1921  
Plate 2, fig. 6

1921a *Lithobelyta reducta* Cockerell Cockerell, p. 22, fig. 27.

**Holotype.** NHMUK In.17091, Bembridge Marls, NW Isle of Wight, UK, Smith coll.

**Remark.** The type was not revised and requires re-study.

### 3.8. Superfamily Ichneumonoidea Latreille, 1802

[By Andrey I. Khalaim.]

**3.8.1. Family Ichneumonidae Latreille, 1802.** Ichneumonidae are one of the largest animal families, including over 30,000 described Recent species; although the real number of species is much greater. Most ichneumonids fly well and are common in all terrestrial biotopes from Arctic tundra to equatorial rainforests. Ichneumonidae are solitary or gregarious parasitoids, their larvae developing inside (endoparasitoids) or on (ectoparasitoids) living arthropod hosts. Most ichneumonids attack eggs, larvae or pupae of holometabolous insects, but some species oviposit on or in spiders and spider egg sacs. Hyperparasitoids (secondary parasites) are also known. They lay eggs into a primary parasite developing in the host.

The oldest representatives of Ichneumonidae are recorded from the uppermost Jurassic or Lower Cretaceous, and belong to the fossil subfamily Tanychorinae Rasnitsyn, 1980 (Zhang & Rasnitsyn 2003). In the Eocene and Oligocene, the family Ichneumonidae began to flourish, and fossil subfamilies Townesitinae and Pherhombinae, and Recent subfamilies and genera appeared.

Thirty-two specimens of the family Ichneumonidae from the collection of the Natural History Museum are studied. 11 species described by Cockerell (1921a) are revised. Taxonomic changes are shown in Table 1.

The Bembridge Marls ichneumonid fauna is represented by the subfamilies Cryptinae, Pimplinae, Townesitinae, Orthocentrinae, Metopiinae and Paxylommatainae, and three specimens of the genus *Lithapectis* Cockerell, which is undetermined to subfamily. Cryptinae are the largest Recent subfamily, which are richly represented in many Cenozoic deposits, and very abundant in Florissant (Brues 1910) and Baltic amber (Kasparyan 1994). The subfamily Cryptinae has a world-wide distribution and a broad range of hosts of various orders (Lepidoptera, Hymenoptera, Coleoptera, Diptera, etc.). All Bembridge Marls Cryptinae belong to the tribe Phygadeuontini Foerster.

Another numerous Bembridge Marls subfamily is Pimplinae. This subfamily has a world-wide distribution and a broad range of hosts. The genera *Exeristes* Foerster and *Scambus* Hartig parasitise hosts hidden within plant tissues (in leaves, galls, buds) or cocoons and use their long ovipositor to reach such hosts. Pimplinae are abundant in most Cenozoic deposits, but not recorded from Baltic amber.

Townesitinae is a fossil subfamily known only from the Bembridge Marls and Baltic amber. This subfamily is very

numerous in both deposits, and in Baltic amber it is second only to Cryptinae (Kasparyan 1994). Townesitinae of the Bembridge Marls and Baltic amber are very similar morphologically, with two out of three Bembridge Marls species belonging to the genus *Marjorietta* Kasparyan described from Baltic amber.

Orthocentrinae are world-wide in distribution and predominantly occur in rainforests and humid forests of the temperate zone. They are mostly small in size and parasitise fungus gnats (Diptera). Orthocentrinae were recorded from Florissant (Brues 1910), Baltic amber (Kasparyan & Humala 1995) and some other Cenozoic deposits. Three species from the genus *Eusterinx* Foerster are recorded in the Bembridge Marls. Species described from Baltic amber also belong to Recent genera (A. E. Humala pers. comm. 2000).

Other Bembridge Marls subfamilies are represented by single specimens only. Metopiinae parasitise various Lepidoptera, oviposit in the larval instar and emerge always from the pupa; distributed world-wide. Metopiinae are first known from the lowermost Eocene Oise amber (Kasparyan & Humala 1995), but are not common there.

Paxylommatainae are a small and morphologically distinct subfamily. Recent species are rare and only occur in the Holarctic region, and fossil species are known from Baltic and Rovno amber (Kasparyan 1988a, 2001; Tolkanitz *et al.* 2005) and the Bembridge Marls only. They are probably parasites of ants.

Ichneumonid specimens are represented most often by isolated forewings and fragments. Sometimes head, meso- and metasoma and parts of legs are present. The body in well-preserved specimens is usually not or weakly deformed and preserves its three-dimensional structure. Wings are complete or sometimes crumpled, so real sizes and measurements may not be equal to such in photos and figures. The sex of the overwhelming majority of specimens is unknown.

Taxonomy is accepted as in the catalogue TaxaPad (Yu *et al.* 2005). The following guides and monographs were used for identification: Kasparyan (1981), Townes (1969, 1970a, b, 1971) and Humala (2003). Morphological terminology predominantly follows Townes.

**3.8.2. Subfamily Pimplinae Wesmäl, 1845.** Large world-wide subfamily with an extremely wide range of hosts. Well represented in most of the Cenozoic deposits.

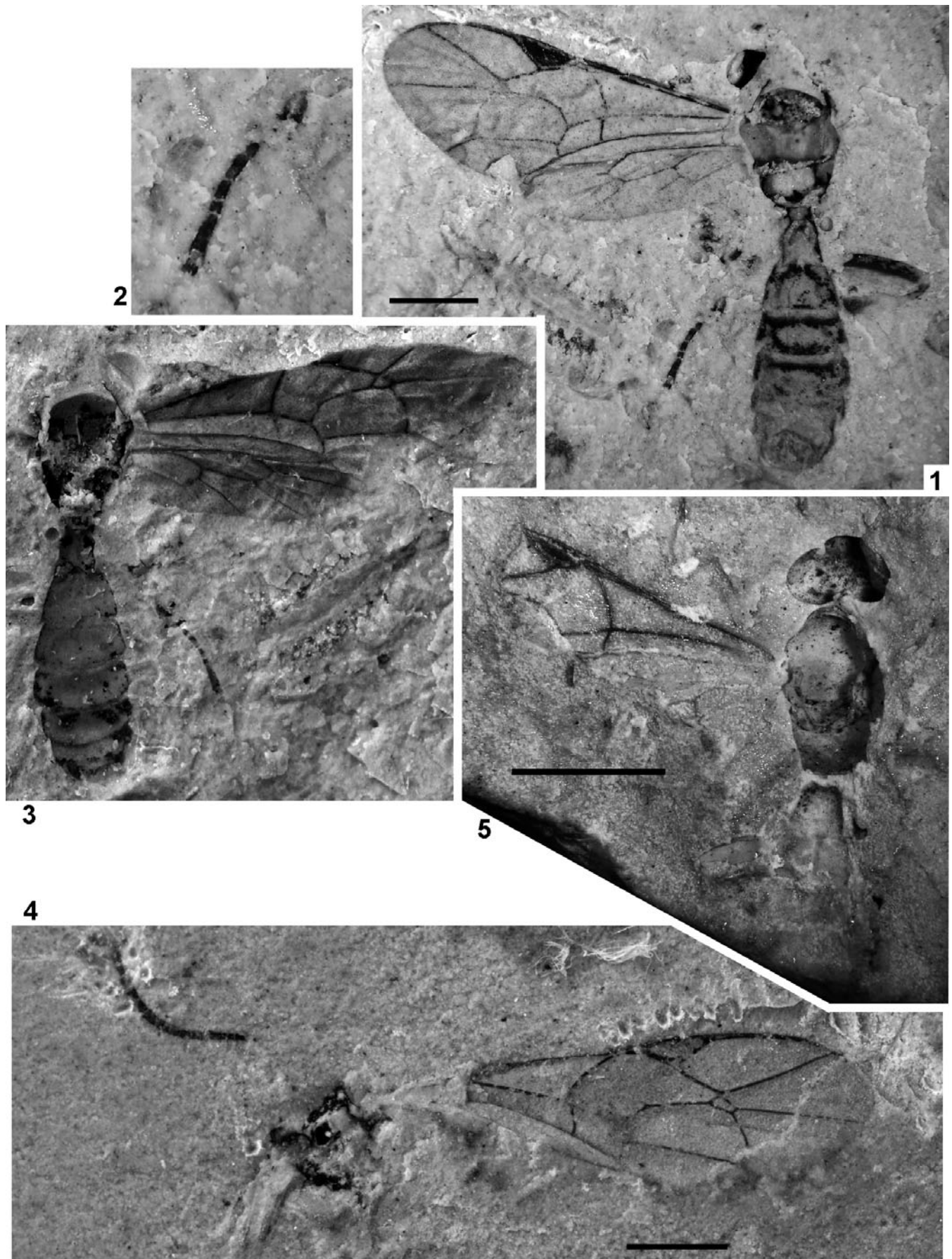
*Exeristes gurnetor* Khalaim, sp. nov.  
Plate 3, figs 1, 2, 3; Text-fig. 4

**Etymology.** After Gurnet Bay, old name of Gurnard Bay where some of the Bembridge insect fossils were collected

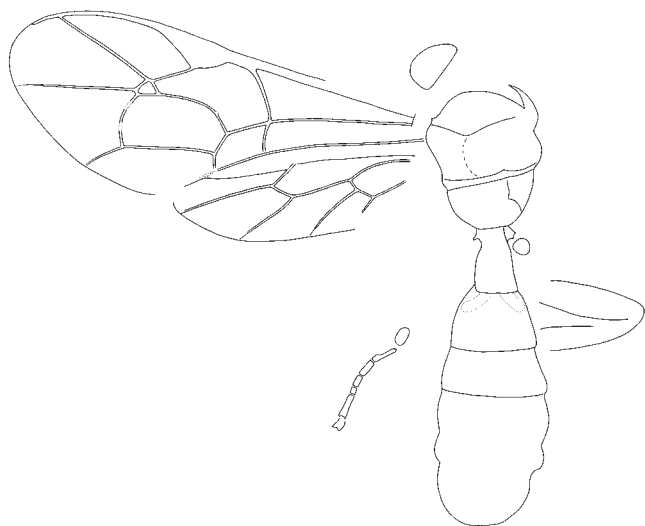
**Holotype.** IWCMS 2012.574, part and counterpart, Thorness Bay; dorsoventral aspect of body with incomplete head, left pair of wings and incomplete hind wings; sex unknown. Yule Coll.

**Diagnosis.** Diagnostic characters of the new species are the shorter discocubital and brachial cells, and first tergite longer than in Recent species. It also differs from all Pimplinae described from the Bembridge Marls in having forewing with the first intercubitus subequal to the second one, and the postnervulus intercepted low.

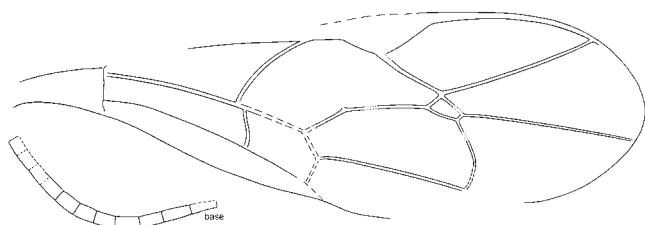
**Description.** Body length as measured from anterior head margin to apex of metasoma) 5 mm, forewing 4.3 mm, mesosoma 1.5 mm, mesosoma width 1 mm. Mesosoma short and stout; notaulus, sternaulus and postpectal carina absent, prepectal carina present. Pterostigma and veins dark-brown. Pterostigma wide, almost three times as long as wide, receiving radius near its middle. First section of radius curved near pterostigma. Second section of radius straight, almost 2.5



**Plate 3** (1–3) *Exeristes gurnetor* sp. nov., holotype, IWCMS 2012.574 (1, 2) and its counterpart (3). (4) *Itopectis saxosa* Cockerell, NHMUK In.24334, holotype. (5) Pimplinae genus and species indet. 2, NHMUK I.9965. Scale bars = 1 mm.



**Text-figure 4** *Exeristes gurnetor* sp. nov., IWCMS 2012.574, holotype.



**Text-figure 5** *Itopectis saxosa* Cockerell, NHMUK In.24334, holotype.

times as long as first section. Metacarp not reaching apex of forewing. Areolet subtriangular, wider than high, receiving second recurrent vein before its outer corner. Second recurrent vein evenly and roundly curved, with two bullae. Discocubitus moderately curved, with ramulus in its 0.36. Second discoidal cell comparatively short. Nervulus hardly postfurcal, slightly inclivous. Postnervulus intercepted below middle; posterior section slightly inclivous. Brachial cell short. Nervellus intercepted in its 0.6. Mediella weakly curved. Hind femur and tibia moderately thick. Hind tibia darkened basally (or sub-basally) and apically. 5th segment of hind tarsus 1.34 times as long as 2nd segment. Length ratio of 2–5 tarsal segments 12:8:5:16. Metasoma dark, strongly depressed. First tergite distinctly elongate, with pair of dorsal keels extending for about half length of tergite. Following tergites strongly transverse. 2nd tergite with pair of oblique basal grooves, 1.3 times as wide as long.

**Remarks.** Short and transverse second and following tergites, long fifth segment of hind tarsus and the intercepted high nervellus indicate that this species belongs to the genus *Exeristes* Foerster.

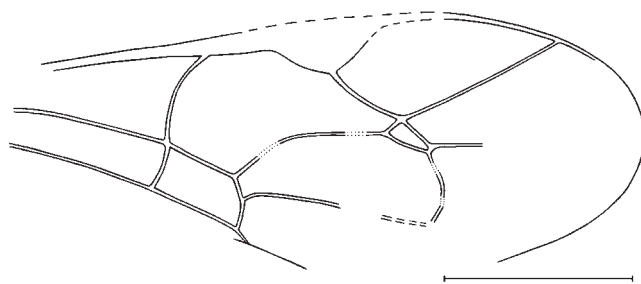
Pimplinae indet.

*"Itopectis" saxosa* Cockerell, 1921

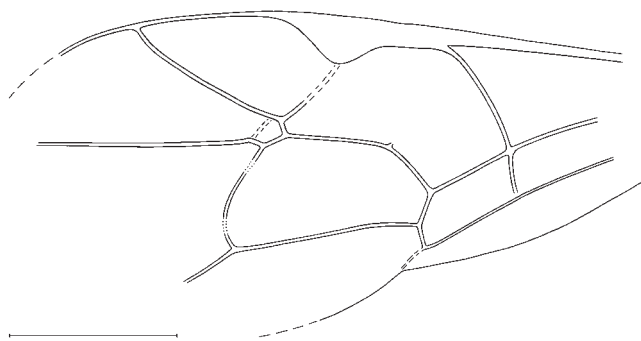
Plate 3, fig. 4; Text-fig. 5

1921a *Itopectis saxosus* Cockerell, 7, fig. 5 (Pimplinae).

**Holotype.** NHMUK In.24334, Bembridge Marls, NW Isle of Wight, UK, Hooley Collection; forewing, part of antenna and mesosoma fragment; sex unknown.



**Text-figure 6** *Scambus fossilis* sp. nov., NHMUK In.64644, holotype. Scale bar = 1 mm.



**Text-figure 7** Pimplinae gen. et sp. indet., NHMUK In.24417. Scale bar = 1 mm.

**Diagnosis.** Differs from other Pimplinae in the Bembridge Marls in having forewing with nervulus postfurcal, distinctly inclivous, and discocubitus very weakly curved.

**Description.** Forewing length 5.1 mm. Basal flagellar segments about twice as long as wide, mid flagellar segments elongate. Pterostigma dark-brown with small yellowish mark, veins brown to dark-brown. Pterostigma wide, 2.9 times as long as wide, receiving radius near its middle. Radius very slightly curved. Second section of radius 2.2 times as long as first section. Areolet pointed, wider than high, receiving second recurrent vein before its outer corner. Second recurrent vein roundly curved, with two bullae. Discocubitus slightly curved, with ramulus. Nervulus curved, distinctly postfurcal, inclivous. Postnervulus intercepted near its middle; posterior section slightly inclivous.

**Remarks.** The species probably belongs to Pimplinae, but cannot be identified closer because many genera of this subfamily (as well as some genera of other subfamilies, in particular of Orthocentrinae) have a similar wing venation.

*"Scambus" fossilis* Khalaim, sp. nov.

Plate 3, fig. 5; Text-fig. 6

**Etymology.** After *fossilis*, the Latin for fossil.

**Holotype.** NHMUK In.64644, Bembridge Marls, Isle of Wight, UK, Coll. Jarzembowski; forewing and dorsal aspect of poorly preserved head, mesosoma and base of metasoma; sex unknown.

**Diagnosis.** Differs from other Pimplinae from the Bembridge Marls in having the forewing with nervulus interstitial, subvertical, and areolet large, strongly oblique.

**Description.** Length of forewing 3.8 mm, mesosoma 1.7 mm. Pterostigma and veins dark brown. Pterostigma wide, receiving radius near its middle. First section of radius evenly curved. Second section of radius straight, 2.15 times as long as first section. Areolet large, subtriangular, receiving second recurrent vein a little before its outer corner. Second recurrent vein

roundly curved, with two bullae. Discocubitus moderately curved, probably without ramulus. Nervulus inclivous, vertical. Postnervulus intercepted high; posterior section inclivous. First tergite with distinct dorsal keels basally.

**Remarks.** This is an evident Pimplinae. *Scambus* is a large and rather polymorphic Recent genus, which corresponds morphologically with this specimen. However, it is possible that the real position of the fossil within the subfamily is somewhat different.

Genus and species indet. 1

Text-fig. 7

**Material.** NHMUK In.24417(1), Bembridge Marls, NW Isle of Wight, UK, Hooley Collection, incomplete forewing.

**Description.** Forewing length as preserved about 4.0 mm, full length over 5.0 mm. Pterostigma and veins brown. Pterostigma wide. Areolet present; second intercubitus unclear. Second recurrent vein roundly curved, with two bullae. Discocubitus slightly curved, with ramulus. Nervulus not curved, interstitial, slightly inclivous. Postnervulus intercepted somewhat below its middle; posterior section slightly inclivous.

Genus and species indet. 2

Plate 3, fig. 5

**Material.** NHMUK I.9965, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; ventral aspect of head, mesosoma and base of metasoma, with base of forewing and leg fragments, sex unknown.

**Description.** Mesosoma length 1.2 mm. Notaulus absent. Propodeum without carinae. Pterostigma and veins dark-brown. Pterostigma wide, receiving radius somewhat basal of its middle. First section of radius curved near pterostigma. Nervulus interstitial, slightly inclivous. Metasoma depressed, first tergite short, about as long as wide, spherically and evenly prominent dorsally, without conspicuous dorsal keels, probably densely and coarsely punctuate, following tergites distinctly transverse.

**3.8.3. Subfamily Orthocentrinae Foerster, 1869.** Large, world-wide subfamily that includes small and moderate-sized ichneumonids, predominantly occurring in humid forests. Orthocentrinae are parasitoids of primitive Diptera, mainly of the superfamily Sciaroidea. Orthocentrinae are also found in Baltic amber (Kasparyan & Humala 1995), Florissant (Brues 1910), Green River (Cockerell 1941) and other Cenozoic deposits. Three species from the Bembridge Marls probably belong to the large and rather polymorphic genus *Eusterinx* Foerster, 1869. *Holomeristus* Foerster, 1869 is considered a subgenus of *Eusterinx* by Humala (2003), and mentioned as a synonym in catalogues of Townes (1971) and Yu *et al.* (2005). Fossil species of this genus are known from the Bembridge Marls only.

*Eusterinx vectensis* (Cockerell, 1921), comb. nov.

Plate 4, fig. 1; Text-fig. 8

1921a *Holomeristus* (?) *vectensis* Cockerell, p. 8, fig. 6 (Orthocentrinae).

**Holotype.** NHMUK I.9370, Bembridge Marls, NW Isle of Wight, UK, Brodie collection; lateral aspect of head with partial antennae, mesosoma, forewing and leg fragments; sex unknown.

**Diagnosis.** Differs from *E. arcuatus* in having the forewing with areolet large, first intercubitus long, and discocubitus moderately curved. Differs from *E. humalai* in having the forewing with areolet.

**Description.** Head and mesosoma combined 1.9 mm long. Forewing length 3.4 mm. Occipital carina present. Scape of antenna suboval. Basal flagellar segments thin and long, proportions of 1–5 segments 19:10:8:8:7.5. Ocelli of moderate size; distance between lateral and frontall ocellus subequal to diameter of ocellus. Clypeus separate from face. Notaulus deep. Postpectal carina probably absent. Hind coxa and femur slender. Pterostigma wide, receiving radius near its middle. First section of radius distinctly curved. Second section of radius almost straight, 2.3 times longer than first section. Areolet large, wider than high. Second recurrent vein inclivous, straight. Discocubitus strongly curved, probably without ramulus. Nervulus interstitial, very slightly inclivous. Postnervulus intercepted in its 0.7; posterior section inclivous.

*Eusterinx arcuatus* (Cockerell, 1921), comb. nov.

Plate 4, fig. 2; Text-fig. 9

1921a *Cremastus* (?) *arcuatus* Cockerell, p. 8, fig. 7 (Cremastinae).

**Holotype.** NHMUK I.9374/I.9246, part and counterpart, Bembridge Marls, NW Isle of Wight, UK, Brodie collection, 1898; lateral aspect of body with scape, forewing and hind leg fragments (counterpart poorly preserved), sex unknown.

**Diagnosis.** Differs from *E. vectensis* in having forewing with small, oblique areolet, very short first intercubitus, and strongly curved discocubitus. Differs from *E. humalai* in having the forewing with areolet.

**Description.** Body length 4.1 mm, forewing 3.5 mm, mesosoma 1.18 mm, mesosoma height 0.86 mm, first tergite length 0.46 mm. Clypeus separate from face. Notaulus deep and short. Prepectal carina and mesopleural fovea present. Postpectal carina present only crossing mesosternal furrow. Propodeum areolate. Pterostigma and veins brown to dark-brown. Pterostigma wide, almost three times as long as wide, receiving radius near its middle. First section of radius curved. Second section of radius slightly curved, 2.2 times as long as first section. Areolet narrow, oblique, first intercubitus very short. Second recurrent vein curved, slightly inclivous, with two bullae. Discocubitus curved, without ramulus. Nervulus interstitial, vertical. Postnervulus intercepted in its 0.6; posterior section very slightly inclivous. First tergite slightly and evenly curved, subcylindrical anteriorly and somewhat depressed posteriorly, with spiracle near its 0.6; dorsal keels weak, but distinct, reaching about 0.7 tergite length; dorso-lateral keel about 0.4 of tergite length, not reaching spiracle; ventrolateral keel distinct and complete. Glymma absent.

**Remark.** The counterpart of the holotype (NHMUK I.9246) was incorrectly identified as an ant (*Leucotaphus gurnetensis*) by Donisthorpe (1920).

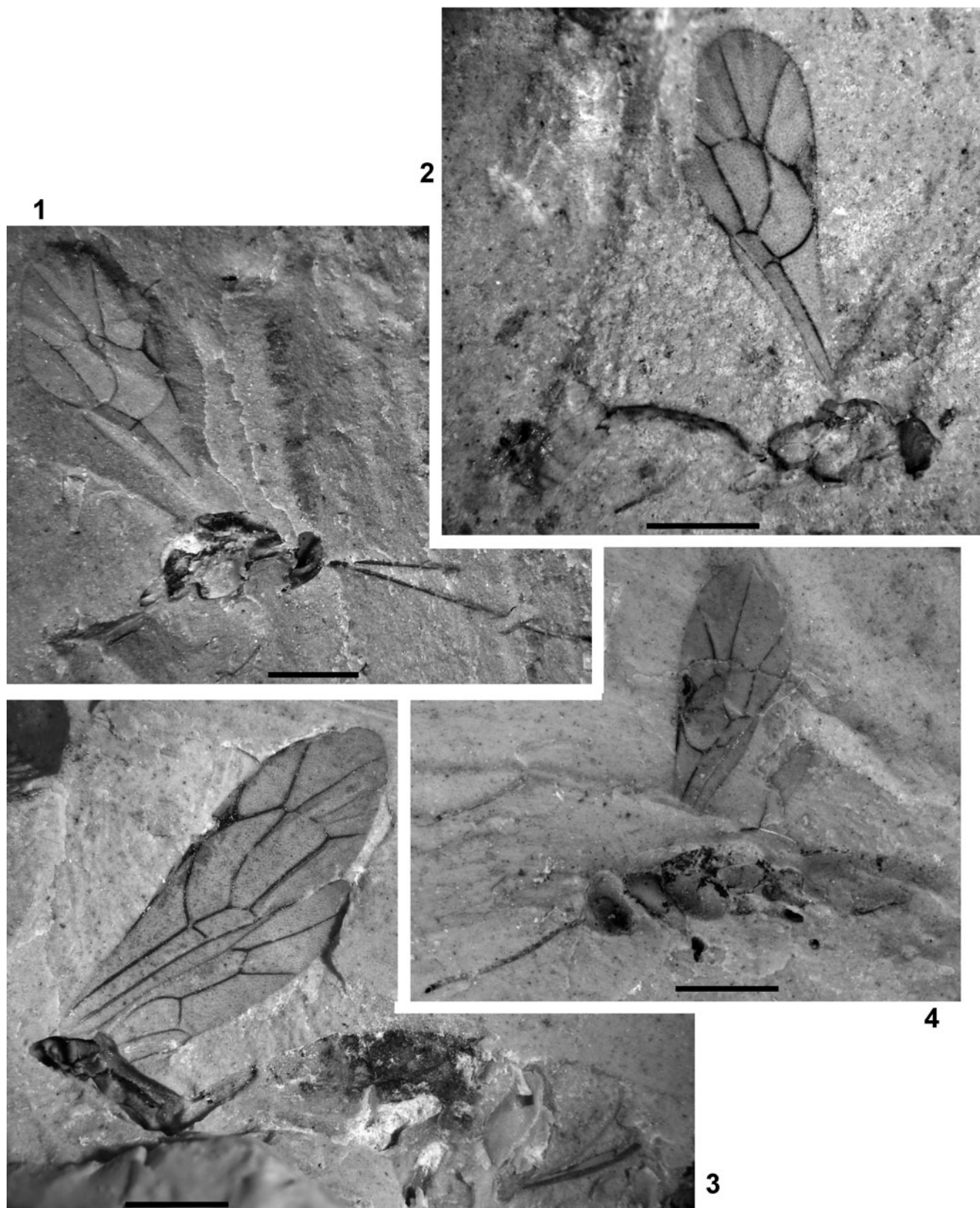
*Eusterinx humalai* Khalaim, sp. nov.

Plate 4, fig. 3; Text-fig. 10

**Etymology.** In honour of Dr. Andrey E. Humala, expert in Orthocentrinae.

**Holotype.** NHMUK In.25730, Bembridge Marls, NW Isle of Wight, UK, Hooley collection; dorso-lateral aspect of female meso- and metasoma with wings.

**Diagnosis.** Differs from other congeners in the Bembridge Marls in having the forewing without areolet and with the first section of radius straight, discocubitus very slightly curved, nervulus inclivous, and postnervulus intercepted low. It differs from Recent *Eusterinx* lacking areolet in having the second discoidal cell unusually long.



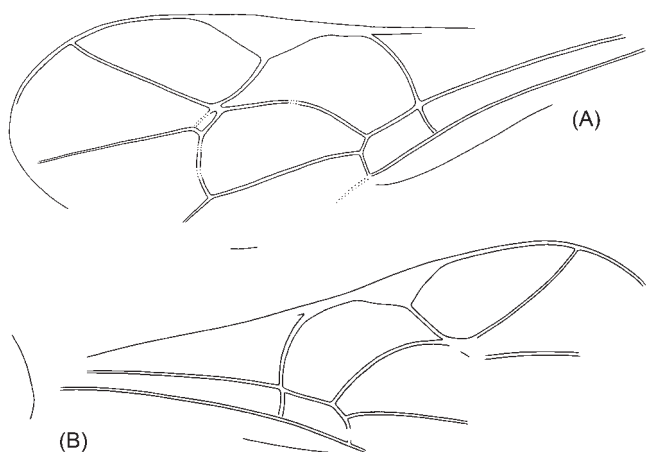
**Plate 4** (1) *Eusterinx vectensis* (Cockerell), NHMUK I.9370, holotype. (2) *Eusterinx arcuatus* (Cockerell), NHMUK I.9374, holotype. (3) *Eusterinx humalai* sp. nov., NHMUK In.25730, holotype. (4) *Hypsicera anglica* (Cockerell), NHMUK I.9292, holotype. Scale bars = 1 mm.

**Description.** Body length over 6 mm, forewing 4.4 mm, first tergite *c.* 1 mm; sheath length 1.6 mm. Notaulus deep, long, reaching centre of mesonotum. Prepectal carina present. Sternaulus deep, reaching from anterior margin to midlength of mesopleuron. Postpectal carina absent. Mesopleural fovea

present. Propodeum areolate. Pterostigma and veins brown to dark-brown. Pterostigma moderately wide, about 3.5 times as long as wide, receiving radius near its middle. First section of radius straight. Second section of radius almost straight, 1.55 times as long as first section. Areolet absent. Second recurrent



**Text-figure 8** *Eusterinx vectensis* (Cockerell), NHMUK I.9370, holotype.



**Text-figure 9** *Eusterinx arcuatus* (Cockerell): (A) NHMUK I.9374, holotype; (B) NHMUK I.9246, counterpart.

vein curved, slightly inclivous, with two bullae. Discocubitus very weakly and evenly curved, without ramulus. Second discoidal cell elongate, 2.3 times as long as wide. Nervulus interstitial, inclivous. Postnervulus intercepted in its 0.4; posterior section slightly inclivous. Nervellus not intercepted. Mediella distinctly curved. Metasoma behind first segment subcylindrical or slightly compressed. First tergite weakly and evenly widened toward apex, with weak ventro-lateral carina, with spiracle probably before middle. Ovipositor thin, slightly upcurved.

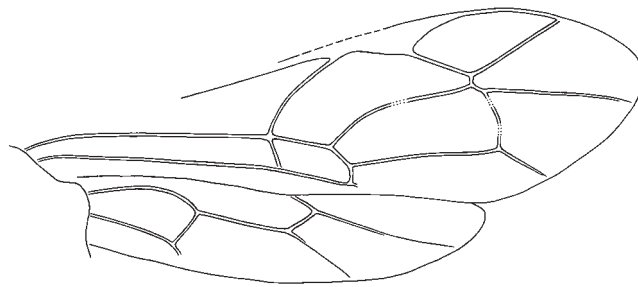
**3.8.4. Subfamily Metopiinae Foerster, 1869.** Moderate-sized, Recent subfamily, parasites of Lepidoptera. Fossil Metopiinae are registered in some Cenozoic deposits, such as the lowermost Eocene Oise amber (Menier *et al.* 2004), the uppermost Eocene of Florissant (Brues 1910), the uppermost Eocene or lowermost Oligocene of Biamo in the Russian Far East (Khalaïm 2008), and Miocene Brick Yard in NW USA (Lewis *et al.* 1990).

*Hypsicera anglica* (Cockerell, 1921), comb. nov.  
Plate 4, fig. 4; Text-fig. 11

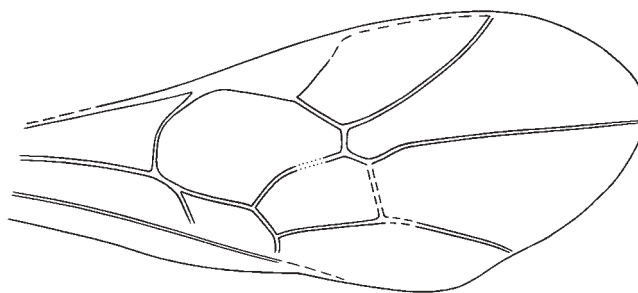
1921a *Polyclistus* (?) *anglicus* Cockerell, p. 10, fig. 10 (Metopiinae).

**Holotype.** NHMUK I.9292, Bembridge Marls, NW Isle of Wight, UK, Brodie collection; lateral aspect of body with metasoma incomplete, antenna, almost complete forewing, and leg fragments; sex unknown.

**Diagnosis.** Differs from recent species by the combination of the strongly inclivous, postfurcal nervulus, the broad pterostigma, and the subvertical, almost straight second recurrent vein.



**Text-figure 10** *Eusterinx humalai* sp. nov., NHMUK In.25730, holotype.



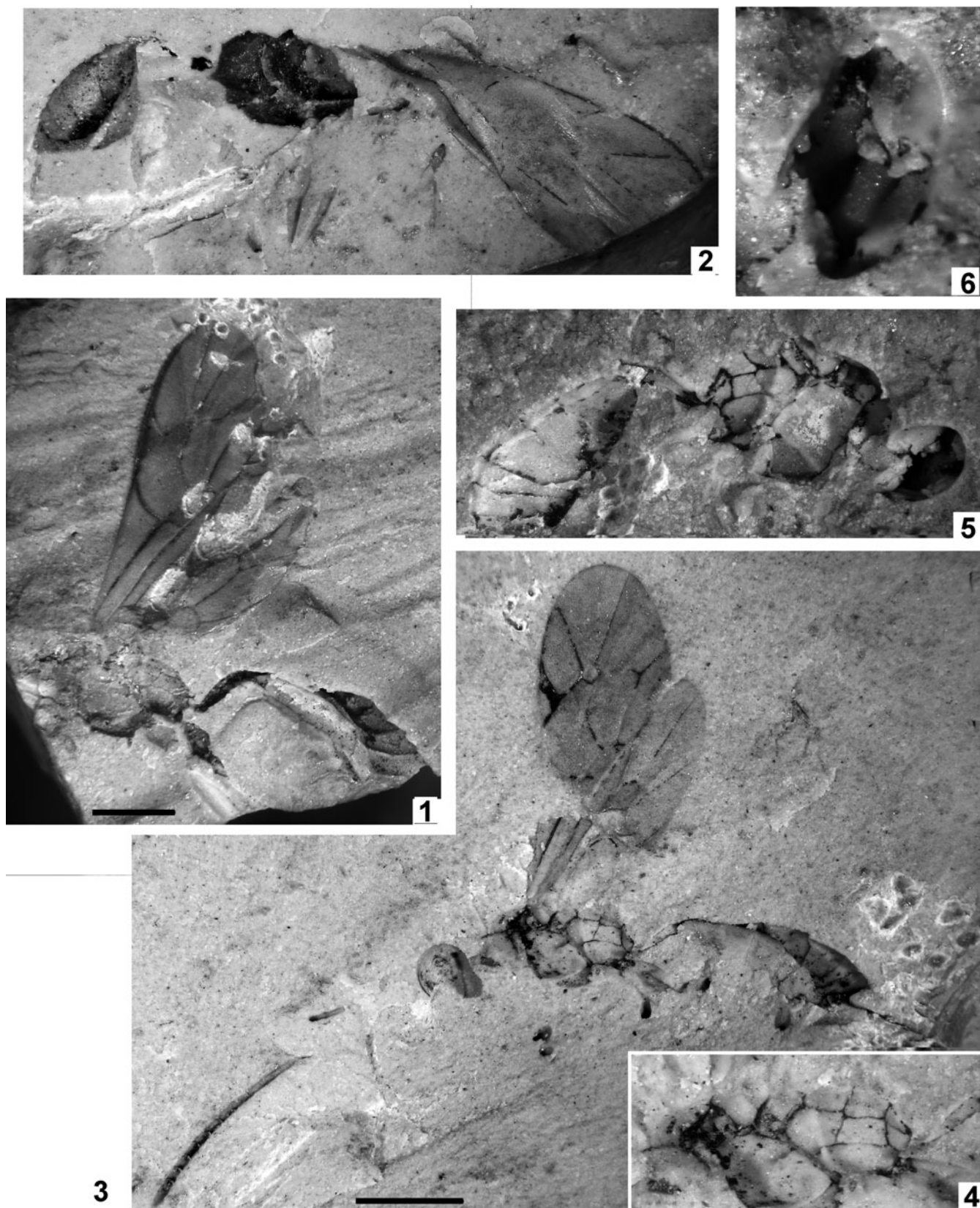
**Text-figure 11** *Hypsicera anglica* (Cockerell), NHMUK I.9292, holotype.

tigma, and the subvertical, almost straight second recurrent vein.

**Description.** Body length *c.* 3.6 mm, forewing 2.9 mm, mesosoma 1.4 mm, height 0.75 mm. Head widened downward, rather abruptly sloping vertically just behind rear margin of hind ocelli. Ocelli of moderate size. Occipital carina absent dorsally and laterally. Scape of antenna short, oval. Flagellomeres at least 18 in number, visible slightly elongate and of subequal width. Mesosoma. Pronotum distinctly swollen, strongly and widely impressed. Notaulus probably absent. Prepectal carina present, indistinct in upper part of mesopleuron. Sternaulus and postpectal carina absent. Mesopleural fovea present. Propodeum areolate. Pterostigma and veins dark-brown. Pterostigma wide, over 2.5 times as long as wide, receiving radius in the middle. First section of radius straight, second section gradually curved. Areolet absent. Second recurrent vein not curved, slightly inclivous. Discocubitus slightly curved, without ramulus. Nervulus distinctly postfurcal and strongly inclivous. Postnervulus intercepted in its lower 0.38. Hind coxa and femur very thick. Metasoma cylindrical or depressed.

**Remarks.** This species belongs to Metopiinae as is justified by a robust, cylindrical body, globose head, thick legs, and characteristic venation of forewing (Text-fig. 11). It belongs most likely to the genus *Hypsicera* Latreille, 1829 which is characterised by the head abruptly sloping vertically just behind rear margin of hind ocelli. *Hypsicera* is a large, world-wide genus comprising over 60 Recent species, most common in tropics, and parasitising various Lepidoptera.

**3.8.5. Subfamily Cryptinae Kirby, 1837.** A large and very diverse subfamily of worldwide distribution. Cryptinae are recorded from any Cenozoic deposits with minimally diverse Ichneumonidae, and always most numerous there. All fossil Cryptinae belong to the tribe Phygadeuontini Foerster, 1869 because of having the second recurrent vein inclivous and propodeum completely areolate. Generic identification of Phygadeuontini is very difficult, and characters available in fossils are usually insufficient for reliable generic identification. I



**Plate 5** (1) "*Hemiteles*" *acourti* (Cockerell), NHMUK In.24332, holotype. (2) "*Hemiteles*" *dirus* sp. nov., NHMUK I.9244, holotype. (3–6) "*Hemiteles*" *protervus* sp. nov.: (1–2) NHMUK I.9609, holotype; (3–4) NHMUK In.25765, paratype; (5–6) NHMUK In.25765, paratype. Scale bars = 1 mm.

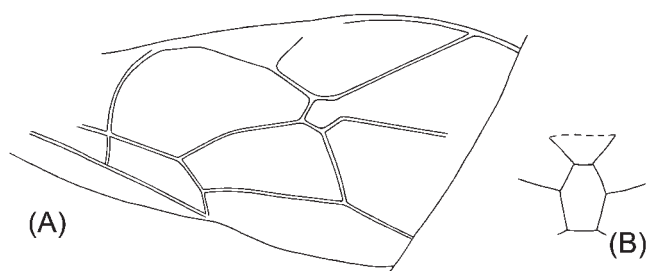
retain Cockerell's original attribution of the fossil using the quotation marks for the generic name to note the very preliminary character of the attribution. New species are described as "*Hemiteles*" because this attribution looks likely but cannot be well grounded at present.

"*Hemiteles*" *acourti* (Cockerell, 1921), comb. nov.  
Plate 5, fig. 1; Text-fig. 12

1921a *Ichneumon* (s. latiss.) *acourti* Cockerell, 5, fig. 2  
(Ichneumoninae).



**Text-figure 12** *Hemiteles acourti* (Cockerell), NHMUK In.24332, holotype.

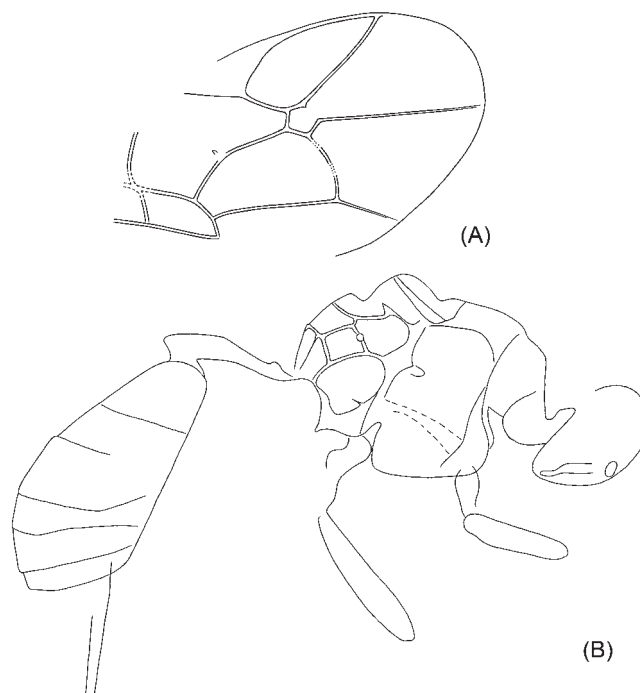


**Text-figure 13** *Hemiteles dirus* sp. nov., NHMUK I.9244, holotype: (A) = forewing; (B) = areola of propodeum.

**Holotype.** NHMUK In.24332, Bembridge Marls, NW Isle of Wight, UK, Hooley collection; lateral view of mesosoma and metasoma (lacking apex) with wings and leg bases, sex unknown.

**Diagnosis.** This species differs from “*Hemiteles*” *protervus* in having the forewing with discocubitus almost straight anteriorly and posteriorly, and strongly curved near the middle, nervulus very slightly inclivous and postfurcal, and radial and second discoidal cells longer, and from “*Hemiteles*” *dirus* and from “*Stilpnus*” *oligocenus* in having the second section of postnervulus weakly inclivous.

**Description.** Size medium, mesosoma length 1.2 mm, forewing 4.4 mm, first metasomal tergite *c.* 1 mm. Prepectal carina present. Sternaulus deep, spanning along whole mesopleuron length. Mesopleural fovea present. Pterostigma and veins brown. Pterostigma wide, 2.6 times as long as wide, receiving radius in the middle. Radius very slightly curved. Radial cell 2.5 times as long as wide. Metacarp not reaching apex of forewing. Areolet pentagonal, second intercubitus unpigmented. Second recurrent vein inclivous, with two bullae. Discocubitus curved, without ramulus. Second discoidal cell 2.15 times as long as wide. Nervulus postfurcal, slightly inclivous. Postnervulus intercepted at somewhat below its middle, posterior section distinctly inclivous. Hind wing with nervellus intercepted below middle. Hind legs of moderate thickness. Metasoma depressed behind first segment. First tergite long, petiolate, with



**Text-figure 14** *Hemiteles protervus* sp. nov.: (A) NHMUK I.9609, holotype; (B) NHMUK In.25765, paratype.

ventro-lateral carinae and probably with dorso-lateral carinae. Glymma probably absent. Posterior tergites strongly transverse.

**Remarks.** Strong and long sternaulus which is never present in Ichneumoninae confirms attribution of the species to Cryptinae.

“*Hemiteles*” *dirus* Khalaim, sp. nov.

Plate 5, fig. 2; Text-fig. 13

**Etymology.** After *dirus*, the Latin for terrible.

**Holotype.** NHMUK I.9244, Bembridge Marls, NW Isle of Wight, UK, Brodie collection, posteroventral aspect of posterior part of mesosoma, side view of metasoma, forewing, and detached leg parts. Sex unknown.

**Diagnosis.** This species differs from “*Hemiteles*” *protervus* and “*Hemiteles*” *acourti* in having forewing with second recurrent vein strongly inclivous and almost straight, nervulus interstitial, and postnervulus strongly inclivous; and from *Stilpnus oligocenus* in having the postnervulus with posterior section inclivous, discocubitus distinctly curved, and size larger.

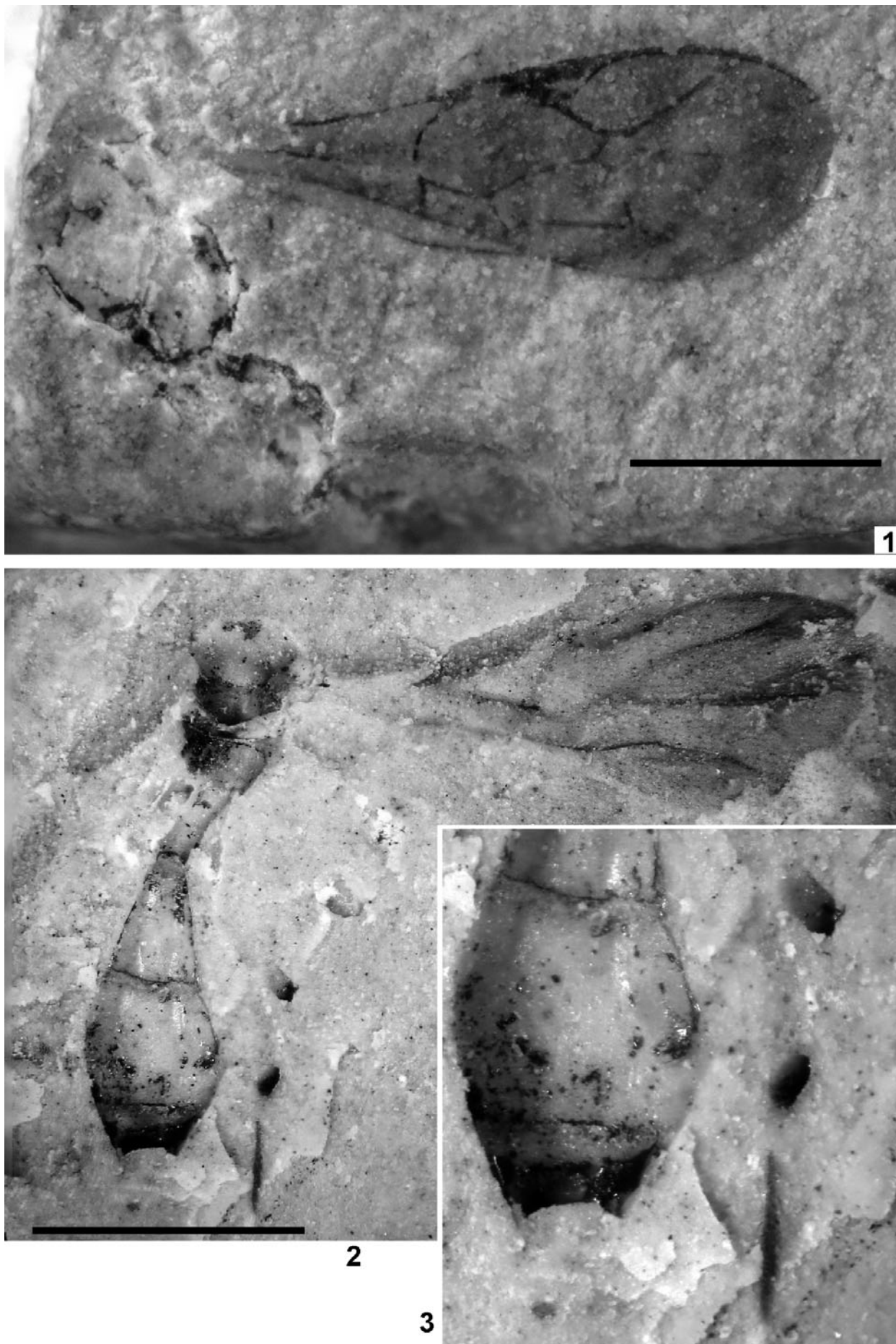
**Description.** Forewing length 3.8 mm. Notaulus present. Mesopleural fovea present. Propodeum areolate; areola hexagonal, elongate. Pterostigma wide, receiving radius near its middle. Radius slightly curved. Areolet pentagonal. Second recurrent vein strongly inclivous, almost straight, not curved near subdiscoideus, bullae indistinct. Discocubitus slightly and evenly curved, without ramulus. Nervulus interstitial, slightly inclivous. Postnervulus intercepted approximately in its 0.35 (posterior part of wing deformed), strongly inclivous, angle between posterior section and brachius 50°. Legs slender. Metasoma behind first segment depressed. Mid and posterior tergites strongly transverse.

“*Hemiteles*” *protervus* Khalaim, sp. nov.

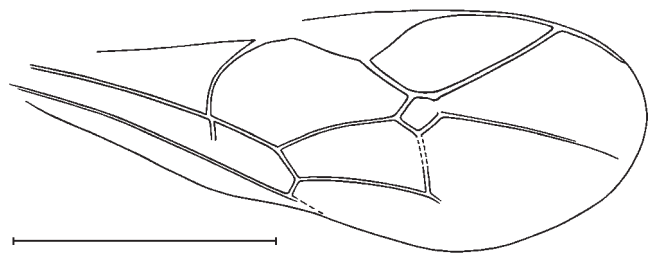
Plate 5, figs 3–6; Text-fig. 14

**Etymology.** After *protervus*, the Latin for bold.

**Holotype.** NHMUK I.9609, Bembridge Marls, NW Isle of Wight, UK, Brodie collection, best preserved female wasp in side view, with legs fragmentary.



**Plate 6** (1) "*Stilpnus*" *oligocenus* Cockerell, NHMUK In.24331, holotype. (2–3) *Paxylobembra kozlovi* gen. et sp. nov., IWCMS 2012.575, holotype. Scale bars = 1 mm.



**Text-figure 15** *Stilpnus oligocenus* Cockerell, NHMUK In.24331, holotype. Scale bar = 1 mm.



**Text-figure 16** *Paxylobembra kozlovi* gen. et sp. nov., IWCMS 2012.575, holotype.

**Paratype.** NHMUK In.25765, Bembridge Marls, NW Isle of Wight, UK, Hooley collection, lateral aspect of almost complete female body lacking appendages.

**Diagnosis.** Differs from “*Hemiteles*” *acourti* in having the forewing with discocubitus weaker curved, and radial and second discoidal cells shorter; from “*Stilpnus*” *oligocenus* in having the second section of postnervulus inclivous; and from “*Hemiteles*” *dirus* in having the second recurrent vein arcuate and stronger curved near subdiscoideus, nervulus slightly postfurcal, and posterior section of postnervulus less inclivous.

**Description.** Body length 4.3 mm, mesosoma 1.4 mm, height 0.86 mm. forewing length 3.3 mm, first tergite 0.7 mm. Antenna filiform; basal flagellar segment strongly elongate, more than five times as long as wide; mid flagellar segments 1.6–2.2 times as long as wide. In paratype, frons with pair of longitudinal impressions behind antennal sockets, face with noticeable large longitudinal ridge. Temple wide, in lower part about of half eye width. Prepectal carina and mesopleural fovea present. Sternaulus long, reaching along whole mesopleuron length from its lower anterior part to posterior margin. Propodeum areolate. Pterostigma and veins dark-brown. Pterostigma wide, receiving radius at midlength. Radius slightly curved. Radial cell 2.2 times as long as wide. Areolet pentagonal, approximately equilateral, second intercubitus absent or unpigmented. Second recurrent vein inclivous, roundly and evenly curved, with two bullae. Discocubitus slightly and evenly curved, with ramulus. Second discoidal cell 1.8 times as long as wide. Nervulus somewhat postfurcal, distinctly inclivous. Postnervulus intercepted hardly below its middle, posterior sec-

tion inclivous. Legs slender. First tergite long, petiolate, petiole rectangular in cross-section, its dorsolateral and ventrolateral carinae distinct, extending along whole petiole length. Glymma absent, lateral area of first tergite flat. Metasoma depressed, widened backward, second and following tergites distinctly transverse. Ovipositor straight, almost as long as first tergite; length 0.6 mm.

**Remarks.** Paratype has no wings, but body is almost identical to that of the holotype, and apparently represents the same species. Paratype has a noticeable longitudinal ridge on the face, but face of holotype is not visible.

“*Stilpnus*” *oligocenus* Cockerell, 1921

Plate 6, fig. 1; Text-fig. 15

1921a *Stilpnus oligocenus* Cockerell, 4, fig. 5 (Cryptinae).

**Holotype.** NHMUK In.24331, Bembridge Marls, NW Isle of Wight, UK, Hooley collection, forewing and poorly preserved mesosoma and base of metasoma. Sex unknown.

**Diagnosis.** This species differs from other Cryptinae in Bembridge Marls in having shorter forewing with postnervulus intercepted low (its posterior section vertical), and with discocubitus very slightly curved.

**Description.** Small wasp with forewing length 2.7 mm. Pterostigma wide, 2.8 times as long as wide, receiving radius at midlength. Radius slightly curved. Metacarp long, but not reaching apex of forewing. Areolet pentagonal, outer side open or second intercubitus unpigmented. Second recurrent vein inclivous. Discocubitus very slightly curved, without ramulus. Nervulus very slightly postfurcal, inclivous. Postnervulus intercepted in its 0.36, its posterior section vertical.

**3.8.6. Subfamily Paxylommatinae Foerster, 1862.** Small subfamily known from Holarctic (Tobias 1988), fossils only known from the Upper Eocene Baltic and Rovno ambers (Kasparyan 1988a, 2001; Tolkanitz *et al.* 2005). Probably parasites of ants.

*Paxylobembra* Khalaim, gen. nov.

**Etymology.** After genus name *Paxylomma* and Bembridge Marls. Gender feminine.

**Type species.** *P. kozlovi* sp. nov. (by monotypy and present designation).

**Diagnosis.** New genus differs from all recent Paxylommatinae in having brachial cell not widened (as wide as submedian cell). It belongs to the tribe Tobiasitini because of lack of notauli. It differs from *Tobiasites* Kasparyan, 1988a and *Paxylommites* Kasparyan, 1988a in having flattened hind tibia and postnervulus intercepted very low (from hind outer corner of brachial cell or somewhat above it). Similarly flattened hind tibia (and hind basitarsus) are known also in the recent genus *Eurypterna* Foerster, 1862.

*Paxylobembra kozlovi* Khalaim, sp. nov.

Plate 6, figs 2, 3; Text-fig. 16

**Etymology.** In memory of Mikhail Kozlov, specialist in Proctotrupoidea.

**Holotype.** IWCMS 2012.575, Jarzembowski collection; ventral view of headless body, forewing with poorly preserved venation, and leg fragments; sex unknown.

**Diagnosis.** As for genus.

**Description.** Body length without head about 2 mm, forewing 2.25 mm. Notaulus absent. Propodeum short, without carinae. Pterostigma moderately wide, receiving radius close

to its base. Second section of radius S-curved, over five times as long as first section. Vein Rs–m short and probably thick. Second recurrent vein absent. Nervulus hardly discernible, interstitial, very slightly inclivous. Postnervulus indiscernible, intercepted at hind outer corner of brachial cell or somewhat above it. Brachial cell not widened (as wide as submedian cell). Hind tibia strongly compressed in cross-section, maximum width near apex about 2.5 times minimum width. Metasoma cylindrical. First metasomal segment slender, at least twice as long as wide, almost not widening rearward. Second metasomal segment almost 1.5 times as long as posterior width.

**3.8.7. Subfamily Townesitinae Kasparyan, 1994.** Small, extinct subfamily described from Baltic amber (Kasparyan 1994). Two Bembridge Marls species undoubtedly belong to the genus *Marjorietta* Kasparyan, 1994 because they have the frons with a keel between impressions (unknown for *M. gigantea* sp. nov.), long sternaulus, characteristic impressions on mesopleuron, and characteristic wing venation, and are very similar to *M. minor* Kasparyan. The third species (*Acourtia perplexa*) is known only from the forewing, which is similar to the wings of the genus *Marjorietta*, but the lack of the body prevents checking other important characters and makes synonymisation of *Marjorietta* under *Acourtia* premature. Accordingly, the genus *Acourtia* is retained for the present. Body length of the three Bembridge species varies from 6 mm to 8.5 mm, far above that in both species from Baltic amber (2.5–4.1 mm) and in Townesitinae in general (maximum 5 mm).

*Acourtia perplexa* Cockerell, 1921

Plate 7, fig. 3; Text-fig. 17

- 1921a *Acourtia perplexa* Cockerell, p. 11, fig. 11 (subfamily not specified).  
 1971 *Acourtia perplexa* Cockerell: Townes, p. 235 (Cremastinae?).  
 1992 *Acourtia perplexa* Cockerell: Carpenter, p. 474.  
 1994 *Acourtia perplexa* Cockerell: Kasparyan, p. 87 (Townesitinae?).

**Holotype.** NHMUK In.24336, Bembridge Marls, NW Isle of Wight, UK, Hooley collection, detached forewing.

**Diagnosis.** Differs from two other species of Townesitinae in Bembridge Marls in having the forewing with vein Rs–m very short and very thick, nervulus slightly postfurcal, and posterior section of postnervulus reclivous.

**Description.** Forewing length 4.9 mm. Pterostigma and veins dark-brown. Pterostigma almost 3.3 times as long as wide, receiving radius in its middle. First section of radius not curved near pterostigma. Second section of radius straight, 4.4 times as long as first section. Metacarp short, not reaching wing apex. Vein Rs–m very short and very thick. Second recurrent vein subtending obtuse angle with first section of subdiscoideus, straight, with single bulla slightly above middle. Discocubitus curved, without ramulus. Second discoidal cell 1.5 as long as wide. Nervulus slightly postfurcal, inclivous. Postnervulus intercepted approximately in its 0.7; posterior section distinctly reclivous. Brachial cell short.

*Marjorietta disrupta* (Cockerell, 1921), comb. nov.

Plate 7, fig. 1; Text-fig. 18

- 1921a *Lampronota disrupta* Cockerell, p. 10, fig. 9 (Banchinae).

**Holotype.** NHMUK I.9711, Bembridge Marls, NW Isle of Wight, UK, Brodie collection, lateral aspect of almost complete female wasp lacking antennal apex and most of legs and

with metasoma detached, obscured by wings basally, and lacking ovipositor apex.

**Diagnosis.** Differs from *A. gigantea* in having the forewing with the first section of radius curved near pterostigma, vein Rs–m moderately thick, second recurrent vein subtend angle 95° with the first section of subdiscoideus, the second discoidal cell longer and postnervulus intercepted lower. Differs from *A. perplexa* in having the vein Rs–m longer and thinner, nervulus almost interstitial, and posterior section of postnervulus inclivous.

**Description.** Body length about 6 mm, forewing 5 mm, mesosoma length 2.1 mm, width 1.5 mm, forewing length 5 mm. Antenna filiform as preserved; scape elongate, oval. Inner margins of eyes convergent ventrally. Temple about one third as wide as eye. Frons with pair of longitudinal impressions behind antennal sockets and with longitudinal keel between these impressions. Impressions deep, with slight slope to frontal ocellus and reaching about its level. Keel high and narrow, higher near antennal sockets and decreasing towards frontal ocellus. Notaulus absent. Prepectal and postpectal carinae distinct. Mesopleuron with obvious longitudinal impression in upper part near tegula, wider and more shallow anteriorly, deeper and more narrow posteriorly, and with more shallow oblique impression extending through mesopleuron from upper anterior to lower posterior corners. Sternaulus strong, deep, crenulate, extending for entire mesopleuron length in its lower part, joining its posterior margin below level of mid coxa. Mesopleural fovea absent. Mesopleural suture straight. Propodeum areolate. Mid and hind coxa and hind femur very slender. Pterostigma and veins dark-brown. Pterostigma narrow, almost five times as long as wide, receiving radius near its middle. First section of radius curved near metacarp. Second section of radius almost straight, 3.5 times as long as first section. Metacarp short, not reaching apex of forewing. Vein Rs–m moderately thickened. Second recurrent vein subtending obtuse angle with first section of subdiscoideus, not curved, with single bulla in the middle. Discocubitus slightly curved, without ramulus. Second discoidal cell 1.65 times longer than wide. Nervulus hardly postfurcal, inclivous. Postnervulus intercepted in its 0.6; posterior section slightly inclivous. Brachial cell short. Nervulus intercepted obviously below middle. Mediella moderately curved. Metasoma cylindrical (first segment not preserved). Ovipositor as preserved straight, longer than height of metasoma.

*Marjorietta gigantea* Khalaim, sp. nov.

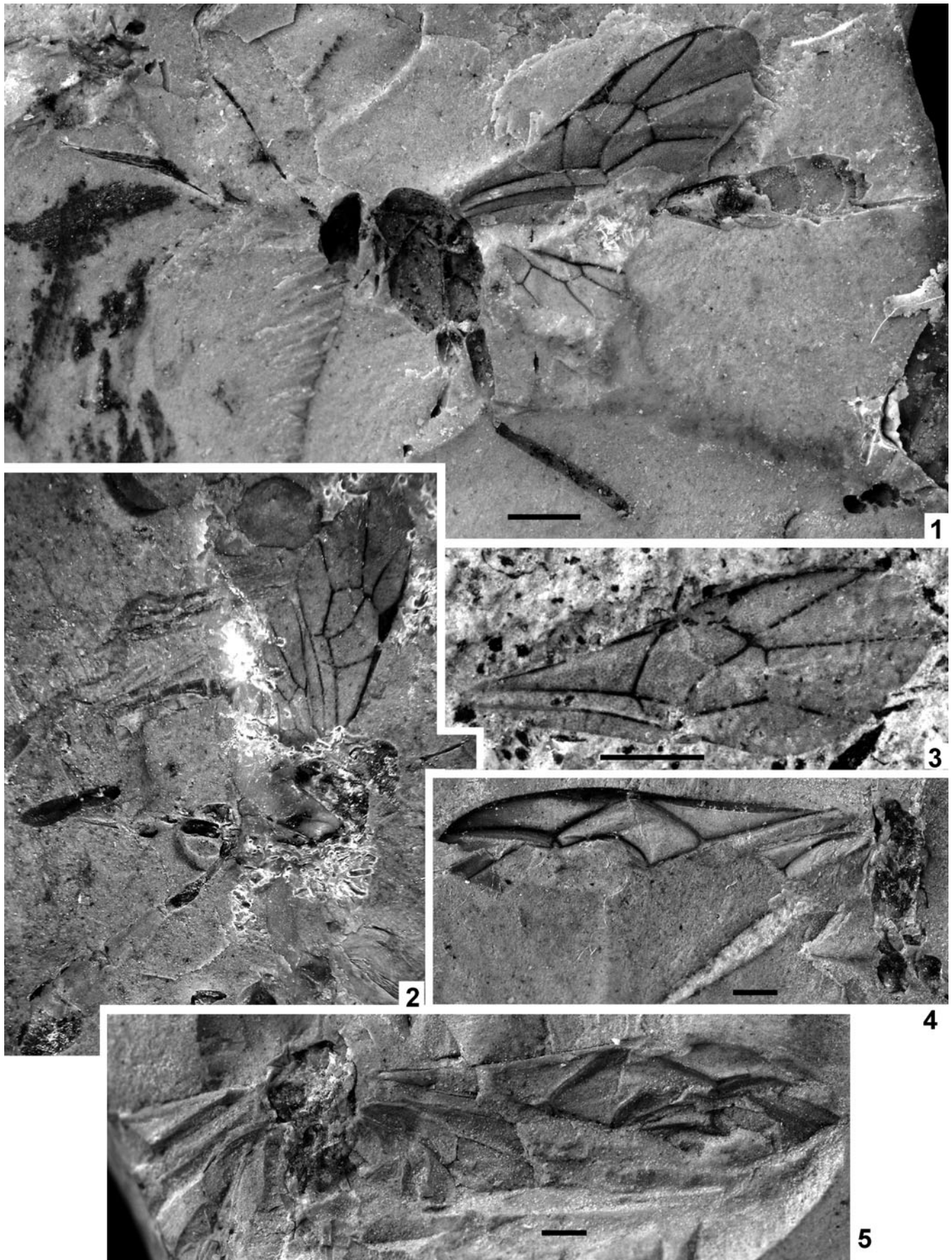
Plate 7, fig. 2; Text-fig. 19

**Etymology.** After *giganteus*, the Latin for giant.

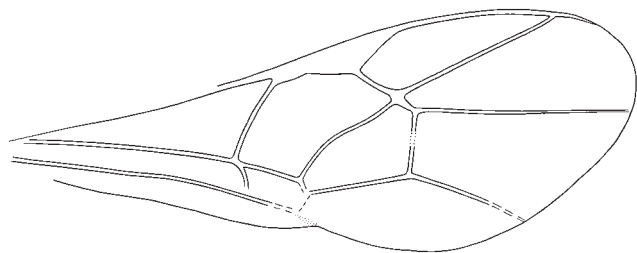
**Holotype.** NHMUK In.43477(1), Bembridge Marls, NW Isle of Wight, UK, Hooley collection, ventrolateral aspect of wasp with head poorly preserved, mesosoma incomplete posteriorly, antenna and legs incomplete; sex unknown.

**Diagnosis.** Differs from *A. disrupta* in having forewing with the first section of radius not curved near pterostigma, vein Rs–m thick, second recurrent vein subtending angle of 105° with first section of subdiscoideus, second discoidal cell shorter, and postnervulus intercepted higher. Differs from *A. perplexa* in having the vein Rs–m longer and thinner, nervulus almost interstitial, and posterior section of postnervulus inclivous.

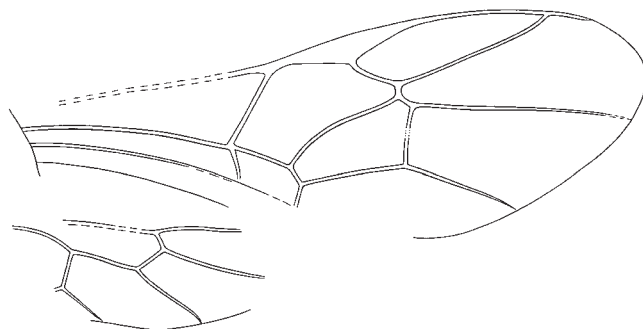
**Description.** Body length 8.5 mm, forewing c. 5 mm. Flagellar base slender; 1st and 2nd flagellomeres at least twice as long as wide. Mesosoma very similar to that of *Marjorietta disrupta*. Prepectal carina distinct. Mesopleuron with distinct subtegular impression, and with oblique impression extending from upper anterior to lower posterior corner of mesopleuron. Sternaulus deep, beginning from anteroventral margin of mesopleuron



**Plate 7** (1) *Marjorietta disrupta* (Cockerell), NHMUK I.9711, holotype. (2) *Marjorietta gigantea* sp. nov., NHMUK In.43477, holotype. (3) *Acourtia perplexa* Cockerell, NHMUK In. 24336, holotype. (4–5) “*Coleocentrus*” *gurnetensis* Cockerell: (4) NHMUK In.17079, holotype; (5) NHMUK In.17296, counterpart. Scale bars = 1 mm.



**Text-figure 17** *Acourtia perplexa* Cockerell, NHMUK In.24336, holotype.



**Text-figure 18** *Marjorietta disrupta* (Cockerell), NHMUK I.9711, holotype.

(posterior part not preserved). Mesopleural fovea absent. Mesopleural suture straight. Pterostigma and veins dark-brown. Pterostigma wide, 3·4 times as long as wide, receiving radius near its middle. First section of radius not curved near metacarp. Second section of radius almost straight, 4·5 times as long as first section. Vein Rs–m short and thick. Second recurrent vein subtending rather obtuse angle with first section of subdiscoideus, not curved, with single bulla somewhat above the middle. Discocubitus slightly curved, without ramulus. Second discoidal cell 1·35 times as long as wide. Nervulus strongly interstitial, inclivous. Postnervulus intercepted in its 0·7; posterior section hardly inclivous (almost vertical). Brachial cell short. Nervellus probably intercepted in the middle. Mediella slightly curved. Metasoma cylindrical, wider posteriorly. First segment thin and long, with distinct petiole and postpetiole. Second segment distinctly elongate. Third segment about as long as wide.

*Ichneumonidae incerta sedis*

“*Coleocentrus*” *gurnetensis* Cockerell, 1921

Plate 7, figs 4, 5; Text-fig. 20

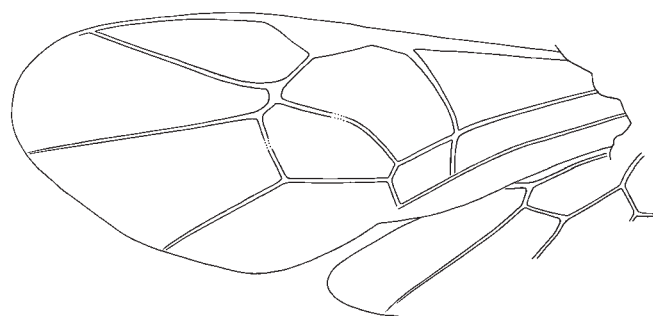
1921a *Coleocentrus gurnetensis* Cockerell, p. 6, fig. 3 (Acaenitinae).

**Holotype.** NHMUK In.17079/In. 17296, Bembridge Marls, NW Isle of Wight, UK, Smith collection; crumpled wings, poorly preserved mesosoma; sex unknown.

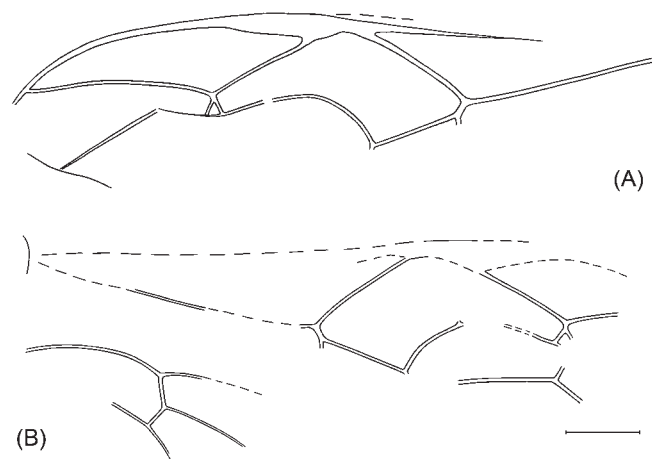
**Diagnosis.** Differs from other Bembridge Marls ichneumonids in having the pterostigma very narrow, long and large.

**Description.** Forewing length 10 mm. Pterostigma narrow, receiving radius basally. First section of radius not curved. Areolet present. Discocubitus curved, ramulus present. Nervulus distinctly postfurcal. Nervellus intercepted in its lower 0·4. Mediella curved.

**Remarks.** Areolet and narrow pterostigma receiving radius basally are characteristic for the tribe Coleocentrini, but the nervellus intercepted low excludes this species from the genus



**Text-figure 19** *Marjorietta gigantea* sp. nov., NHMUK In.43477, holotype.



**Text-figure 20** *Coleocentrus gurnetensis* Cockerell: (A) NHMUK In.17079 (holotype); (B) NHMUK In.17296 (counterpart). Scale bar = 1 mm.

*Coleocentrus*, and from this tribe that has the nervellus intercepted in the upper 0·32 or higher.

“*Polysphincta*” *atavina* Cockerell, 1921

Plate 8, fig. 1; Text-fig. 21

1921a *Polysphincta* (?) *atavina* Cockerell, p. 9, fig. 8 (Pimplinae).

**Holotype.** NHMUK In.24335, Bembridge Marls, NW Isle of Wight, UK, Hooley collection, poorly preserved forewing, sex unknown.

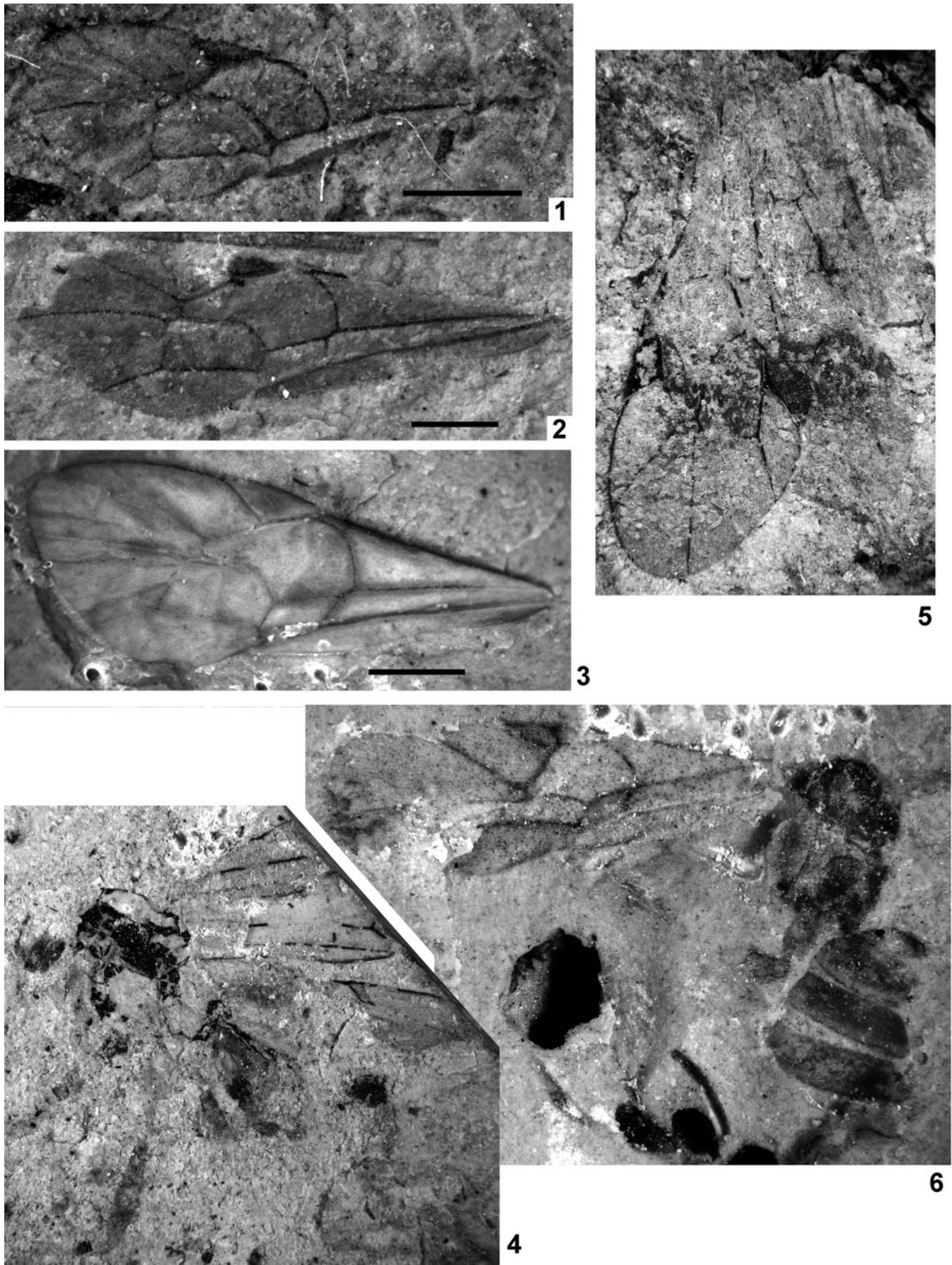
**Diagnosis.** Differs from other Bembridge Marls ichneumonids in the unusual shape of areolet which adjoins broadly to the radius (Text-fig. 21).

**Description.** Forewing length over 4 mm. Pterostigma and veins predominantly brown, stigma yellowish medially. Pterostigma wide, almost three times as long as wide, receiving radius near its middle. First section of radius curved. First intercubitus very short. Second intercubitus probably present, unpigmented. Second recurrent vein moderately curved, with two bullae. Discocubitus curved, ramulus absent. Nervulus interstitial or hardly postfurcal, inclivous. Postnervulus intercepted near its middle.

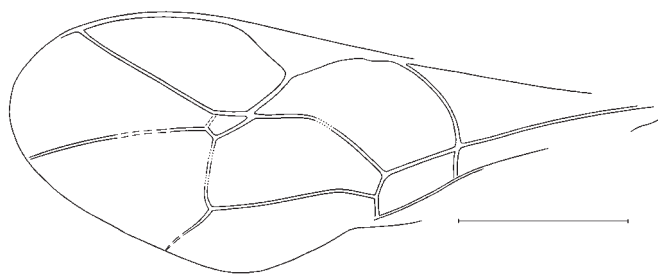
**Remarks.** Forewing venation generally similar to venation in Pimplinae, but areolet is not typical for this subfamily (second intercubitus unclear).

Genus *Lithapecthis* Cockerell, 1921

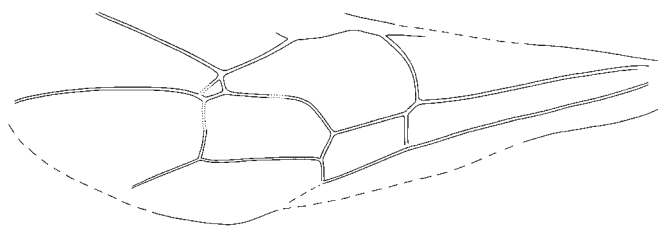
**Type species.** *Lithapecthis fumosus* Cockerell, 1921 (by monotypy).



**Plate 8** (1) "*Polysphincta*" *atavina* Cockerell, NHMUK In.24335, holotype. (2) *Lithapectis fumosus* Cockerell, NHMUK In.24333, holotype. (3) *Lithapectis salmacidus* sp. nov., NHMUK In.24839, holotype. (4–6) Ichneumonidae indet.: (4) NHMUK I.9311; (5) NHMUK I.9354; (6) NHMUK I.9812. Scale bars = 1 mm.



**Text-figure 21** *Polysphincta atavina* Cockerell, NHMUK In.24335, holotype. Scale bar = 1 mm.



**Text-figure 22** *Lithapectis fumosus* Cockerell, NHMUK In.24333, holotype.

**Diagnosis.** Pterostigma more or less wide, receiving radius near its middle. Areolet small, oblique, with long petiole; first intercubitus shorter than second intercubitus; second intercubitus unpigmented. Second recurrent vein almost straight, slightly curved in middle, probably with two bullae (Text-fig. 23), inclivous or subvertical. Discocubitus rather strongly curved, without ramulus. Second discoidal cell short, 1.7–1.9 times as long as wide. Nervulus postfurcal, distance between basal vein and nervulus is half the length of nervulus. Postnervulus intercepted near or below its middle.

**Remarks.** Cockerell supposed that this genus belongs to Pimplinae. I have studied the type and two additional specimens which I refer to the same genus, all known from isolated forewings. *Lithapectis* is well distinguishable from other ichneumonid genera found in the Bembridge Marls, mainly by the combination of the narrow pedunculate areolet, characteristic form of the second recurrent vein, and postfurcal nervulus, but I cannot recognise its subfamily. Similar forewing venation can be found in extant specimens of the subfamilies Ctenopelmatinae, Tryphoninae, Metopiinae and Banchinae.

*Lithapectis fumosus* Cockerell, 1921

Plate 8, fig. 2; Text-fig. 22.

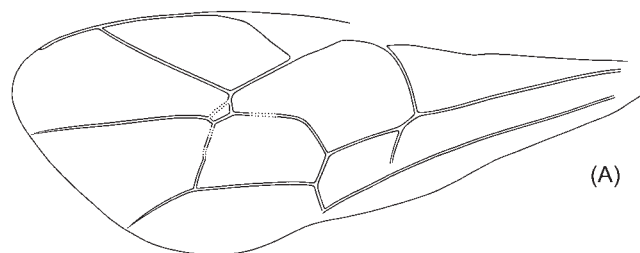
1921a *Lithapectis fumosus* Cockerell, p. 6, fig. 4 (Pimplinae).

1992 *Lithapectis fumosus* Cockerell: Carpenter, p. 481.

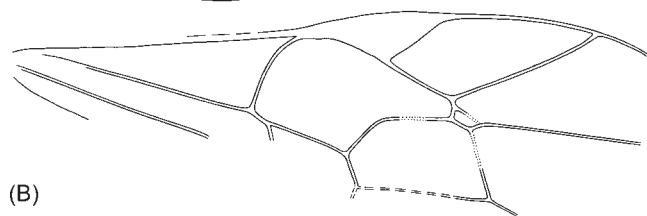
**Holotype.** NHMUK In.24333, Bembridge Marls, NW Isle of Wight, UK, Hooley collection, poorly preserved forewing; sex unknown.

**Diagnosis.** Differs from *L. salmacidus* sp. nov. in having the forewing with the second recurrent vein subvertical, areolet with shorter petiole, second discoidal cell longer, discocubitus moderately curved, and nervulus weakly inclivous.

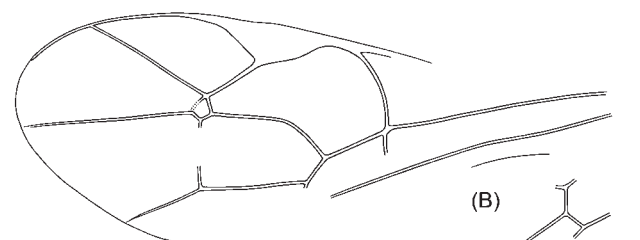
**Description.** Forewing length over 6.5 mm. Pterostigma and veins predominantly brown; costa and veins in basal part of wing dark-brown to black; stigma yellowish basally. Pterostigma more or less wide, receiving radius near its middle. Areolet petiolate, first intercubitus shorter than second inter-



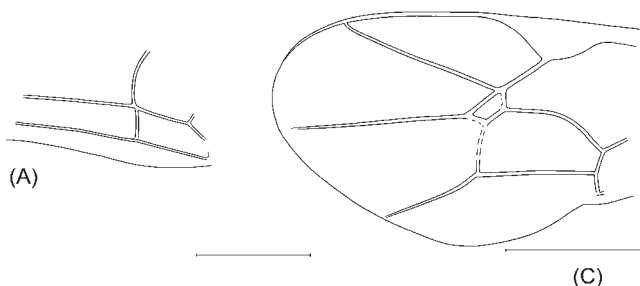
(B)



**Text-figure 23** *Lithapectis salmacidus* sp. nov., holotype: (A) part, NHMUK In.24839; (B) counterpart, NHMUK In.24791.



(A)



**Text-figure 24** Ichneumonidae indet.: (A) NHMUK I.9311; (B) NHMUK I.9354; (C) NHMUK In.17430. Scale bars = 1 mm.

cubitus. Second recurrent vein slightly curved, subvertical. Discocubitus curved, without ramulus. Second discoidal cell 1.9 times as long as wide. Nervulus weakly inclivous, postfurcal (distance between basal vein and nervulus equal to half nervulus length). Postnervulus intercepted in its middle.

*Lithapectis salmacidus* Khalaim, sp. nov.

Plate 8, fig. 3; Text-fig. 23

**Etymology.** After *salmacidus*, the Latin for salty sour.

**Holotype.** NHMUK In.24839, NHMUK In. 24791 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Hooley collection; well-preserved complete forewing. Sex unknown.

**Diagnosis.** Differs from *L. fumosus* in having the forewing with the second recurrent vein distinctly inclivous, areolet with long petiole, second discoidal cell shorter, discocubitus stronger curved, and nervulus strongly inclivous.

**Description.** Forewing length 5.5 mm. Veins yellowish; pterostigma whitish. Pterostigma wide, receiving radius near

its middle. Metacarp not reaching apex of forewing. Radius more or less straight; second section 2.2 times longer than first section. Areolet petiolate, with long petiole; first intercubitus shorter than second intercubitus. Second intercubitus unpigmented. Second recurrent vein almost straight, slightly curved in the middle, inclivous. Discocubitus strongly curved, without ramulus. Second discoidal cell 1.7–1.75 times longer than wide. Nervulus strongly inclivous, postfurcal (distance between basal vein and nervulus half as long as nervulus). Postnervulus intercepted somewhat below its middle.

#### Ichneumonidae indet.

**Remarks.** Additionally found are six unnamed ichneumonid fossils which cannot be identified even up to subfamily level: NHMUK I.9214, NHMUK I.9311 (Plate 8, fig. 4); NHMUK I.9354 (Plate 9, fig. 5, Text-fig. 24B); NHMUK I.9812 (Plate 8, fig. 6); and NHMUK In.17430, some of which are worth further discussion.

Specimen NHMUK I.9311: Bembridge Marls, NW Isle of Wight, UK, Brodie collection, dorsal view of poorly preserved mesosoma and base of metasoma, wing bases and parts of mid and hind legs (Plate 8, fig. 4, Text-fig. 24A). Nervulus and postnervulus in forewing are similar to these in *Hemiteles acourti*, but those characters are insufficient to identify the position of the fossil.

Specimen NHMUK I.9812: Bembridge Marls, NW Isle of Wight, UK, Brodie collection, poorly preserved mesosoma, part of metasoma and crumpled forewing (Plate 8, fig. 6). Strongly transverse mid-metasomal segments might indicate Pimplinae.

Specimen NHMUK In.17430: Bembridge Marls, NW Isle of Wight, UK, Smith collection, poorly preserved apical half of forewing (Fig. 24C), possibly Pimplinae.

#### 3.8.8. Key to species of Bembridge Marls Ichneumonidae

- 1 – Forewing with second recurrent vein entirely absent (Text-fig. 16); second section of radius S-curved, over five times as long as first section; small species with forewing 2.25 mm; brachial cell not widened (as wide as submedian cell); notauli absent; hind tibia strongly compressed in cross-section, maximum width near apex about 2.5 times minimum width. (Paxylommatinae) = *Paxylobembra kozlovi* sp. nov.
- Forewing with second recurrent vein present. Second section of radius straight or arcuate, usually much shorter. . . . .go to 2.
- 2 – Second recurrent vein straight, with one bulla, distinctly reclivous (Text-figs 17–19); areolet absent, vein Rs–m moderately to very thick; second section of radius 3.5–4.5 times as long as first section; postnervulus intercepted always distinctly above its middle. (Townesitinae). . . . .go to 3.
- Second recurrent vein often arcuate, with one or two bullae, usually inclivous or rarely subvertical; areolet absent or present, vein Rs–m thin; second section of radius usually shorter; postnervulus intercepted near or below, or rarely above its middle. . . . .go to 6.
- 3 – Body length 2.5–4.0 mm; forewing length at the most 3.3 mm = Baltic amber species of *Marjorietta* Kasparyan.
- Body length 6.5–8.5 mm; forewing about 5 mm. Bembridge Marls. . . . .go to 4.
- 4 – Vein Rs–m very short and very thick (Text-fig. 17); nervulus slightly postfurcal; posterior section of postnervulus reclivous = *Acourtia perplexa* Cockerell.
- Vein Rs–m longer and thinner; nervulus almost interstitial; posterior section of postnervulus more or less inclivous. . . . .go to 5.
- 5 – First section of radius not curved near pterostigma (Text-fig. 19); vein Rs–m thick; second recurrent vein subtend angle 105° with first section of subdiscoideus; second discoidal cell 1.35 times as long as wide; postnervulus intercepted in its 0.7 = *Marjorietta gigantea* sp. nov.
- First section of radius distinctly curved near pterostigma (Text-fig. 18); vein Rs–m moderately thick; second recurrent vein subtend angle 95° with first section of subdiscoideus; second discoidal cell 1.5 times as long as wide; postnervulus intercepted in its 0.6 = *Marjorietta disrupta* (Cockerell).
- 6 – Areolet quadrangular, strongly oblique, with long petiole above (Text-figs 22, 23); second recurrent vein slightly inclivous, almost straight, weakly sinuate centrally, probably with two bullae; nervulus strongly postfurcal, distance between basal vein and nervulus equal to half of nervulus length. (*Lithapechtis* Cockerell). . . . .go to 7.
- Areolet (if present) quadrangular or pentagonal, sometimes pointed above but without distinct petiole (Text-figs 5, 8); second recurrent vein straight or arcuate, not sinuate centrally, with one or two bullae; nervulus interstitial or weakly postfurcal (except for *Hypsicera anglica* with this vein strongly postfurcal). . . . .go to 8.
- 7 – Second recurrent vein distinctly inclivous (Text-fig. 23); areolet with long petiole; second discoidal cell 1.7–1.75 times as long as wide; discocubitus stronger curved; nervulus strongly inclivous = *Lithapechtis salmacidis* sp. nov.
- Second recurrent vein subvertical (Text-fig. 22); areolet with short petiole; second discoidal cell 1.9 times as long as wide; discocubitus moderately curved; nervulus weakly inclivous = *Lithapechtis fumosus* Cockerell.
- 8 – Areolet complete (except for *Eusterinx humalai* sp. nov. which has no areolet), quadrangular, more or less pointed above (Text-figs 4–6, 8–10); second recurrent vein moderately to strongly arcuate (except for *Eusterinx vectensis* which has this vein almost straight); nervulus interstitial or slightly postfurcal. . . . .go to 9.
- Areolet incomplete (open distally), pentagonal, not pointed above (Text-figs 11–15); second recurrent vein straight or arcuate; nervulus sometimes far postfurcal. . . . .go to 14.
- 9 – Second recurrent vein strongly arcuate and areolet weakly to strongly oblique (Text-figs 3–6). (Pimplinae). . . . .go to 10.
- Second recurrent vein almost straight and areolet weakly oblique (subrhombic), or areolet absent (Text-fig. 10). (Orthocentrinae). . . . .go to 12.
- 10 – Nervulus postfurcal, distinctly inclivous (Text-fig. 5); discocubitus very weakly curved ? = *Itopectis saxosa* Cockerell.
- Nervulus interstitial, slightly inclivous or subvertical. Discocubitus distinctly curved. . . . .go to 11.
- 11 – Areolet strongly oblique (Text-fig. 6); first intercubitus much shorter than second intercubitus; postnervulus intercepted high = *Scambus fossilis* sp. nov.
- Areolet weakly oblique (Text-fig. 4); first intercubitus subequal to second intercubitus; postnervulus intercepted low. = *Exeristes gurnetor* sp. nov.
- 12 – Forewing without areolet; first section of radius straight; discocubitus very slightly curved; nervulus inclivous; postnervulus intercepted low = *Eusterinx humalai* sp. nov.
- Forewing with areolet; first section of radius more or less curved; discocubitus distinctly curved; nervulus vertical; postnervulus intercepted low. . . . .go to 13.
- 13 – Areolet large, first intercubitus long; discocubitus moderately curved = *Eusterinx vectensis* (Cockerell).

- Areolet small, oblique, first intercubitus very short; discocubitus strongly curved = *Eusterinx arcuatus* (Cockerell).
- 14 – Nervulus strongly postfurcal and strongly inclivous (Text-fig. 11); second recurrent vein straight; first intercubitus and abscissa of cubitus between first intercubitus and second recurrent vein long; head globose; body robust; hind coxa and femur thick. (Metopiinae) = *Hypsicera anglica* (Cockerell).
- Nervulus interstitial or slightly postfurcal, weakly inclivous. Second recurrent vein usually arcuate. First intercubitus and abscissa of cubitus between first intercubitus and second recurrent vein usually much shorter. (Cryptinae). . . . go to 15.
- 15 – Postnervulus intercepted in its 0.35; posterior section vertical; discocubitus very slightly curved; small insect with forewing length 2.7 mm = *Stilpnus oligocenus* Cockerell.
- Postnervulus sometimes intercepted higher; posterior section inclivous; discocubitus distinctly curved; forewing usually longer. . . . go to 16.
- 16 – Second recurrent vein strongly inclivous, almost straight, not curved near subdiscoideus; nervulus interstitial; postnervulus strongly inclivous, angle between posterior section and brachius 50°; discocubitus slightly and evenly curved = *Hemiteles dirus* sp. nov.
- Second recurrent vein inclivous, roundly curved, stronger curved near subdiscoideus; nervulus more or less postfurcal; posterior section of postnervulus less inclivous; discocubitus usually stronger curved. . . . go to 17.
- 17 – Discocubitus at most very weakly curved anteriorly and posteriorly, and strongly curved near the middle; nervulus very slightly inclivous, distinctly postfurcal; radial cell 2.5 times as long as wide; second discoidal cell 2.15 times as long as wide = *Hemiteles acourti* (Cockerell).
- Discocubitus more or less evenly curved; nervulus distinctly inclivous, somewhat postfurcal; radial cell 2.2 times as long as wide; second discoidal cell 1.8 times as long as wide = *Hemiteles protervus* sp. nov.

**3.8.9. Discussion.** Taxonomic composition of Ichneumonidae of the Bembridge Marls gives little information about ecological and climatic conditions. Orthocentrinae indicate humid forests; other subfamilies do not show clear biotopic preferences. Parasites of Lepidoptera (Metopiinae, and probable Pimplinae and Cryptinae), mycetophagous Diptera (Orthocentrinae) and probable ants (Paxylommatidae) are found.

The Bembridge Marls ichneumonid fauna is generally closest to that of Baltic amber, both being dominated by Cryptinae and Townesitinae with presence of Paxylommatinae. Cryptinae are abundant in many Cenozoic insect sites, but Townesitinae are known only from the Bembridge Marls and Baltic amber. However, these two assemblages differ strikingly in two respects. First, the extinct subfamily Pherhombinae is rather common in the Baltic and Rovno ambers (Kasparyan 1988b, Tolkanitz *et al.* 2005) but not recorded in the Bembridge Marls (nor in any other Cenozoic deposits). Secondly, the subfamily Pimplinae dominates in the Bembridge Marls assemblage and is rather common in other Cenozoic deposits since the Paleocene, but is not recorded in the Baltic amber. The lack of Pherhombinae in the Bembridge Marls indicates it is somewhat younger than Baltic amber.

Both the Bembridge Marls and the Baltic amber deposits are apparently older than the Florissant beds. This inference is based on the subfamily Ichneumoninae, a relatively progressive group of Ichneumonidae (Gokhman 1992) which is abundant in the Florissant and recorded neither in the Bembridge Marls nor in the Baltic amber. Additionally, the Florissant assemblage lacks the fossil subfamily Townesitinae which

constitutes an appreciable part of both the Bembridge Marls and the Baltic amber ichneumonid assemblages. Many Recent subfamilies are recorded from the Florissant which are absent from the Bembridge Marls (Brues 1910), but that material deserves revision before being used in more detailed comparisons.

### 3.9. Family Braconidae Nees, 1812

[By Sergey A. Belokobylskij.]

The oldest fossils of Braconidae are known from the Cretaceous Period (Carpenter *et al.* 1937; Zherikhin 1978; Rasnitsyn 1983; Basibuyuk *et al.* 1999; Perrichot *et al.* 2007, 2009; Ortega-Blanco *et al.* 2009), but the most numerous and diverse members of this family were discovered and described from Cenozoic (Oligocene, Miocene and Eocene) ambers in different parts of the World (Baltic: Brues 1923, 1933, 1939; Tobias 1987; Dominican: Zuparko & Poinar 1997; Achterberg 2001; Engel & Bennett 2008; Mexican: Muesebeck 1960; Chinese: Hong 2001, 2002), and different rock deposits (Cockerell 1919, 1920a, 1921a; Brues 1906; Statz 1936, 1938).

In spite of the quite extensive published information about braconid fossils, the positions of many taxa are very problematic, because they were not described by braconid experts. As a result, many fossils have been described in incorrect genera and subfamilies. Unfortunately, the largest collection of Baltic amber from Königsberg University, with numerous types of Hymenoptera, was mostly destroyed during the Second World War. This includes the Braconidae described by Brues and now it is almost impossible to check and correct the attribution of many of Brues' taxa because of incomplete descriptions and often insufficiently informative figures. Undoubtedly, revision of the all available described taxa is one of the main steps toward the real estimation of the taxonomic position of fossil Braconidae.

All the types of species described by Cockerell (1921a) from the Bembridge Marls of the Isle of Wight are kept in good condition in the collection of the Palaeontological Department of the Natural History Museum in London. The study of this collection has enabled the modern taxonomic position of these taxa to be established. Additional material from this collection shows that the diversity of the braconid fossil assemblage is wide and includes several additional subfamilies not recorded previously.

Most of the fossils studied show meso- and metasoma preserved three-dimensionally (including depth), and their study requires observations from different visual angles. As a result, several characters mentioned in the descriptions cannot be seen on photos or line drawings. Also, the sex of the fossils under study is often unknown, because the respective characters are not available due to incomplete preservation.

The venational terminology and symbols follow that of Achterberg (1979) (see Text-fig. 29B and additions in Text-figs 26 and 43A).

**3.9.1. Subfamily Agathidinae Foerster, 1862.** Koinobiont endoparasitoids of lepidopteran larvae of the subfamily Agathidinae are not abundant in the fossil record, including amber. Three species of *Agathis* Latreille, 1804 (*A. juvenilis* Brues, 1910, *A. saxatilis* Brues, 1910 and *A. velatus* Brues, 1910) described by Brues (1910) from the Florissant beds are now considered as uppermost Eocene (Evanoff *et al.* 2001). Of these taxa, *A. saxatilis* is undoubtedly a species of the genus *Earinus* Wesmael, 1837 because of the presence of the distinct and complete 1-SR+M in the forewing. Two others are probably species of *Bassus* Fabricius, 1804, judging from the head being figured as not elongate in its lower half and with the mouth apparatus short. Two species with broad second submarginal cell

in the forewing from the Florissant (*Microdus miocenicus* Cockerell, 1927) and from the Baltic amber (*M. quadrangularis* Brues, 1933) belong to the genus *Bassus* in the modern sense as well. The generic position of *Cremnops florissanticola* Cockerell, 1919 (Cockerell 1919; Brues 1933) from the Florissant needs verification: its wing venation as figured is very close to that of other fossil *Bassus* with the enlarged second submarginal cell. Lastly, *Snellenius succinalis* Brues, 1933 described in this subfamily, but in the microgastrine genus *Snellenius* Westwood, 1882 (Brues 1933), is not an agathidine: its taxonomic position needs revision.

Two well preserved forewings discovered in the collection from the Bembridge Marls are described below. Several agathidine genera (*Cremnops* Foerster, 1862, *Labagathis* Enderlein, 1920, *Zacremnops* Sharkey & Wharton, 1985), as well as some species of other genera (*Bassus*, *Agathis*, *Euagathis* Szepligeti, 1900, *Braunsia* Kriechbaumer, 1894) have similar venation. That is why the new species are provisionally included in *Bassus* as one of the less derived agathidine genera, and additionally because a number of species of comparable age and with similar wing venation are already known to belong to *Bassus* (see above).

Genus *Bassus* Fabricius, 1804

*Bassus magnareola* Belokobylskij, sp. nov.

Plate 9, fig. 1; Text-fig. 25A

**Etymology.** From *magnus*, the Latin for large, and *areola*, the Latin for cell, because of the enlarged second submarginal cell of forewing.

**Holotype.** NHMUK In.17127, Bembridge Marls, NW Isle of Wight, UK, Smith Collection; isolated forewing.

**Diagnosis.** Similar to *B. quadrangularis* (Brues, 1933) in large second submarginal cell but differs in having 1-R1 shorter than pterostigma, second submarginal cell very weakly narrowed above, and r much shorter than 3-SR. Differs from insufficiently described *B. miocenicus* (Cockerell, 1927) (= *Microdus miocenicus* Cockerell, 1927, Florissant), comb. nov., in having subsquare second submarginal cell.

**Description.** Forewing length  $c.3.3$  mm. C+SC+R thick. Pterostigma subtriangular, wide, rather short,  $c.2.5$  times as long as maximum width. 1-R1 0.75 times as long as pterostigma, almost as long as distance from apex of marginal cell to wing apex; r arising almost from middle of pterostigma, short,  $c.0.2$  times as long as maximum pterostigmal width, 0.5 times as long as 3-SR. Marginal cell much shortened, narrow, pointed apically, 4.5 times as long as maximum width. 3-SR 0.15 times as long as weakly concave SR1, 0.6 times as long as 2-SR; r-m distinctly sclerotised, weakly broken in anterior 0.4. Second submarginal cell subsquare, weakly narrowed anteriorly, 0.9 times as long as wide, its anterior side 0.8 times as long as posterior side. Short basal and apical sections of 1-SR+M developed; m-cu distinctly antefurcal, as long as 2-SR, 0.7 times as long as weakly curved 1-M and subparallel to it; cu-a distinctly postfurcal. Colour: Forewing hyaline; pterostigma and C+SC+R dark brown; remaining veins brown to pale brown.

*Bassus quadrangularis* (Brues, 1933), comb. nov.

Plate 9, fig. 2; Text-fig. 25B

1933 *Microdus quadrangularis* Brues, 95. Baltic amber; Upper Eocene.

**Holotype.** Female, Baltic amber, without number (possibly lost).

**Material examined.** NHMUK In.24958, Bembridge Marls, NW Isle of Wight, UK, Hooley Collection; isolated forewing with basal part lost.

**Diagnosis.** Similar to *B. magnareola* sp. nov. but differs in having 1-R1 almost as long as pterostigma, second submarginal cell distinctly narrowed above, and r almost as long as 3-SR.

**Description.** C+SC+R thick. Pterostigma subtriangular, wide, rather short,  $c.2.5$  times as long as maximum width. 1-R1 almost as long as pterostigma,  $c.1.3$  times as long as distance from apex of marginal cell to wing apex; r arising almost from middle of pterostigma, short,  $c.0.4$  times as long as maximum width of pterostigma, almost as long as 3-SR. Marginal cell distinctly shortened, narrow, pointed apically, almost five times as long as maximum width. 3-SR 0.1 times as long as the straight SR1, 0.4 times as long as 2-SR; r-m distinctly sclerotised, weakly broken in anterior 0.3. Second submarginal cell trapezoid, narrowed anteriorly, about 0.8 times as long as wide, its anterior side 0.6 times as long as posterior side. Short basal and apical sections of 1-SR+M present; m-cu distinctly antefurcal, almost as long as 2-SR, 0.6 times as long as distinctly curved 1-M and weakly divergent from it. Cu-a postfurcal, distant from 1-M for 0.3 times cu-a length. Subdiscal cell wide,  $c.3.0$  times as long as wide; CU1b absent.

**Remarks.** According to the short original description of the forewing, the present fossil shows no differences from *B. quadrangularis*. I prefer to use this name until new material is available.

**3.9.2. Subfamily Brachistinae Foerster, 1862.** Brachistinae are ovo-larval parasitoids of various beetles. One of the main features discriminating this group from the closely related subfamily Helconinae is the absence of r-m and, as a result, of the second submarginal cell. Unfortunately, not every fossil has its forewing preserved well enough to permit reliable identification. This results in tentative placement of insufficiently preserved fossils.

In the fossil record, the subfamily is represented by only a few species of two genera. These are *Eubazus crassicornis* (Brues, 1933) and *E. gracilicornis* (Brues, 1939) (originally described in *Blacus*: Achterberg 1982), *E. normalis* Brues, 1923, comb. nov. and *E. wilmattae* Brues, 1910, comb. nov. (originally described in *Brachistes* and *Calyptus* respectively) and *Triaspis anomala* Brues, 1939 (all from the Baltic amber, except for *E. wilmattae* from the Florissant). Described from the Florissant, *Urosigalphus aternus* Brues, 1910 is possibly also a member of the brachistine genera *Triaspis* or *Schizoprymnus*. *Calyptoides veteris* Cockerell, described putatively as a brachistine wasp (Cockerell 1921a), actually belongs to subfamily Helconinae (see below).

Five new species of the genus *Eubazus* are described from the Insect Limestone, Bembridge Marls, in this study. The subgeneric position of these new species is not clear because of insufficient morphological information available from these rock fossils.

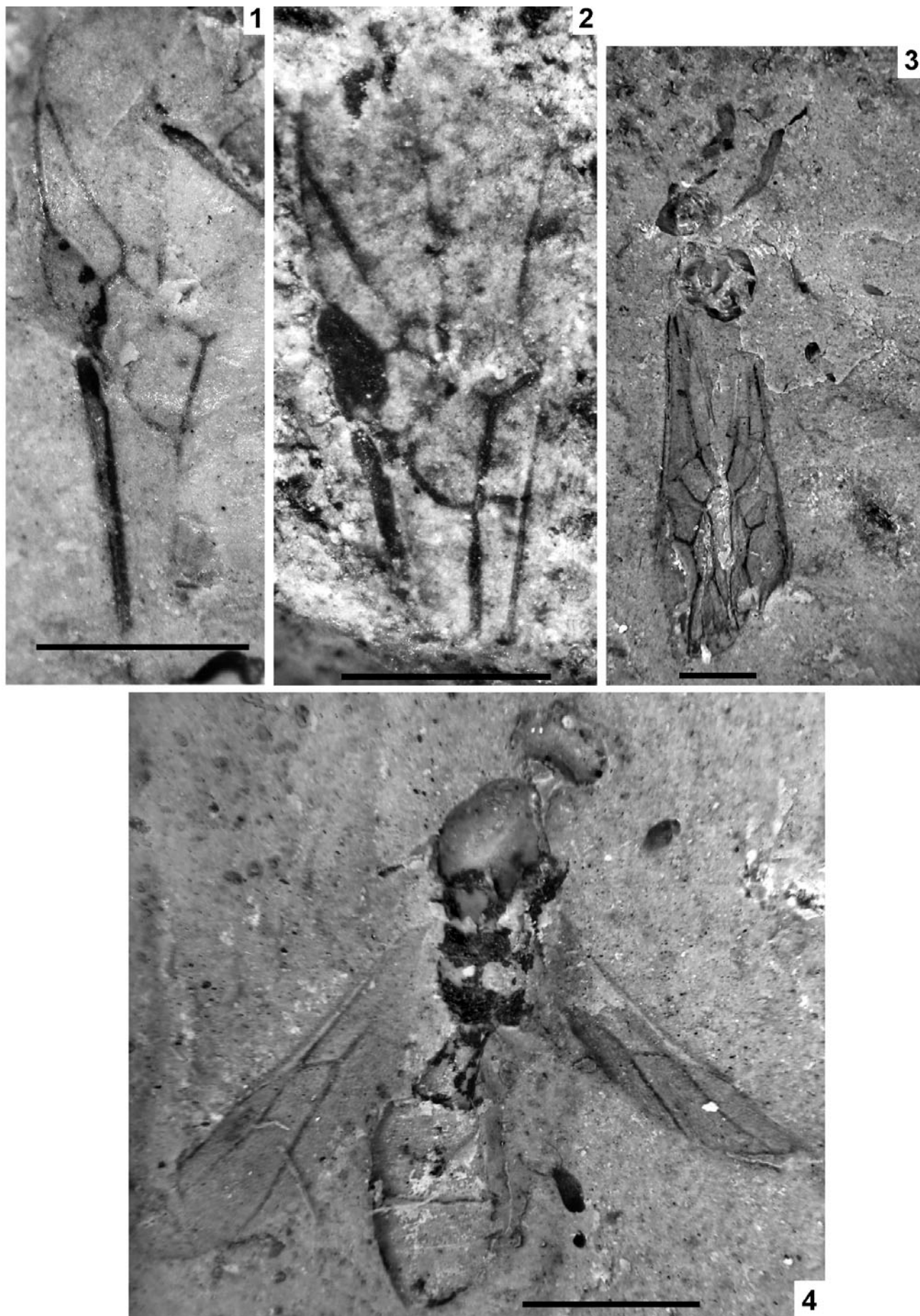
Genus *Eubazus* Nees, 1816

*Eubazus ? brodiei* Belokobylskij, sp. nov.

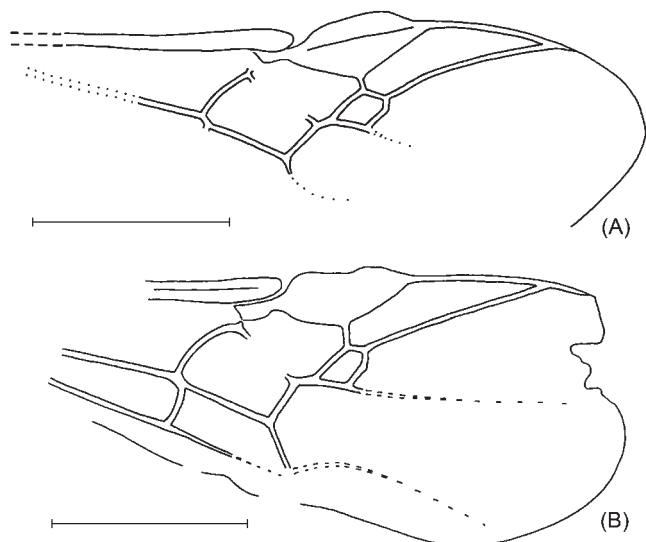
Plate 9, fig. 4; Text-fig. 26

**Etymology.** After P.B. Brodie, whose collection includes the holotype.

**Holotype.** NHMUK I.9239, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; wasp in dorsoventral aspect lacking antennae, legs, forewings, and metasomal apex.



**Plate 9** (1) *Bassus magnareola* sp. nov., holotype, NHMUK In.17127. (2) *Bassus quadrangularis* (Brues), NHMUK In.24958. (3) *Bracon* ? *antefurcalis* sp. nov., holotype, NHMUK I.9061. (4) *Eubazus* ? *brodiei* sp. nov., holotype, NHMUK I.9239. Scale bars = 1 mm.



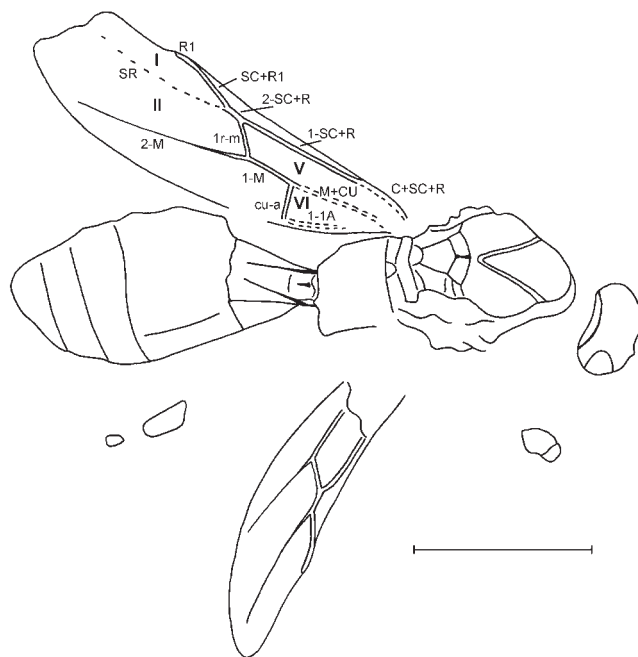
**Text-figure 25** (A) *Bassus magnareola* sp. nov., holotype, NHMUK In.17127. (B) *B. quadrangularis* (Brues), NHMUK In.24958. Scale bars = 1 mm.

**Diagnosis.** Characteristic of the species is the propodeum lacking delineated areas and the first tergite long and with high dorsal carinae.

**Description.** Length of body 3.8 mm, mesosoma 1.5 mm, metasoma 1.7 mm. Head transverse, about twice as wide as median length, rather strongly and somewhat roundly narrowed behind the eyes. Occipital carina complete dorsally. Mesosoma about 2.5 times as long as maximum width. Median lobe of mesoscutum distinctly protruding forwards and rounded anteriorly. Notauli complete, rather deep, more or less narrow entirely, fused one with another shortly before prescutellar depression. Prescutellar depression wide, deep, with high medial carina, possibly weakly sculptured. Scutellum convex. Metanotum medially with two fine carinae convergent posteriorly, and with high and rather large tubercle in posterior half. Propodeum long, irregularly rugose-reticulate, possible smooth basilaterally. Hind wing with sub basal cell large; M+CU 3.0 times as long as 1-M. Basal cell rather narrow, almost seven times as long as its maximum width. Marginal cell weakly widened in basal half and then weakly narrowed towards apex. Metasoma 1.1 times as long as mesosoma. First tergite elongate, regularly and not strongly widened towards apex, with high and weakly convergent dorsal carinae developed in basal 0.8 of tergite, dorsope small or almost indistinct; length of tergite 1.5 times its apical width, apical width 1.6 times its basal width. Second suture absent. Combined median length of second and third tergites 1.4 times basal width of second tergite and 0.8 times their maximum width. Following segments distinctly protruding behind third one. Colour: mesosoma and first metasomal tergite black; rest of metasomal tergites possibly brown; hind wing hyaline; veins brown.

**Remarks.** The absence of forewings is a serious problem for understanding the real taxonomic position of this fossil because similar morphological structures are known in subfamilies Brachistinae and Helconinae (Diospilini). On the base of hind wing venation, structures of the dorsal part of mesosoma and the first metasomal tergite, the new species is included tentatively in the genus *Eubazus*.

*Eubazus flavistigma* Belokobylskij, sp. nov.  
Plate 10, fig. 1; Text-fig. 27A



**Text-figure 26** *Eubazus ? brodiei* sp. nov., holotype, NHMUK I.9239. Scale bars = 1 mm.

**Etymology.** From flavus, the Latin for yellow, and pterostigma, referring to its colouration.

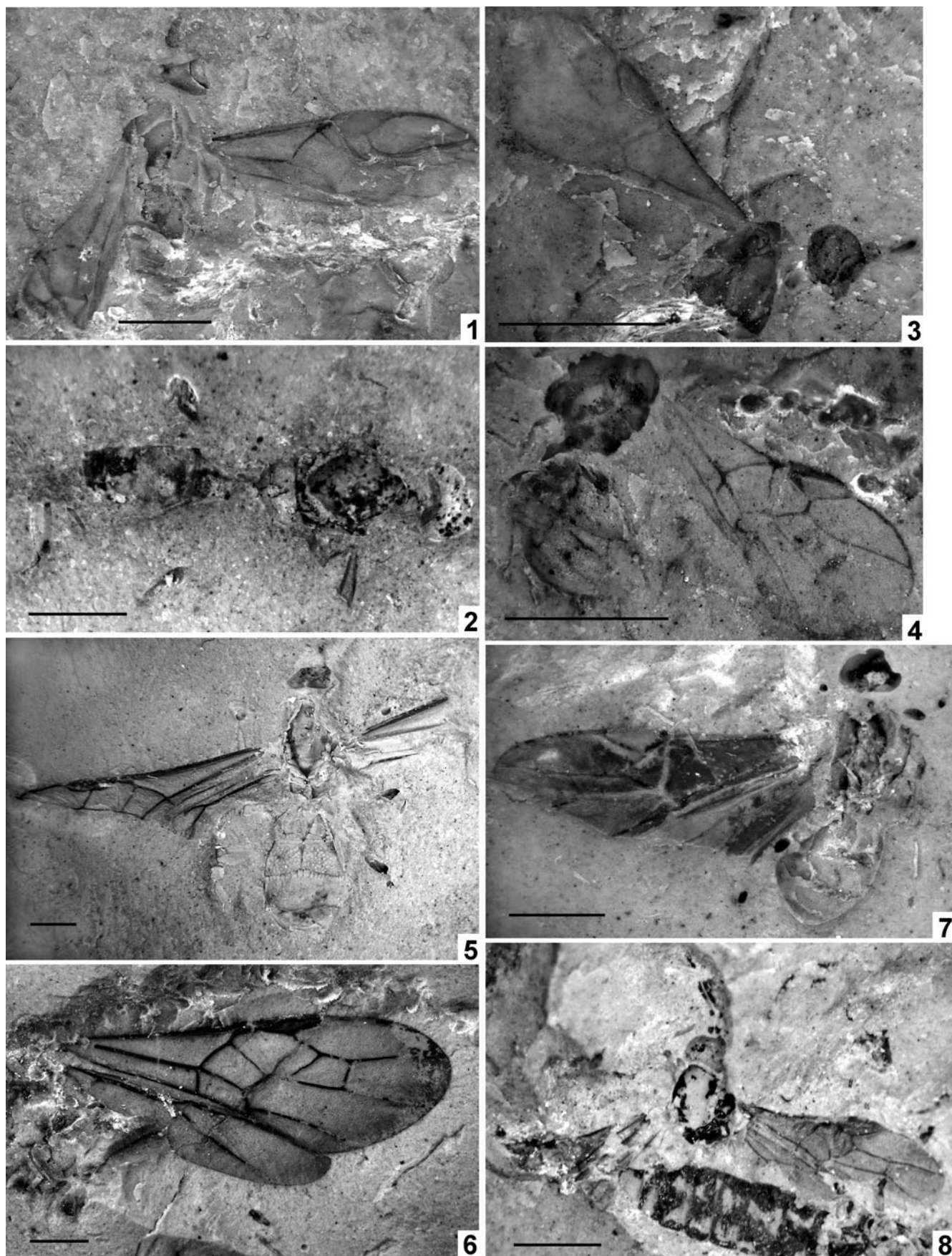
**Holotype.** NHMUK In.17244, Bembridge Marls, NW Isle of Wight, UK, Smith Collection; incomplete fossil showing head in dorsal view, with antennal bases, mesosoma in oblique dorsal-lateral view, and forewings, one lacking base and another without fore and distal parts.

**Diagnosis.** The present species differs distinctly from the all other fossil *Eubazus* in yellow coloration of pterostigma.

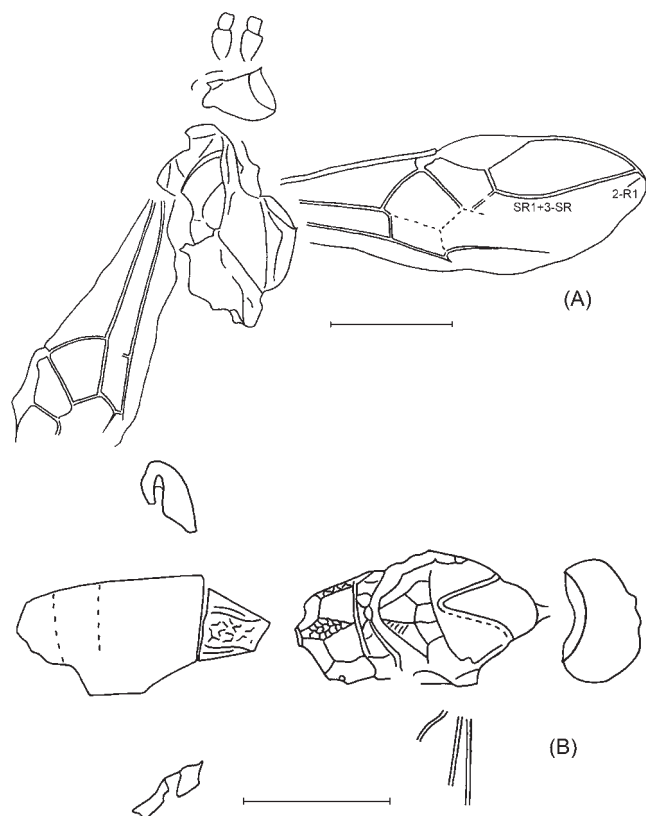
**Description.** Length of mesosoma 1.6 mm, forewing 3.0 mm. Head behind eyes rather strongly and roundly narrowed. Scapus thick, 1.4 times as long as wide, almost twice as long as pedicel. Mesosoma 1.5 times as long as maximum height. Mesoscutum highly and roundly elevated above pronotum. Notauli complete, very deep, wide anteriorly and narrow posteriorly. Prepectal carina distinct. Sternauli deep and wide medially, smooth, situated in anterior 0.8 of mesopleuron. Propodeum possibly without areolation. Forewing 2.6 times as long as maximum width. Pterostigma subtriangular, wide, 2.6 times as long as maximum width. 1-R1 1.25 times as long as pterostigma, 2-R1 distinct. Marginal cell weakly shortened, pointed apically, 2.6 times as long as maximum width; r arising a little behind midlength of pterostigma, 0.7 times as long as maximum pterostigmal width. SR1+3-SR distinctly curved in basal 0.3, otherwise almost straight, 5.8 times as long as r, 3.8 times as long as 2-SR. 2-SR 1.5 times as long as r, 1.2 times as long as m-cu. 1-SR+M straight. Discal cell sessile anteriorly, 1.5 times as long as wide; m-cu distinctly antefurcal; cu-a postfurcal, distant from 1-M for half cu-a length. Subdiscal cell wide, closed antero-posteriorly by CU1b, 2.5 times as long as wide. CU1a arising from posterior 0.25 of distal side of subdiscal cell. Colour: head and possibly mesosoma pale brown; wings faintly infusate; pterostigma entirely yellow; veins brown.

*Eubazus ? grandareola* Belokobylskij, sp. nov.  
Plate 10, fig. 2; Text-fig. 27B

**Etymology.** After *grandis*, the Latin for large, and areola, referring to the propodeum with large areola.



**Plate 10** (1) *Eubazus flavistigma* sp. nov., holotype, NHMUK In.17244. (2) *Eubazus* ? *grandareola* sp. nov., holotype, NHMUK I.8754. (3) *Eubazus* ? *hooleyi* sp. nov., holotype, NHMUK In.24790. (4) *Eubazus nanus* sp. nov., holotype, NHMUK I.10108. (5) *Bembracon acourtsmithi* sp. nov., holotype, NHMUK In.17224. (6) *Bembracon medialis* sp. nov., holotype, NHMUK Pl II.2760. (7) *Bracon brodiei* sp. nov., holotype, NHMUK I.9351. (8) *Bracon micrarche* (Cockerell), holotype, NHMUK I.9026. Scale bars = 1 mm.

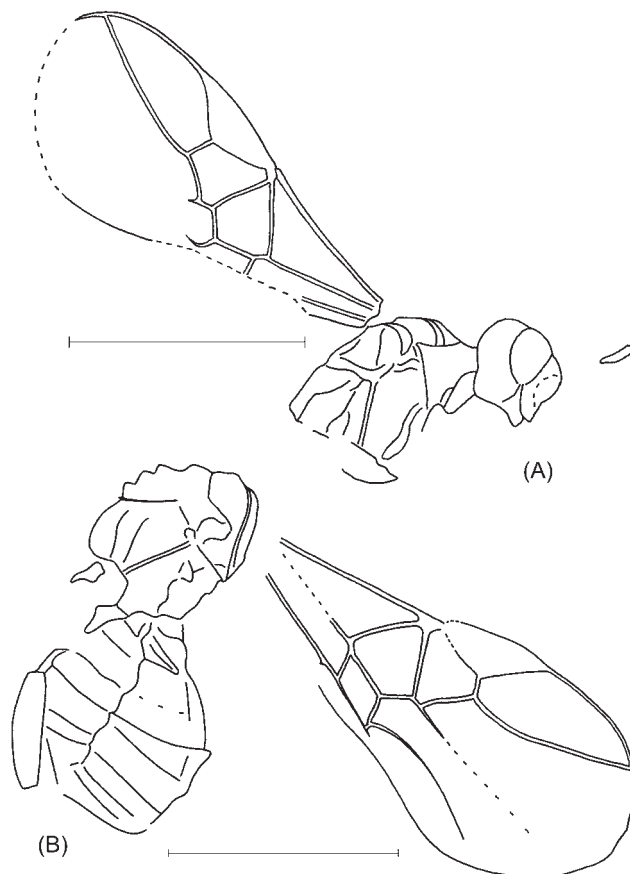


**Text-figure 27** (A) *Eubazus flavistigma* sp. nov., holotype, NHMUK In.17244. (B) *E. ? grandareola* sp. nov., holotype, NHMUK I.8754. Scale bars = 1 mm.

**Holotype.** NHMUK I.8754, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; body in dorsoventral view, lacking antennae, wings, almost all legs, and rear part of metasoma.

**Diagnosis.** The present species is very similar to *Charmon*, but differs in presence of sternauli. It differs distinctly from the all other fossil *Eubazus* in its short temple and large propodeal areola.

**Description.** Length of body 4.5 mm (mesosoma 1.7 mm, metasoma 1.7 mm). Head transverse, about twice as wide as median length. Occipital carina dorsally complete. Mesosoma 2.3 times as long as maximum width. Mesoscutum 1.15 times as long as maximum width. Median lobe of mesoscutum distinctly protruding forwards and rounded anteriorly. Notauli complete, rather deep, more or less wide throughout, crenulated, fused shortly before prescutellar depression. Prescutellar depression wide, deep posteriorly, with high median carina, possibly weakly sculptured. Scutellum convex, without transverse posterior depression. Metanotum medial with two fine carinae weakly convergent posteriorly, and with high and rather large tubercle in posterior half. Propodeum long, possibly finely and irregularly rugose-reticulate at least in part, with distinct areolation, areola large, long subrhomboid, distinctly narrowed posteriorly, 2.6 times as long as maximum width, with very short basal carina; petiolar area short and semicircular, distinctly separated by carina from areola. Metasoma: First tergite relatively short, regularly and strongly widened towards apex, with high and weakly convergent dorsal carinae in basal half of tergite, with strongly convex and wide median area; length of tergite 0.9 times its apical width, apical width about 2.5 times its basal width. Second suture absent. Combined median length of second and third tergites 1.2 times basal width of second tergite and 0.9 times their maximum



**Text-figure 28** (A) *Eubazus ? hooleyi* sp. nov., holotype, NHMUK In.24790. (B) *E. nanus* sp. nov., holotype, NHMUK I.10108. Scale bars = 1 mm.

width. Following segments distinctly protruding behind third one. Colour: head and mesosoma black; metasoma brown, dark brown posteriorly.

**Remarks.** Characteristic of the species is the propodeum, with its rather large and long areola which almost reaches the basal margin of the propodeum. In the absence of information about wing morphology, the species is tentatively attributed to *Eubazus* for the same reasons as *E. brodiei* sp. nov.

*Eubazus ? hooleyi* Belokobylskij, sp. nov.

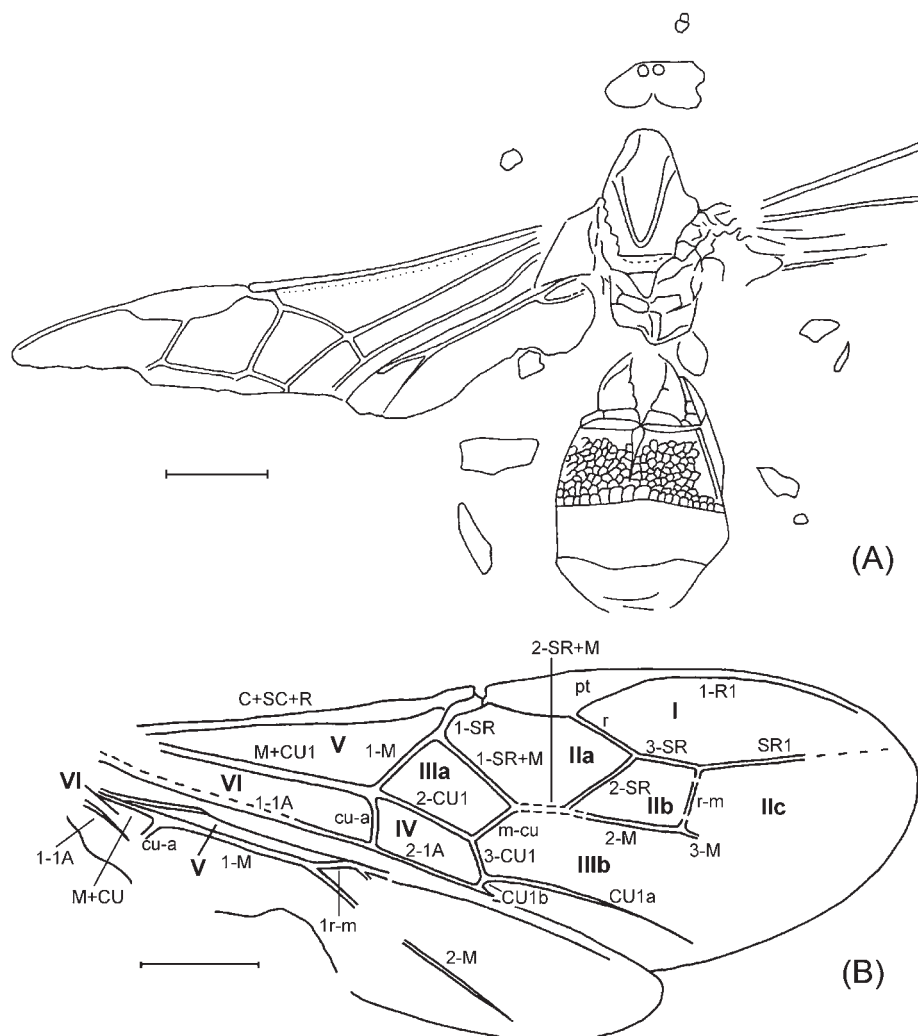
Plate 10, fig. 3; Text-fig. 28A

**Etymology.** After R.W. Hooley, who compiled the collection including the holotype.

**Holotype.** NHMUK In.24790, Bembridge Marls, NW Isle of Wight, UK, Hooley Collection; head and incomplete mesosoma in side view, and one forewing lacking part of hind and apical margin.

**Diagnosis.** The main diagnostic characters of this species are long temple, deep and S-shaped sternauli, distinctly shortened marginal cell, and long 2-R1.

**Description.** Length of head 0.35 mm, mesosoma 0.8 mm, forewing 1.9 mm. Head 1.2 times as high as long. Temple rather long, eye almost as long as temple. Occipital carina present at least laterally. Clypeus convex. Mesosoma high, 1.35 times as long as maximum height. Mesoscutum high, roundly elevated above pronotum. Notauli apparently present. Scutellum weakly convex. Subalar depression rather deep and narrow. Prepectal carina present. Sternauli deep, rather narrow, S-shaped, oblique, long, almost smooth. Forewing 2.3 times as long as wide. Pterostigma subtriangular, wide, 2.8 times as



**Text-figure 29** (A) *Bembracon acourtsmithi* sp. nov., holotype, NHMUK In.17224. (B) *B. medialis* sp. nov., holotype, NHMUK P 1.II2760. Cells: I = marginal; II = submarginal; III = discal; IV = subdiscal; V = basal; VI = sub basal; a, b, and c indicate first, second and third cell, respectively. Scale bars = 1 mm.

long as maximum width, almost as long as 1-R1. Marginal cell distinctly shortened, 2.7 times as long as maximum width. 2-R1 rather long; r arising from middle of pterostigma, 0.6 times as long as maximum pterostigmal width. SR1+3-SR weakly curved anteriorly and almost straight posteriorly, c. 6.0 times as long as r. 2-SR twice as long as r, 4.7 times as long as 2-SR+M, 1.6 times as long as m-cu. 1-SR+M straight. Discal cell shortly sessile anteriorly, almost as long as wide; cu-a postfurcal, distant of 1-M for 0.6 times cu-a length. Colour: head black; wings hyaline; pterostigma (possibly) and veins brown.

**Remarks.** Wing venation, particularly the shape of SR1+3-SR, and presence of sternauli and notauli, suggest attribution of the species to subfamily Brachistinae. The absence of the metasoma does not permit certain identification to genus, so I ascribe it tentatively to *Eubazus*.

*Eubazus nanus* Belokobylskij, sp. nov.

Plate 10, fig. 4; Text-fig. 28B

**Etymology.** After *nanus*, the Greek for dwarf, referring to the very small body size.

**Holotype.** NHMUK I.10108, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; meso- and metasoma in side view with hind femur and one forewing.

**Diagnosis.** The main diagnostic characters of this new taxon are sternauli shallow, propodeum without delineated areas, forewing short, discal cell narrow, and marginal cell shortened.

**Description.** Length of metasoma 0.9 mm, forewing 1.9 mm. Mesosoma c. 1.2 times as long as maximum height. Mesoscutum highly and roundly elevated above pronotum. Notauli complete, deep and rather narrow in anterior 0.7, deeper and rather wide in posterior 0.3, fused just before prescutellar depression. Prescutellar depression rather wide, deep, finely sculptured at least partly, with distinct median keel. Prepectal carina present laterally, possibly very fine ventrally. Sternauli very shallow, wide, almost smooth, situated along almost whole length of lower part of mesopleuron. Metapleural suture distinctly and shortly crenulated. Propodeum short, almost not narrow in anterior 0.3 (lateral view) and strongly and almost linearly abruptly in posterior 0.7, without marginal areas, almost entirely and rather finely rugose-reticulate. Forewing 2.2 times as long as maximum width. Pterostigma subtriangular, wide, 2.5 times as long as maximum width. 1-R1 1.1 times as long as pterostigma; 2-R1 short. Marginal cell shortened, pointed apically, 2.8 times as long as maximum width; r arising almost from middle of pterostigma, 0.45 times as long as maximum pterostigmal width. SR1+3-SR weakly and evenly curved in basal 0.5 and weakly S-shaped in apical 0.5, 8.3 times as long as r, 3.5 times as long

as 2-SR. 2-SR 2.4 times as long as r, 1.35 times as long as m-cu. 1-SR+M almost straight. Discal cell shortly sessile anteriorly, 1.1 times as long as wide; m-cu distinctly antefurcal, 3.3 times as long as 2-SR+M; cu-a postfurcal, distant from 1-M for half cu-a length. Subdiscal cell wide, closed antero-posteriorly by CU1b, 2.4 times as long as wide. CU1a arising from posterior 0.25 of distal side of subdiscal cell, a present and unsclerotised. Hind femur about 4.0 times as long as maximum width. Metasoma not forming carapace, tergites rather soft, apical segments distinctly protruding behind third tergite. Second suture very fine. Second tergite almost as long as third tergite. Colour: mesosoma and possibly metasoma almost entirely dark brown; wing hyaline; entire pterostigma and veins brown.

**Remarks.** The position of this species in the genus *Eubazus* is clear from the venation of the forewing, the mesosoma with prepectal carina and sternaui, and the structure of the metasoma. The new species is very similar to *E. crassicornis* (Brues) described from Baltic amber (Brues 1933) and redescribed by Achterberg (1982) in having the widely sessile discal cell of the forewing, the short first metasomal tergite and the wide hind femur. Unfortunately, the supporting characters of the fossil are not numerous, which hinders the comparison of the taxa, but *E. nanus* sp. nov. possibly differs in having the subdiscal cell wide and short, 2-SR long, first and second metasomal tergites short, and forewing short. *E. nanus* sp. nov. differs from *E. hooleyi* sp. nov. in having the discal cell short and narrow sessile anteriorly, 3-SR mostly straight, pterostigma and sternaui narrow; from *E. flavistigma* sp. nov. in having pterostigma dark coloured, discal cell narrow and widely sessile, and r short.

**3.9.3. Subfamily Braconinae Nees, 1812.** The subfamily Braconinae is one of the largest and most diverse groups of idiobiont braconids, with numerous extant genera described from tropical and subtropical regions (Quicke 1987). Used for designation of these genera, the special characters of antennae, mesosomal and metasomal tergites are usually not visible in rock fossils. This presents problems in the identification of the taxonomic position of these fossils.

Only a few Cenozoic Braconinae have been described so far. Their generic names have changed with time; I prefer to include the majority of described taxa, especially those described by old authors, to the polymorphic genus *Bracon* used in the broad sense, until future revisions have been undertaken. These are *Bracon cockerelli* Brues, 1910 (Florissant, USA), *B. diffusiventris* Théobald, 1937 (Oligocene of Kleinkembs, Germany), *B. laceolatus* Zhang, 1989 (Miocene of Shangwang, China), *B. macrostigma* Heyden, 1858 (Oligocene of Sieblos, Germany), *B. praeteritus* Foerster, 1891 (Oligocene of Brunstatt, Germany), *B. rottensis* Meunier, 1915 (Rott, Germany), possibly also *B. abstractus* Brues, 1910 and *B. resurrectus* Brues, 1910 from Florissant, as well as *B. micrarche* (Cockerell, 1921), comb. nov., from the Bembridge Marls. The status of *Bracon laminarium* Scudder, 1878 from Green River (USA) is unclear and perhaps it does not belong to the Braconidae (A. P. Rasnitsyn, pers. comm. 2006). According to the figure, *Bracon pallidus* Heer, 1867 is not of the subfamily Braconinae, but its correct taxonomic position is obscure and requires re-study of the type. Unfortunately, most of the aforementioned species of this genus described in the 19th or beginning of the 20th centuries have no distinct descriptions with correct modern diagnostic features, and it is quite possible some of these species belong to other genera or subfamilies. There is only one braconine species described from a rock fossil that can be identified with greater confidence, *Iprobracon vectensis* Cockerell, 1921a, apparently belonging to the genus *Cyanopterus* (see below).

Worth mentioning is that all Bembridge Marls Braconinae, except those attributed to a new genus, have distinctly curved 1-SR+M.

Genus *Bembracon* Belokobylskij, gen. nov.

**Etymology.** After Bembridge and genus *Bracon*. Gender masculine.

**Type species.** *Bembracon acourtsmithi* sp. nov., by present designation.

**Diagnosis.** Differs from extant genera in the combination of the following characters: notauli distinct and complete; pre-scutellar depression wide and deep; first tergite with median area distinctly and acutely narrowed towards apex; and with apico-lateral transverse areas delineated by carina.

**Description.** Head with antennal sockets separated by only half their diameter. Mesoscutum long, with notauli distinct, complete and anteriorly deep. Prescutellar depression wide, oblique anteriorly, almost vertical posteriorly. Pterostigma rather narrow; r arising from middle of pterostigma; r-m present. Discal cell distinctly petiolate anteriorly; cu-a interstitial or a little antefurcal; m-cu distinctly antefurcal. Second sub-marginal cell short and wide. Subdiscal cell closed by CU1b in posterodistal corner. Hind wing with basal cell very narrow and sub basal cell very small. First metasomal tergite with median area wide, pointedly narrowed posteriorly, and with two apicolateral transverse areas delineated by carinae. Second tergite moveably connected with first tergite, with very narrow and long triangle median area following to second suture. Second suture deep and wide, coarsely crenulated.

**Remarks.** The genus is tentatively included in the subfamily Braconinae, because the studied fossils present no information about the occipital and prepectal carinae. At the same time, the new genus shows unique characters, unknown for Braconinae, in the structure of the first metasomal tergite, which has a pointedly narrowed posterior median area and delimited apico-lateral transverse areas. These characters, taken together with the distinct and complete notauli and deep and the wide prescutellar depression, might allow proposing a separate tribe or even subfamily for the new genus. However, this seems premature until more information is accumulated for this interesting group.

*Bembracon acourtsmithi* Belokobylskij, sp. nov.

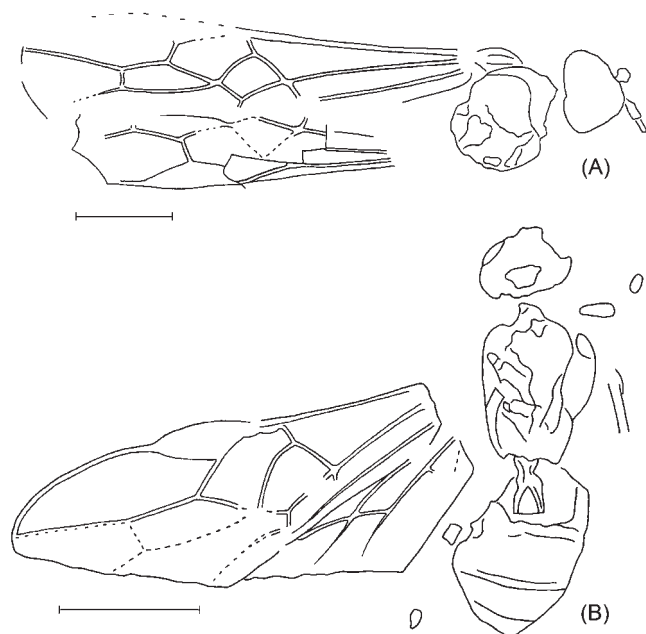
Plate 10, fig. 5; Text-fig. 29A

**Etymology.** After the collector's name.

**Holotype.** NHMUK In.17224, Bembridge Marls, NW Isle of Wight, UK, Smith Collection; head (damaged) and body in dorsal aspect with incomplete wings (missing hind and apical areas) and small leg fragments.

**Diagnosis.** Differs from the second species of this genus, *B. medialis* sp. nov., in having R and particularly 2-SR+M short, and 1-SR short.

**Description.** Length of metasoma 2.7 mm. Mesosoma: Mesoscutum long, 1.4 times as long as maximum width. Notauli present, complete, shallow. Prescutellar depression almost smooth. Propodeum smooth, possibly with complete high longitudinal keel. Forewing: pterostigma oval triangular, c.4.0 times as long as maximum width; r arising from middle of pterostigma and distinctly oblique in respect to it, 1.6 times as long as maximum pterostigmal width, 0.65 times as long as 2-SR. 1-SR+M almost straight; m-cu 0.7 times as long as 2-SR, 0.6 times as long as 1-SR+M and 1-M. Discal cell 1.7 times as long as wide. 1-M about 5.0 times as long as 1-SR; m-cu distinctly antefurcal. Subdiscal cell wide. Hind wing:



**Text-figure 30** (A) *Bracon ? antefurcalis* sp. nov., holotype, NHMUK I.9061. (B) *B. brodiei* sp. nov., holotype, NHMUK I. 9351. Scale bars = 1 mm.

m-cu absent. Basal cell subparallel-sided. M+CU 0.4 times as long as 1-M. Legs: hind femur wide. Metasoma 1.65 times as long as maximum width. First tergite wide, 0.75 times as long as wide apically, with median area subtriangular (2.4 times as long as maximum width), smooth except narrowly areolate laterally. Second tergite almost entirely rather densely and sub-roundly areolate; median length of tergite (without suture) 0.5 times its basal width, 0.8 times length of third tergite. Second suture rather narrow laterally and wide medially, weakly curved. Third tergite entirely smooth. Posterior tergites shortly protruding behind third one. Colour: wings entirely infusate; pterostigma and veins black.

*Bembracon medialis* Belokobylskij, sp. nov.

Plate 10, fig. 6; Text-fig. 29B

**Etymology.** After medial vein, which is distinctive in the length of its 2-SR+M.

**Holotype.** Pl II2760a, b (part and counterpart), north end of Thorness Bay. Collected by R. Szadziewski, 23/05/05; joined fore and hind wings.

**Diagnosis.** Differs from the type species in having r and particularly 2-SR+M long, 1-SR long, and the discal cell less widened basally.

**Description.** Length of forewing 7.3 mm, hind wing 5.2 mm. Forewing length 2.7 times its maximum width. Pterostigma narrow, oval-triangular, 4.3 times as long as maximum width; r arising weakly before middle of pterostigma and distinctly oblique in respect to it, 1.85 times as long as maximum pterostigmal width, 0.9 times as long as 2-SR. Marginal cell not shortened; 1-R1 1.25 times as long as pterostigma. 3-SR 0.85 times as long as r, 0.3 times as long as SR1, 0.75 times as long as 2-SR, 1.1 times as long as r-m. 2-SR 1.45 times as long as r-m; latter with two bullae. Second submarginal cell short, 1.8 times as long as maximum width, almost as long as discal cell. 1-SR+M straight. 2-SR+M long, 0.6 times as long as 2-SR, 0.9 times as long as m-cu; m-cu 0.7 times as long as 2-SR, 0.5 times as long as 1-SR+M and 0.7 times as long as 1-M. Discal cell 1.85 times as long as wide. 1-M c. 3.0 times

as long as 1-SR; m-cu strongly antefurcal; cu-a very weakly antefurcal. Subdiscal cell wide, 2.5 times as long as wide. Hind wing: m-cu absent. Basal cell very narrow and subparallel-sided. M+CU 0.45 times as long as 1-M.

Genus *Bracon* Fabricius, 1804

*Bracon ? antefurcalis* Belokobylskij, sp. nov.

Plate 9, fig. 3; Text-fig. 30A

**Etymology.** After the antefurcal position of the forewing cu-a.

**Holotype.** NHMUK I.9061, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; poorly preserved head with scape and pedicel and partial mesosoma in lateral view, two damaged forewings and badly damaged hind wing.

**Description.** Length of head 0.6 mm, of forewing 4.6 mm. Head 1.3 times as high as long. Face rather convex and short. Hypoclypeal cavity rather large. Occipital carina absent. Antenna: scape rather wide and short; pedicel possibly very short. Mesosoma possibly with prepectal carina absent. Forewing: pterostigma subtriangular, rather narrow, 3.8 times as long as maximum width. Marginal cell not shortened, pointed apically, almost 3.0 times as long as maximum width. 1-R1 c. 1.2 times as long as pterostigma; r arising almost from middle of pterostigma, 0.9 times as long as maximum pterostigmal width, forming distinct obtuse angle with 3-SR. 3-SR twice as long as r, 0.5 times as long as the almost straight SR1, almost as long as 2-SR; r-m present and distinct, 0.35 times as long as 2-SR. Second submarginal cell 3.2 times as long as width, about 1.8 times as long as subdiscal cell. 1-SR+M straight basally, curved in apical half; m-cu distinctly antefurcal, 3.7 times as long as 2-SR+M, 0.6 times as long as 2-SR. Discal cell long petiolate anteriorly, 1.8 times as long as width. 1-SR almost 0.5 times as long as 1-M; cu-a distinctly antefurcal, distant from 1-M for 0.35 times cu-a length. Subdiscal cell wide. Hind wing with sub basal cell short, m-cu possibly absent. Colour: head possibly brown; mesosoma brown; wings almost hyaline; pterostigma brown, veins dark brown.

**Remarks.** I tentatively include this new species in the genus *Bracon* until additional information about its morphology is known. The most important characters of this taxon are the distinctly antefurcal position of cu-a, the equal lengths of 3-SR and 2-SR, the relatively narrow pterostigma and the second submarginal cell. Antefurcal position of cu-a is known for braconine species of the genus *Eubrocon* Ashmead, 1900, but the new species distinctly differs from *Eubrocon*, besides its small body size, in the different shape and large size of the second submarginal cell.

*Bracon brodiei* Belokobylskij, sp. nov.

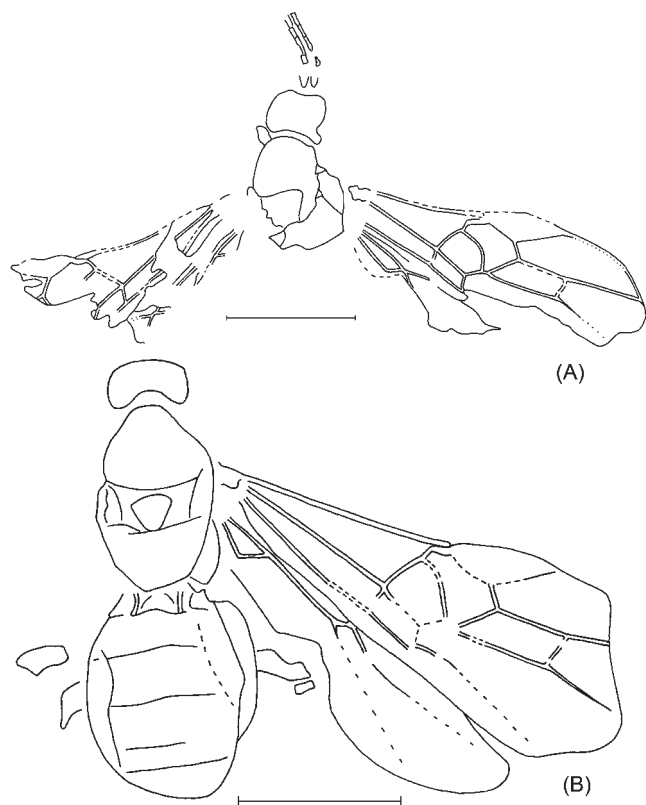
Plate 10, fig. 7; Text-fig. 30B

**Etymology.** After P.B. Brodie who compiled the collection which includes the holotype.

**Holotype.** NHMUK I.9351, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; dorsal aspect of poorly preserved head and mesosoma and better preserved metasoma, incomplete wings (lacking base and posteroapical area) and leg fragments.

**Diagnosis.** Very similar to *B. micrarche* (Cockerell, 1921), but differs in having r and the second submarginal cell long, and the 1-SR short. Additional material is necessary to decide if the above difference is really of specific importance.

**Description.** Length of body c.3.0 mm, mesosoma c.1.1 mm, forewing 3.2 mm. Head more or less transverse, distinctly and convex-roundly narrowed behind eyes. Antennal



**Text-figure 31** *Bracon micrarche* (Cockerell): (A) holotype NHMUK I.9026/I.9760; (B) NHMUK I.9182. Scale bars = 1 mm.

sockets large, separated for almost their diameters and  $c.1.8$  times distance between socket and eye. Transverse diameter of eye  $1.6$  times length of temple. Ocelli weakly enlarged. Mesosoma  $1.4$  times as long as maximum width. Forewing  $2.8$  times as long as maximum width. Pterostigma subtriangular, wide,  $2.8$  times as long as maximum width.  $1-R1$   $1.4$  times as long as pterostigma;  $r$  arising from basal  $0.4$  of pterostigma and oblique in respect to it, almost as long as maximum pterostigmal width. Marginal cell not shortened, pointed apically,  $3.4$  times as long as maximum width.  $3-SR$  about twice as long as  $r$ ,  $0.75$  times as long as  $SR1$ ,  $1.5$  times as long as  $2-SR$ ;  $r-m$  possibly sclerotised medially and with two short bullae subanteriorly and subposteriorly,  $0.6$  times as long as  $2-SR$ . Second submarginal cell long,  $3.4$  times as long as wide,  $1.7$  times as long as subdiscal cell.  $1-SR+M$  strongly curved in basal half and almost straight in apical half;  $m-cu$  antefurcal,  $0.6$  times as long as  $2-SR$ ,  $0.9$  times as long as  $r$ ,  $0.3$  times as long as  $2-M$ . Discal cell distinctly petiolate anterior,  $1.6$  times as long as wide.  $1-M$  almost  $3.0$  times as long as  $1-SR$ ;  $cu-a$  interstitial. Subdiscal cell rather wide, closed antero-posteriorly by  $CU1b$ ,  $3.0$  times as long as wide.  $CU1a$  arising from posterior  $0.3$  of distal side of subdiscal cell. Hind wing:  $SR$  arising from  $2-SC+R$  very closely to  $1r-m$ . Basal cell weakly widened anteriorly. Marginal cell widened in basal half. Sub basal cell short;  $M+CU$   $0.6$  times as long as  $1-M$ . Metasoma: first tergite rather long and more or less narrow, its length about  $1.3$  times apical width, with distinct convex basal area separated by rather deep lateral furrows, moveably connected with second tergite. Colour: head and mesosoma black; metasoma light reddish brown; wings distinctly infusate, paler distally; pterostigma and veins dark brown.

*Bracon micrarche* (Cockerell, 1921), comb. nov.  
Plates 10, fig. 8, 11, fig. 1; Text-fig. 31

1921a *Ipobracon* (?) *micrarche* Cockerell, p. 17, fig. 19.

**Holotype.** NHMUK I.9026/I.9760 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; dorso-ventral view of head with incomplete antennae and damaged mesosoma, and damaged wings.

**Other material.** NHMUK I.9182, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; sublateral view of head and dorso-ventral aspect of body lacking appendages except slightly incomplete fore and hind wings and leg fragments.

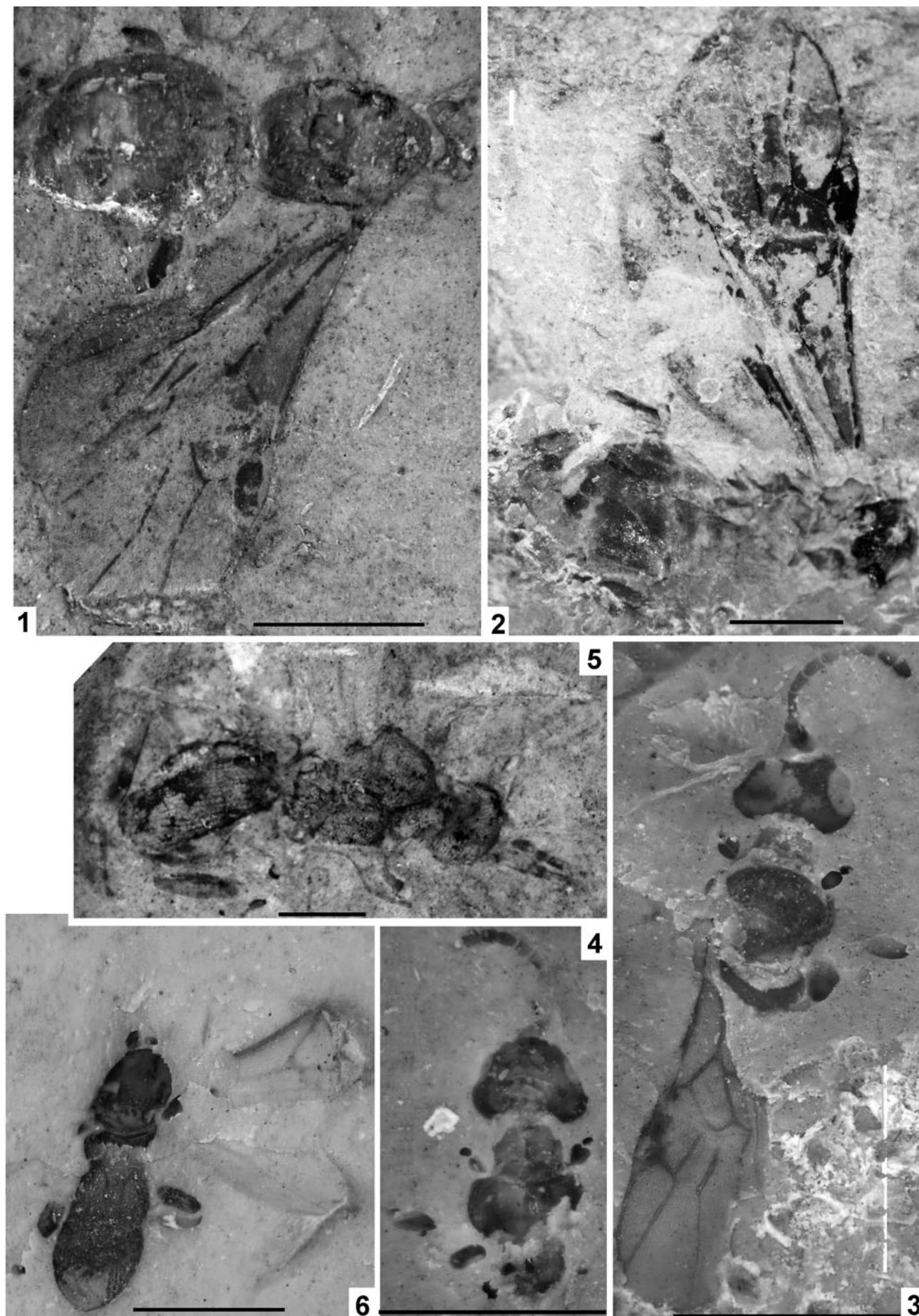
**Diagnosis.** Characteristic of this species is the distinctly curved  $1-SR+M$ . This species differs from *B. cockerelli* Brues, 1910 from Florissant in having the wings less strongly infusate,  $3-SR$  and second submarginal cell longer, and  $m-cu$  distinctly antefurcal.

**Description of holotype.** Length of mesosoma  $c. 0.8$  mm, forewing  $2.4$  mm. Head  $1.3$  times as high as maximum length. Occipital carina absent. Scape short and rather thick,  $1.3$  times as long as wide. Sub-basal segments  $2.8-3.0$  times as long as wide. Mesosoma: notauli absent. Prescutellar depression narrow. Scutellum wide, its width  $1.2$  times median length. Propodeum entirely smooth. Forewing  $c. 3.0$  times as long as maximum width. Pterostigma subtriangular, rather wide,  $2.8$  times as long as maximum width.  $1-R1$   $1.3$  times as long as pterostigma;  $r$  arising distinctly before middle of pterostigma (interior distance between parastigma and  $r$   $0.7$  times interior distance between  $r$  and apex of pterostigma), distinctly oblique in respect to pterostigma,  $0.6$  times as long as maximum pterostigmal width. Marginal cell not shortened, pointed apically,  $3.0$  times as long as maximum width.  $3-SR$   $3.2$  times as long as  $r$ ,  $0.75$  times as long as the almost straight  $SR1$ ,  $1.5$  times as long as  $2-SR$ ;  $r-m$  sclerotised, with two short bullae subanteriorly and subposteriorly,  $0.6$  times as long as  $2-SR$ . Second submarginal cell long, about  $3.0$  times as long as wide, about twice as long as rather wide subdiscal cell.  $1-SR+M$  more or less distinctly and rather strongly and evenly curved;  $m-cu$  antefurcal,  $0.5$  times as long as  $2-SR$ ,  $0.2$  times as long as  $2-M$ . Discal cell distinctly petiolate anteriorly,  $1.6$  times as long as wide.  $1-M$  about  $2.5$  times as long as  $1-SR$ ;  $cu-a$  almost interstitial. Subdiscal cell rather wide, closed antero-posteriorly by  $CU1b$ ,  $c. 3.0$  times as long as wide.  $CU1a$  arising from posterior  $0.3$  of distal side of subdiscal cell. Hind wing:  $m-cu$  absent. Basal cell widened anteriorly. Marginal cell widened in basal  $0.3$ , then weakly narrowed. Colour: head, antennae and mesosoma black; wings distinctly infusate, paler distally; pterostigma entirely brown; veins brown.

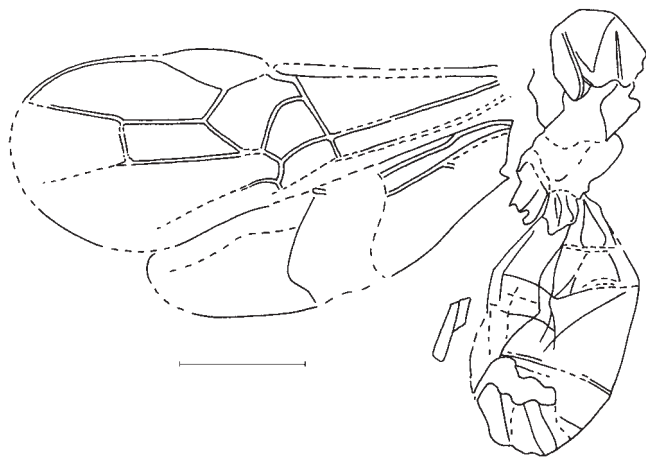
**Non-type specimen.** Generally similar to holotype, but differs in the following characters. Length of mesosoma  $c. 1.1$  mm, metasoma  $1.2$  mm, forewing  $c. 2.8$  mm. Mesosoma  $1.6$  times as long as maximum width. Mesoscutum  $0.85$  times as long as maximum width. Notauli absent. Width of scutellum  $1.3$  times its median length. Forewing  $c. 2.5$  times as long as maximum width as preserved. Pterostigma  $2.7$  times as long as maximum width.  $3-SR$   $2.9$  times as long as  $r$ ,  $1.6$  times as long as  $2-SR$ ;  $r-m$   $0.7$  times as long as  $2-SR$ . Second submarginal cell  $1.8$  times as long as subdiscal cell.  $1-SR+M$  rather strongly curved in basal half and almost straight in apical half;  $m-cu$   $0.25$  times as long as  $2-M$ . Discal cell  $1.4$  times as long as width. Subdiscal cell  $2.8$  times as long as wide. In hind wing, sub basal cell short,  $M+CU$   $0.6$  times as long as  $1-M$ . Metasoma  $c. 1.5$  times as long as wide. First tergite incompletely preserved, with median area distinct at least basally, moveably connected with second tergite. Second suture more or less distinct. Second tergite medially as long as third tergite. Metasoma light reddish brown.

Genus *Cyanopterus* Haliday, 1835  
*Cyanopterus vectensis* (Cockerell, 1921), comb. nov.  
Plate 11, fig. 2; Text-fig. 32

1921a *Ipobracon* (?) *vectensis* Cockerell, p. 17, fig. 18.



**Plate 11** (1) *Bracon micrarche* (Cockerell), NHMUK I.9182. (2) *Cyanopterus vectensis* (Cockerell), holotype, NHMUK I.9415. (3–4) *Ascogaster* (*Ascogaster*) *pygmaea* sp. nov., holotype, NHMUK In.17379: (3) part; (4) counterpart. (5) *Ascogaster* (*Syntaphus*) *wheeleri* (Donisthorpe), holotype, NHMUK I.8744. (6) *Ascogaster* (*Ascogaster*) *brodiei* sp. nov., holotype, NHMUK I. 9924. Scale bars = 1 mm.



**Text-figure 32** *Cyanopterus vectensis* (Cockerell), holotype NHMUK I.9415. Scale bar = 1 mm.

**Holotype.** NHMUK I.9415, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; headless body in sublateral view with incomplete wings.

**Diagnosis.** Differs from extant species in the strongly curved 1-SR+M in the forewing.

**Description.** Length of mesosoma 1.4 mm, metasoma 2.0 mm, forewing 3.9 mm. Mesosoma: mesoscutum possible finely and densely punctulate with very fine granulation on median lobe, with distinct, not deep, narrow notauli, fused posteriorly. Propodeum smooth throughout. Forewing 2.4 times as long as maximum width. Pterostigma subtriangular, wide, 2.7 times as long as maximum width. 1-R1 1.3 times as long as pterostigma; r arising distinctly before middle of pterostigma, distinctly oblique in respect to it, 0.9 times as long as maximum pterostigmal width. Marginal cell very weakly shortened, pointed apically, 3.1 times as long as maximum width. 3-SR 2.3 times as long as r, 0.9 times as long as the straight SR1, 1.4 times as long as 2-SR; r-m sclerotised, possibly with two short bullae subanteriorly and subposteriorly, 0.7 times as long as 2-SR. Second submarginal cell long, 3.3 times as long as wide, about twice as long as subdiscal cell. 1-SR+M strongly and somewhat irregularly curved. 2-SR+M about 0.8 times as long as m-cu, 0.2 times as long as 2-M. M-cu 0.4 times as long as 2-SR, 0.2 times as long as 2-M. Discal cell distinctly petiolate anteriorly, 1.45 times as long as wide. 1-M 2.5 times as long as 1-SR; m-cu weakly antefurcal; cu-a interstitial. Subdiscal cell rather wide, closed antero-posteriorly by CU1b, 2.4 times as long as wide. CU1a arising from posterior 0.3 of subdiscal cell; 3-CU1 distinctly curved. In hind wing, sub basal cell weakly widened anteriorly; marginal cell weakly narrowed submedially. Metasoma: first tergite with distinctly subtriangular median area delineated by rather deep furrows. Second tergite with rather distinct lateral furrows, oblique and divergent posteriorly. Second suture present. Colour: mesosoma black; metasoma pale brown; wings distinctly infusate; entire pterostigma and veins dark brown.

**Remarks.** This species belongs to *Cyanopterus* (senior synonym of *Ipobracon* Thomson, 1892) based on the presence of relatively distinct lateral divergent furrows on the second tergite combined with the absence of the median subtriangular area.

**3.9.4. Subfamily Cheloninae Foerster, 1862.** The subfamily Cheloninae is one of the most specialised groups within the microgasteroid phylogenetic branch of Braconidae. The members of this subfamily are easily recognised by the coarsely sclerotised and immoveably fused three anterior metasomal tergites, together with some features of forewing venation (such as r-m

present, marginal cell usually shortened, r often arising from the anterior third of pterostigma, m-cu usually postfurcal, 3-SR often short), basically reticulate-areolate sculpture of mesosoma and the presence of a postpectal carina.

The chelonine wasps are common as fossils, particularly as inclusions in the Eocene Baltic amber (Brues 1933; Tobias 1987). It is noteworthy that they mainly belong to the genus *Ascogaster*, one of the less derived groups in the subfamily (*A. adentata* Tobias, 1987, *A. dilatata* Brues, 1933, *A. gracilicornis* Brues, 1933, *A. longicauda* Tobias, 1987, *A. pentagona* Brues, 1933, *A. pinicola* Brues, 1933, *A. praevolans* Brues, 1933, *A. robusta* Brues, 1933, *A. rutilipes* Tobias, 1987, *A. submersa* Brues, 1933, *A. sylvestris* Brues, 1933, and *A. thoracica* Tobias, 1987). *Syntaphus wheeleri* Donisthorpe, 1920 described from the Bembridge Marls as an ant belongs here as well (see below). Moreover, all three species of the genus *Chelonus* Jurine, 1801 described from Florissant (*Chelonus depressus* Brues, 1910, *C. muratus* Brues, 1910 and *C. solidus* Brues, 1910) show no reliable features that would discriminate *Chelonus* from *Ascogaster*. Hence, no reliable records of the fossil *Chelonus* are known so far.

Only two species of *Phanerotoma* Wesmael, 1838 (*P. baltica*, Brues, 1933 and *P. extensa* Brues, 1933) have been recorded from Baltic amber (Brues 1933). The genus *Diodontogaster* Brues, 1933 (type species *D. bidentata* Brues, 1933 in Baltic amber) is the third chelonine genus in the fossil record which is venationally similar to *Ascogaster* and differs from it only in the presence of a pair of conical lobes on the posteroventral sides of the carapace (Brues 1933). The genus *Anacanthobracon* Brues, 1933 (type species *A. femorator* Brues, 1933) was described in the subfamily Doryctinae (Brues 1939), but its position is debatable: the good published figure shows no hypoclypeal cavity characteristic of that subfamily. This feature, together with the structure of the metasoma and very specific forewing venation, indicates this genus belongs to the subfamily Cheloninae. Finally, the genus *Chelonohelcon* Brues, 1933, described originally in the subfamily Helconinae, has been recently transferred to Cheloninae in the monobasic tribe Chelonohelconini (Tobias 1987).

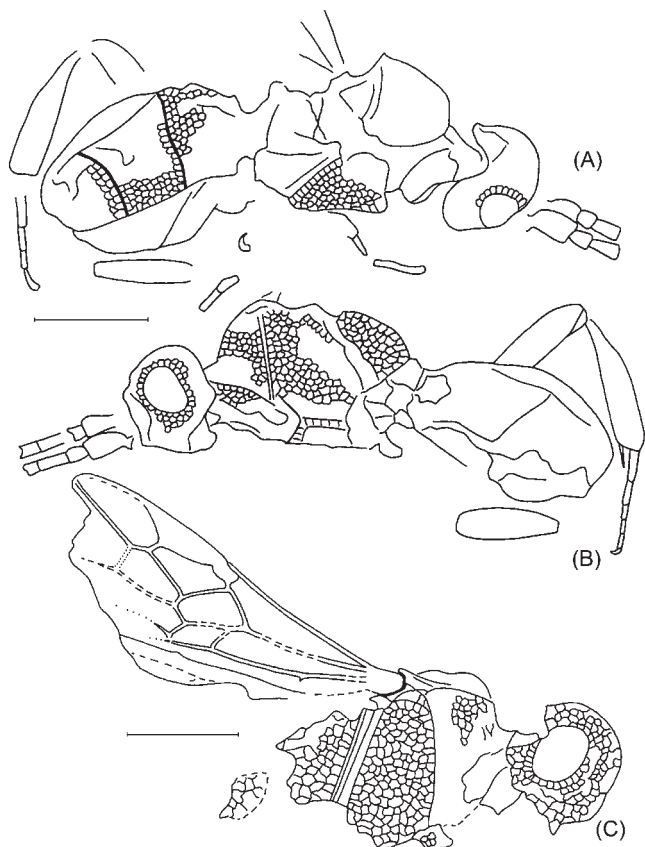
The position of the genus *Eobracon* Cockerell, 1920 (type species *E. cladurus* Cockerell, 1920a) described in the subfamily Cheloninae from the Eocene of Colorado (Cockerell 1920; Brues 1933) is also disputable. According to the wing venation (supposing that r-m is correctly shown there: see Cockerell 1920a, p. 259), this genus may be a specialised member of Helconinae.

Four genera and 20 species are known in the fossil record for Cheloninae. Three additional species of *Ascogaster* are described here. Worth mentioning is that all known fossil Cheloninae have a less derived forewing venation of the *Ascogaster* type (with 1-SR+M developed), supposing that the earlier authors (Gravenhorst 1834; Brischke 1886) have erroneously recorded *Chelonus* sp. in Baltic amber.

#### Genus *Ascogaster* Wesmael, 1835

**Remarks.** The species of *Ascogaster* are the most abundant fossil Cheloninae. They show striking variation of several important characters recorded in the described species, e.g., petiolate anterior to sessile basal cell and postfurcal vs. interstitial position of m-cu in the forewing (Brues 1933), or presence vs. absence of the first suture on the carapace (Tobias 1987). This suggests the need for care in assessing the real position of these taxa.

Development of both metasomal sutures is a previously unknown character for *Ascogaster*. The only species with clear



**Text-figure 33** *Ascogaster (Syntaphus) wheeleri* (Donisthorpe): (A, B) holotype, NHMUK I.8744: (A) part; (B) counterpart, sculpture partially drawn; (C) paratype, NHMUK I.9325. Scale bars = 1 mm.

sutures was described as an ant under the generic name *Syntaphus* Donisthorpe, 1920 (see below). Otherwise, a few extant and fossil species of *Ascogaster* have at most the first suture more or less developed (Tobias 1987). Nevertheless, I prefer to retain *Syntaphus* as a subgenus of *Ascogaster*.

The presence of two deep metasomal sutures is the main diagnostic character of the chelonine tribe Phanerotomini. The discovery of a similar character in *Ascogaster* reduces the gap between these genera, but does not close it. In fact, species of *Ascogaster* have the discal cell petiolate and body predominantly dark coloured, in contrast to *Phanerotoma* which have the discal cell distinctly and more-or-less widely sessile anteriorly and body colour basically pale. It is of importance that several species from Baltic amber described by Brues (1933) in *Ascogaster* (*A. dilatata* Brues, 1933, *A. gracilicornis* Brues, 1933, *A. praevolans* Brues, 1933, *A. pentagona* Brues, 1933, *A. sylvestris* Brues and *A. submerse* Brues) have the discal cell sessile or subsessile. However, his descriptions do not mention the presence or absence of metasomal sutures. All species described by Tobias (1987) have the discal cell distinctly petiolate.

Subgenus *Syntaphus* Donisthorpe, 1920, stat. nov.

**Type species.** *S. wheeleri* Donisthorpe, 1920, by monotypy.

**Diagnosis.** Differs from the nominate subgenus in having the metasomal carapace with two sutures developed.

*Ascogaster (Syntaphus) wheeleri* (Donisthorpe, 1920), comb. nov.

Plate 11, fig. 5; Text-fig. 33

1920 *Syntaphus wheeleri* Donisthorpe, p. 84, plate V. fig. 1, 2.

**Holotype.** NHMUK I.8744/I. 9936 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; practically wingless body in sublateral aspect with partial antenna, hind leg and small fragments of other legs.

**Paratype.** NHMUK I.9325, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; head and incomplete mesosoma in side view, and incomplete forewing.

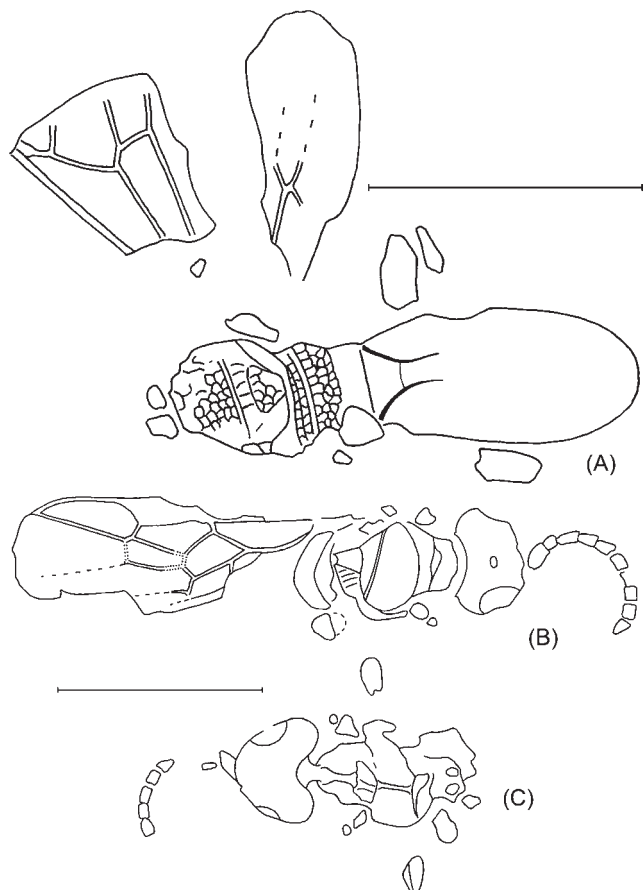
**Description of holotype.** Length of body 4.3 mm. Head sub-circular, entirely covered by coarse and irregularly areolate sculpture of chelonine type. Longitudinal diameter of eye 1.4 times its transverse diameter. Scape large, 2.0–2.3 times as long as wide, 2.7 times as long as pedicel. First flagellar segment 2.3–2.5 times as long as wide. Mesosoma 1.3–1.4 times as long as high, covered by chelonine type sculpture including propodeum but excluding apparently smooth mesosternum. Mesopleuron 1.6 times as high as maximum width. Notauli rather wide and shallow. Prescutellar depression rather shallow and short. Sternauli indistinct. Propodeum with rather long dorsal process (tooth) subacute apically. Hind femur rather wide, about 3.3 times as long as wide. Hind tibia strongly widened, 4.6 times as long as maximum width, 1.5 times as long as hind femur. Hind tarsus 0.8 times as long as hind tibia. Ratio of hind tarsal segments is 16:7:5:4:5. Carapace of metasoma with two rather distinct sutures, entirely covered by sculpture of chelonine type, 2.0–2.2 times as long as width. First and second tergites of equal median length and 1.1 times as long as third tergite. Colour: antenna black; mesosoma at least partly, and metasoma entirely, black; hind leg brown.

Paratype is generally similar to holotype, but differs in the following characters (the wing is absent in the holotype). Length of head and mesosoma (with spine of propodeum) 2.1 mm; mesosoma 1.4 mm. Longitudinal diameter of eye 1.3 times its transverse diameter. Mesopleuron about twice as high as maximum width. Propodeum with long dorsal sub-pointed apical tooth and short rather thick rounded ventral tubercle. Forewing (distal part absent): pterostigma subtriangular, wide, about 3.0 times as long as maximum width. 1-R1 0.9 times as long as pterostigma; r arising distinctly behind middle of pterostigma (internal distance of pterostigma between parastigma and r about twice internal distance between r and apex of pterostigma), distinctly oblique to pterostigma, 0.4 times as long as maximum pterostigmal width. Marginal cell distinctly shortened, pointed apically, 2.7 times as long as maximum width, 1.15 times as long as pterostigma. 3-SR 1.9 times as long as r, 0.25 times as long as weakly curved SR1, 0.4 times as long as 2-SR; r–m unsclerotised, 0.5 times as long as 2-SR. Second submarginal cell rather long, about twice as long as wide, 1.4 times as long as subdiscal cell. 1-SR+M weakly S-shaped. 2-SR+M 0.6 times as long as m–cu, 0.15 times as long as 2-M; m–cu 0.4 times as long as 2-SR, 0.3 times as long as 2-M. Discal cell shortly petiolate anteriorly, 1.85 times as long as wide. 1-M about 8.0 times as long as 1-SR; m–cu distinctly postfurcal; cu–a inclined posteroapically, distinctly postfurcal, distant from 1-M for 0.65 times cu–a length. Subdiscal cell wide, closed antero-posteriorly by CU1b, about twice as long as wide. CU1a arising from posterior 0.25 of subdiscal cell; 3-CU1 straight. 2A present and sclerotised at most part. Hind wing absent. Wings infusate; entire pterostigma and veins dark brown.

Subgenus *Ascogaster* Wesmael, 1835 s. str.

**Type species.** *A. instabilis* Wesmael, 1835

**Species included.** About 100 species are known in the world fauna, including 15 fossil ones (Brues 1933; Huddleston 1984; Tobias 1987, 2000).



**Text-figure 34** (A) *Ascogaster (Ascogaster) brodiei* sp. nov., holotype, NHMUK I.9924; (B, C) *Ascogaster (Ascogaster) pygmaea* sp. nov., holotype NHMUK In.17379: (B) part; (C) counterpart. Scale bars = 1 mm.

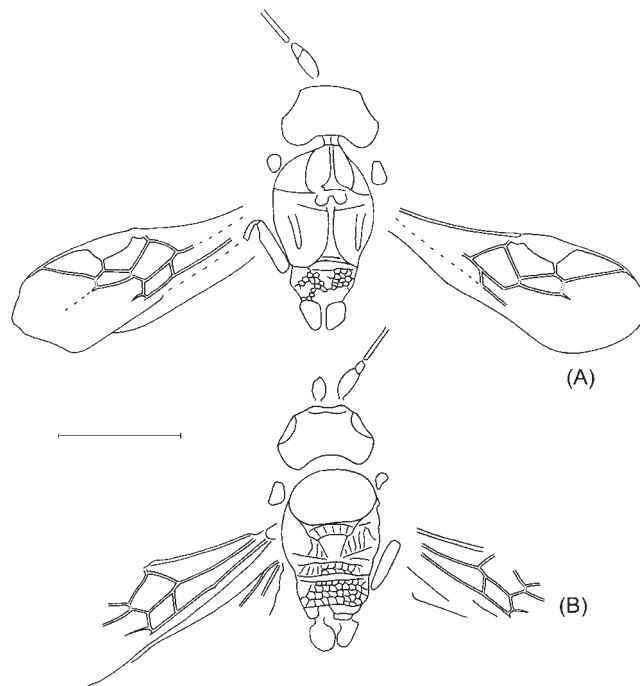
*Ascogaster (Ascogaster) brodiei* Belokobylskij, sp. nov.  
Plate 11, fig. 6; Text-fig. 34A

**Etymology.** After P.B. Brodie, who compiled the collection which includes the holotype.

**Holotype.** NHMUK I.9924, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; incomplete headless body in dorsoventral aspect, incomplete wings and leg fragments.

**Diagnosis.** The new species is similar to *A. robusta* Brues, 1933 with petiolate discal cell but differs in having the carapace long, narrow and apparently pale in basal part, and the propodeum without tubercles. The new species differs from *A. pinicola* Brues, 1933, another fossil species with a petiolate discal cell, in having notauli indistinct, scutellum areolate, carapace lacking longitudinal sculpture, and propodeum without a transverse carina. From all *Ascogaster* species described by Tobias (1987) in Baltic amber, this new species differs by the absence of the first metasomal suture. Comparison with described recent species is not possible because of the limited number of diagnostic characters available.

**Description.** Length of mesosoma 1.1 mm. Mesosoma including propodeum covered by coarse and irregularly areolate sculpture of chelonine type, scutellum less strongly sculptured. Mesoscutum without notauli. Prescutellar depression wide and distinctly crenulated. Propodeum without any teeth or tubercles, with rather distinct semicircular transverse carina submedial. Forewing: discal cell distinctly petiolate. 1-SR 0.2 times as long as 1-M; cu-a distinctly postfurcal, inclined posteroapically, distant from 1-M for 0.7 times cu-a length. Carapace of metasoma rather long and narrow, twice as long as wide,



**Text-figure 35** *Ascogaster (Ascogaster) yulei* sp. nov., holotype NHMUK I.9209/I. 9379 (A) part; (B) counterpart. Scale bar = 1 mm.

rather distinctly roundly narrowed in posterior quarter, without sutures, entirely densely reticulate-areolate with elongate areolae, without distinct striation. Basal carinae distinct, convergent in their anterior half (where connected by distinct transverse carina), then subparallel for rather long distance. Colour: mesosoma black, metasoma black; possibly pale in wide basomedial area; forewing almost hyaline in basal half.

*Ascogaster (Ascogaster) yulei* Belokobylskij, sp. nov.  
Plate 12, figs 1, 2; Text-fig. 35

**Etymology.** After A. Yule, who has collected a lot of important fossil specimens in Bembridge Marls.

**Holotype.** NHMUK I.9209/I.9379 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; showing head with incomplete antenna and mesosoma with forewings and leg fragments in dorsal and ventral aspects.

**Diagnosis.** Different types of preservation of the present and previous species make them difficult to compare, leaving only larger body size, shorter prescutellar depression and presence of a transverse propodeal carina in *A. yulei* to discriminate them. The new species is also very similar to *A. pinicola* Brues, 1933, except that it has notauli indistinct, scutellum finely reticulate-areolate, propodeum lacking teeth, 3-SR almost equal to r, and m-cu distinctly postfurcal. *A. yulei* differs from *A. wheeleri*, besides the supposed absence of the metasomal sutures, in having 3-SR short and almost equal to r, and the sternauli present. The new species differs from all the *Ascogaster* species described by Tobias (1987) from Baltic amber in having temple large, notauli indistinct, and 1-R1 shorter than pterostigma.

**Description.** Median length of head 0.4 mm, mesosoma 1.25 mm, forewing 2.2 mm. Head almost twice as wide as long medially, 1.5–1.6 times as wide as maximum length. Head behind eyes distinctly roundly narrowed. Transverse diameter of eye 0.9–1.0 times length of temple. Occipital carina distinct, complete dorsally, fused with hypostomal carina above base of mandible. Malar space about as long as basal width of mandible. Antenna: scape rather long, almost twice as long as maximum width. Basal segments of flagellum rather slender.

Mesosoma 1.55–1.65 times as long as maximum width. Notauli absent. Mesoscutum rather long, its length 0.65 times as long as maximum width, finely areolate. Prescutellar depression rather deep, more or less short, with five carinae. Scutellum finely reticulate-areolate. Sternauli shallow, weakly S-shaped, distinctly crenulated. Mesopleuron apparently almost smooth in lower half. Propodeum without any distinct teeth or tubercles, with rather distinct semicircular transverse submedial carina. Wings: forewing 2.6 times as long as wide. Pterostigma wide, rather short and almost semicircular, 2.3 times as long as maximum width. Marginal cell strongly shortened, almost twice as long as wide. 1-R1 0.9 times as long as pterostigma, 1.3 times as long as distance from apex of marginal cell to apex of wing; r arising almost from posterior 0.3 of pterostigma, about half as long as maximum pterostigmal width. 3-SR almost equal to r, 0.2 times as long as the straight SR1. Second submarginal cell rather long, 2.6 times as long as wide; m-cu distinctly but not strongly postfurcal. Discal cell distinctly petiolate, about 1.5 times as long as wide, almost as long as second submarginal cell. 1-SR c. 0.2 times as long as 1-M; cu-a distinctly postfurcal, inclined posteroapically, distant from 1-M for 0.6–0.65 times cu-a length. Subdiscal cell wide, closed antero-posteriorly by CU1b, almost twice as long as wide. CU1a arising from posterior 0.3 of subdiscal cell; 3-CU1 straight. Colour: head and mesosoma black. Forewing faintly infusate. Pterostigma and veins brown to dark brown.

*Ascogaster (Ascogaster) pygmaea* Belokobylskij, sp. nov.  
Plate 11, figs 3, 4; Text-fig. 34B, C

**Etymology.** After *pygmaeus*, the Latin for tiny, referring to small body size of the species.

**Holotype.** NHMUK In.17379 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Smith Collection; dorsoventral aspect of head with incomplete antenna and partial mesosoma with incomplete forewing and leg fragments.

**Diagnosis.** The new species differs distinctly from all known fossil *Ascogaster* species in its small size, with forewing length 1.7 mm. It is similar to *A. pinicola* Brues, 1933 in small body size and discal cell petiolate anteriorly, but differs in having the basal segments of the antenna short and thick, m-cu distinctly postfurcal, marginal cell long, and 3-SR less strongly shortened. *A. pygmaea* is similar to *A. pentagona* Brues, 1933 in the shape of the second submarginal cell, but distinctly differs by having the forewing faintly infusate, the notauli absent, the mesonotum without striation, and the discal cell distinctly petiolate anteriorly. It differs from *A. thoracica* Tobias, 1987 in having the basal flagellar segments short and thick, the notauli indistinct, the propodeum without teeth, and 3-SR long. Comparison with *A. brodiei* is limited, because of their different states of preservation, yet they can be discriminated by the small size and distinctly transverse mesoscutum in the present species. *A. pygmaea* can be distinguished from *A. yulei* by the narrower head, short temple, wide mesoscutum, weakly shortened marginal cell, and long discal cell.

**Description.** Median length of head 0.25–0.3 mm, mesosoma 0.8–0.85 mm, forewing 1.7 mm. Head about twice as wide as long medially, 1.2 times as wide as maximum length, distinctly convex and roundly narrowed behind eyes. Transverse diameter of eye 1.1–1.2 times length of temple. Occipital carina distinct, fused below with hypostomal carina. Clypeus distinctly convex. Ocelli weakly enlarged. Antenna thick basally. Scape long and rather thick, 1.8 times as long as maximum width, 2.5 times as long as pedicel. First flagellar segment twice as long as apical wide, 1.1 times as long as second segment. Seventh flagellar segment thickened, 1.35–1.45 times as

long as maximum width. Mesosoma 1.5 times as long as maximum width. Pronotum distinctly elongate. Mesoscutum 0.55 times as long as maximum width, finely areolate. Notauli absent. Prescutellar depression rather deep, wide, with several carinae. Sternauli indistinct. Mesopleuron distinctly finely areolate. Prepectal and postpectal carinae present. Forewing 2.8 times as long as wide. Pterostigma wide, rather long, subtriangular, 2.7 times as long as maximum width. Marginal cell weakly shortened, 2.8 times as long as wide. 1-R1 1.1 times as long as pterostigma, about 3.0 times as long as distance from apex of marginal cell to apex of wing; r arising from posterior 0.3 of pterostigma, about half as long as maximum pterostigmal width. 3-SR 0.8 times as long as r, 0.15 times as long as the straight SR1, 0.25 times as long as 2-SR. Second submarginal cell rather long, 2.3 times as long as wide; m-cu strongly postfurcal. Discal cell distinctly petiolate, about twice as long as wide, 1.1 times as long as second submarginal cell. 1-SR 0.25 times as long as 1-M; cu-a distinctly postfurcal, inclined posteroapically, distant from 1-M for 0.8 times cu-a length. Subdiscal cell wide, closed antero-posteriorly by CU1b, 2.3 times as long as wide. CU1a arising from posterior 0.25 of subdiscal cell; 3-CU1 straight. Colour: head and mesosoma dark reddish brown; forewing faintly infusate; pterostigma and veins brown to dark brown.

**3.9.5. Subfamily Doryctinae Foerster, 1862.** Doryctinae is one of the most diversified subfamilies of idiobiont Braconidae (Belokobylskij *et al.* 2004b), with more than 200 known genera.

Many doryctine taxa have been described or recorded as fossils. The most species (eight) were described in the genus *Doryctes* Haliday, 1836 (s.l., including *Ontsira* Cameron, 1900) (Brues 1933; Statz 1938; Zhang 1989). In many cases, it is not easy to establish the modern generic position for these taxa because of insufficient descriptions and/or preservation state. The position of most of the species described by Brues in the genus *Doryctes* (*D. contectus* Brues, 1933, *D. filiformis* Brues, 1933, *D. hyalinus* Brues, 1933, and *D. succinalis* Brues, 1933) seems to be correct, but *D. minutus* Brues, 1933 is hardly a member of this genus, judging from its different wing venation and the absence of basiventral tubercles on the hind coxa. The species described in the genus *Clinocentrus* (*C. microps* Brues, 1933, *C. caudatus* Brues, 1933 and *C. debilis* Brues, 1933) probably also belong to *Ontsira* or *Doryctes* (Belokobylskij 1995). The three species described by G. Statz (1938) from the Aquitanian (latest Oligocene or earliest of Miocene) of Rott in Germany as *Doryctes*. *D. conjectus* Statz, 1938 and *D. rotundatus* Statz, 1938 have a correct position in this genus, but *D. longulus* Statz is not a Doryctinae, because its forewing discal cell is sessile, the hind wing sub basal cell is very long and the m-cu is absent. *D. huadongensis* Zhang, 1989 from the Middle Miocene of Shanwang, China, is perhaps a real *Doryctes* (Zhang 1989). Three species from Rott have been described in the genus *Spathius* Nees, 1818 (*S. pedicularis* Statz, 1936, *S. longicornis* Statz, 1938 and *S. macroradialis* Statz, 1938) the first two of them probably belong to *Doryctes*, and the last one to subfamily Opiinae. The elongate first metasomal tergite not transformed into a real petiole can be found in some species of doryctine genera (including *Doryctes*), as well as in other braconid groups, and especially in males (cf. *S. pedicularis*). Brues suggested *Ichneumon petrinus* Scudder, 1890 from White River, Colorado (USA) to be a member of *Spathius* (Brues, 1910) or *Rhaconotus* Ruthe, 1854 (Brues, 1933), but these suggestions need verification. Finally, two braconid wasps from the Bembridge Marls, *Phanomeris colenutti* Cockerell, 1921a and *Xenarcha distracta* Cockerell, 1921a should be transferred to the genus *Ontsira* (see below).

Two genera of the tribe Hecabolini are mentioned in the fossil record as well. The diagnostic characters of *Hecabolus*

*gladiator* Statz, 1936 described from Rott (forewing discal cell sessile anteriorly, subdiscal cell closed apico-posteriorly, shape of body with long ovipositor) suggest its position rather in the brachystine genus *Eubazus*. Real diagnostic characters of the genus *Polystenus* Foerster, 1862 have been obscured for long time (Papp 1984). As a result, *P. obduratus* Brues, 1933 from the Baltic amber is actually not a member of this genus (if it is a real member of Doryctinae). The description, even if supported with no illustration, indicates this species should form a genus of its own, based on the described combination of characters: occiput very faintly margined above; malar space long; hind coxa with no basiventral tubercle; m-cu antefurcal; and cu-a far postfurcal.

Very interesting is the discovery of specialised genera of the tribe Ecphylini *Ecphyllus oculatus* Muesebeck, 1960 and *Aivalylus dominicanus* Zuparko & Poinar, 1997, in the Miocene Mexican and Dominican ambers. Genus *Heterospilus* Haliday, 1836 from the Santonian (Late Cretaceous) Taimyr amber is reported by Zherikhin (1978), but this information needs verification. The record of the genus *Rhaconotus* Ruthe, 1854 (as *Hormiopterus* Girault, 1869; Scudder 1890; Brues 1910) in the Florissant (latest Eocene of Colorado) is doubtful as well.

There are taxa erroneously described in Doryctinae. The genus *Promonolexis* Brues, 1933 (type species *P. klebsi* Brues, 1933) from the Eocene Baltic amber (Brues 1933) is most probably a junior synonym of *Blacus* Nees. The characters shared by these genera are: discal cell sessile (the character state practically unknown in Doryctinae); ovipositor thickened and strongly curved down (practically unknown in doryctines and more characteristic of Euphorinae s. l.); antenna 18-segmented (characteristic of some species of *Blacus*); and long and narrow mesonotum and metasoma slender basally (also the characters often met in *Blacus*). The genus *Semirhytus* Szepilgeti, 1902, having been included in Rogadinae by Brues (1933), may differ radically from the modern one. Then *S. caudatus* Brues, 1933 from the Baltic amber is probably a member of the tribe Clinocentrini (Exothecinae s.l.) rather than of Doryctinae. Lastly, *Anacanthobracon* Brues, 1939 described in Doryctinae, also from the Baltic amber, is apparently a member of Cheloninae (see above).

The taxonomic position of one doryctine fossil is not yet clear. *Doryctomorpha tertiaria* Brues, 1933 (Baltic amber) was described in the genus *Doryctomorpha* Ashmead, 1900, which is now included in Rhyssalinae (Belokobylskij *et al.* 2004a). The original description is insufficient, so progress in understanding that species depends on finding the type or new material.

Only three doryctine species are recorded from the Bembridge Marls, all belonging to the genus *Ontsira*.

#### Genus *Ontsira* Cameron, 1900

**Remarks.** This is the first reliable fossil record of the genus *Ontsira*, because all earlier records appearing under the name *Doryctes* are not easy to fit to these genera based on their original descriptions.

*Ontsira colenutti* (Cockerell, 1921), comb. nov.  
Plate 12, fig. 3; Text-fig. 36

1921a *Phanomeris* (?) *colenutti* Cockerell, p. 18, fig. 20.

**Holotype.** NHMUK I.9702/ I.9096 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; head and mesosoma in lateral aspect with forewing and incomplete legs.

**Diagnosis.** The species differs from all known fossil *Doryctes* s.l. (*Ontsira* included) in having mesosoma short and high.

**Description.** Length of mesosoma 1.5 mm, hind femur 0.85 mm, forewing 3.5 mm. Head behind eyes roundly narrowed. Occipital carina present below. Mesosoma short and high, 1.45 times as long as maximum height. Mesoscutum highly and almost roundly elevated above pronotum. Notauli complete, wide anterior and narrow posterior. Scutellum distinctly convex. Prepectal carina distinct. Sternauli deep, rather wide, crenulated at least partly, situated in anterior 0.6 of mesopleuron. Propodeum with basilateral areas delineated by carinae. Forewing 2.7 times as long as maximum width. Pterostigma subtriangular, rather wide, 3.0 times as long as maximum width. 1-R1 1.4 times as long as pterostigma, 2-R1 very short; r arising almost from middle of pterostigma, 0.8 times as long as maximum pterostigmal width. Marginal cell not shortened, pointed apically, 3.0 times as long as maximum width. 3-SR twice as long as r, 0.5 times as long as the almost straight SR1, 1.2 times as long as 2-SR; r-m present, weakly sclerotised, 0.6 times as long as 2-SR. Second submarginal cell 2.3 times as long as wide, 1.25 times as long as subdiscal cell. 1-SR+M straight, 9.0 times as long as 2-SR+M. 2-SR+M 0.15 times as long as 2-SR, 0.2 times as long as m-cu, 0.1 times as long as sclerotised part of 2-M. Discal cell distinctly petiolate anterior, 1.8 times as long as wide; m-cu distinctly antefurcal. Cu-a postfurcal, distant from 1-M for 0.4 times Cu-a length. Subdiscal cell wide, 3.0 times as long as width; cula arising from posterior 0.3 of distal margin of subdiscal cell, rather distinctly pigmented in about its basal half. Legs: hind coxa with basiventral tubercle, 1.3 times as long as wide (tubercle included). Hind femur widened, 3.3 times as long as wide. Hind tibia not thickened, 1.6 times as long as femur, 2.5 times as long as hind basitarsus. Colour: head pale brown; mesosoma brown to pale brown; hind leg brown; hind tibia pale in basal 0.3; wings almost hyaline; pterostigma brown, pale basally and apically; veins brown.

**Remarks.** The position of this species in *Ontsira* is justified by the following characters: prepectal carina and sternauli present, notauli distinct and complete, hind coxa with basiventral tooth, hind femur thick, propodeum with basilateral areas delineated by carinae.

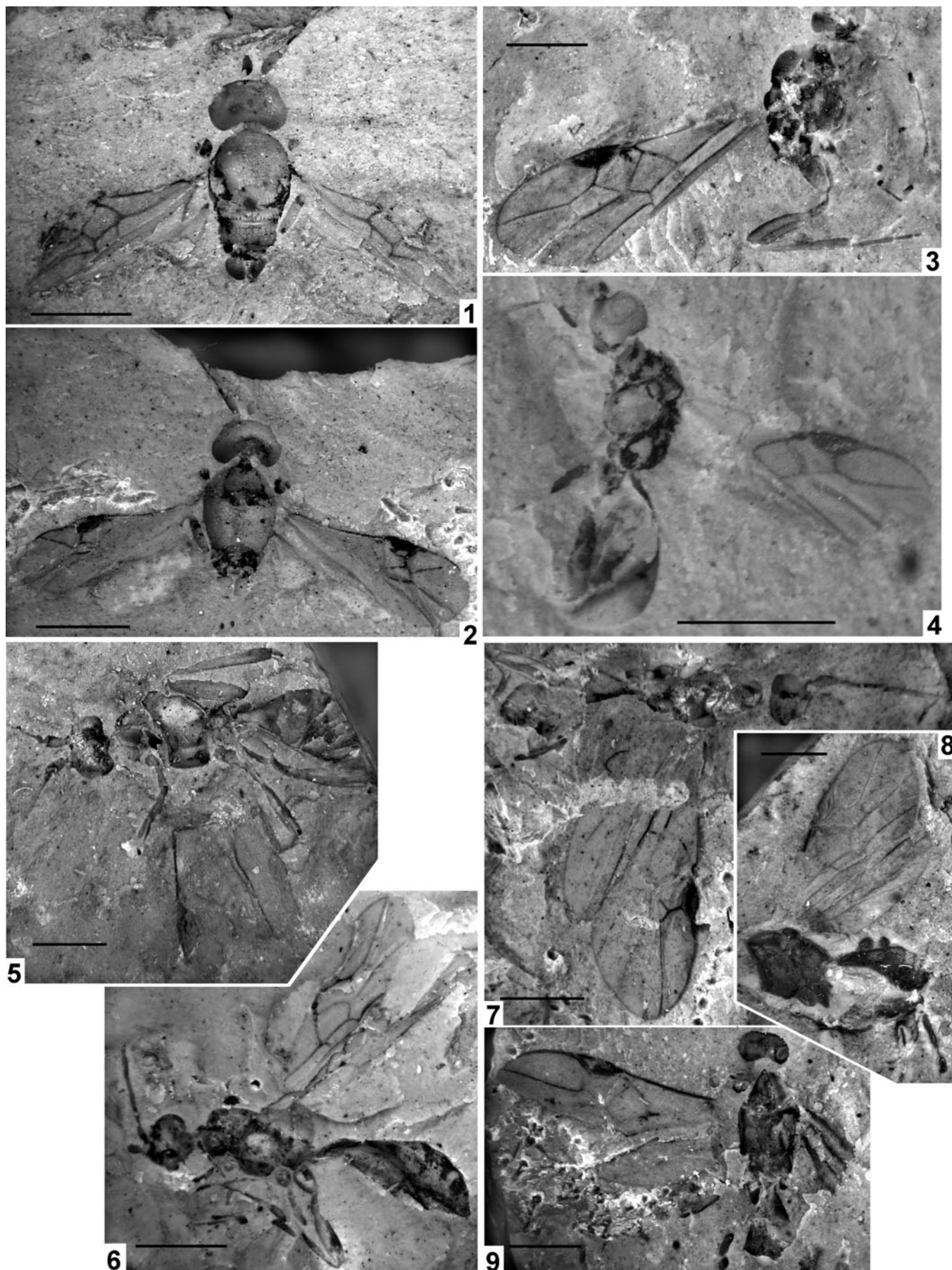
*Ontsira distracta* (Cockerell, 1921), comb. nov.  
Plate 12, fig. 4; Text-fig. 37

1921a *Xenarcha* (?) *distracta* Cockerell, 15, fig. 16.

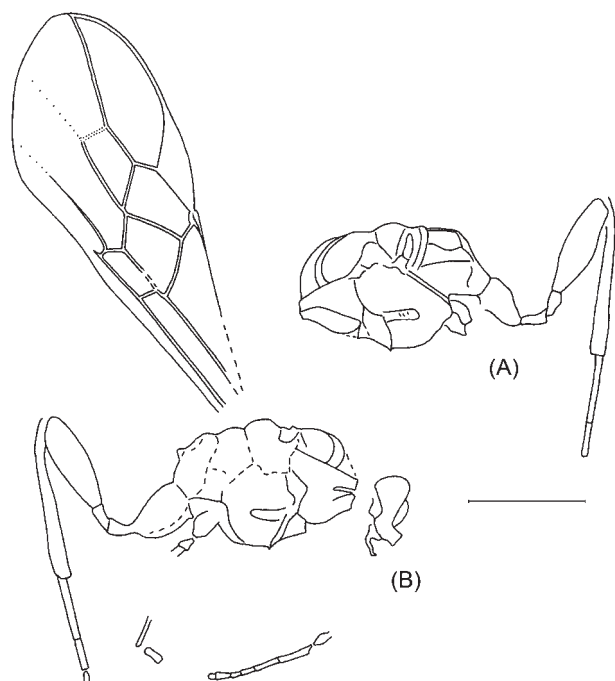
**Holotype.** NHMUK I.9099, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; insect in side view lacking hind wings and with incomplete antenna and legs.

**Diagnosis.** The species differs from the other known fossil *Ontsira* species in having body size small, basilateral areas of propodeum short, forewing distinctly infusate and with anterior position of CU1a.

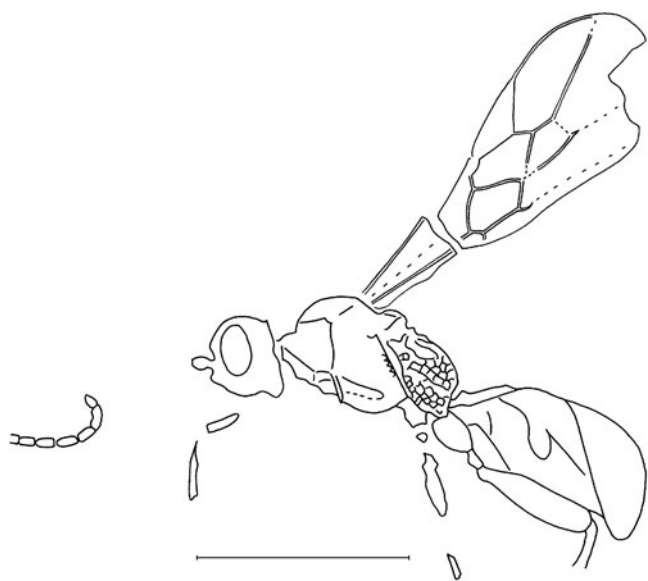
**Description.** Length of body 2.0 mm, mesosoma 0.8 mm, metasoma 0.95, hind femur 0.5 mm, forewing 1.6 mm. Head rather long, 1.3 times as high as long. Occipital carina developed. Hypoclypeal depression distinct, rather wide. Face transversely striate. Occipital carina distinct, fused below with hypostomal carina distinctly above mandibular base. Eye c.1.5 times as long as temple in lateral view. Segments in apical half of antenna elongate but rather thick, c. 3.0 times as long as wide. Mesosoma 1.7 times as long as maximum height. Mesoscutum highly and roundly elevated above pronotum. Scutellum convex. Prepectal carina distinct. Sternauli rather wide, situated



**Plate 12** (1–2) *Ascogaster (Ascogaster) yulei* sp. nov., holotype, NHMUK I.9209/I.9379: (1) part; (2) counterpart. (3) *Ontsira colenutti* (Cockerell), holotype, NHMUK I.9702 (part). (4) *Ontsira distracta* (Cockerell), holotype, NHMUK I.9099. (5) *Meteorus crassitergum* sp. nov., holotype, NHMUK I.9065. (6) *Ontsira cenozoica* sp. nov., holotype, NHMUK I.9601. (7–9) *Meteorus applanatus* sp. nov.: (7) holotype, NHMUK I.9656; (8) paratype, NHMUK I.8894; (9) paratype, NHMUK I.8674. Scale bars = 1 mm.

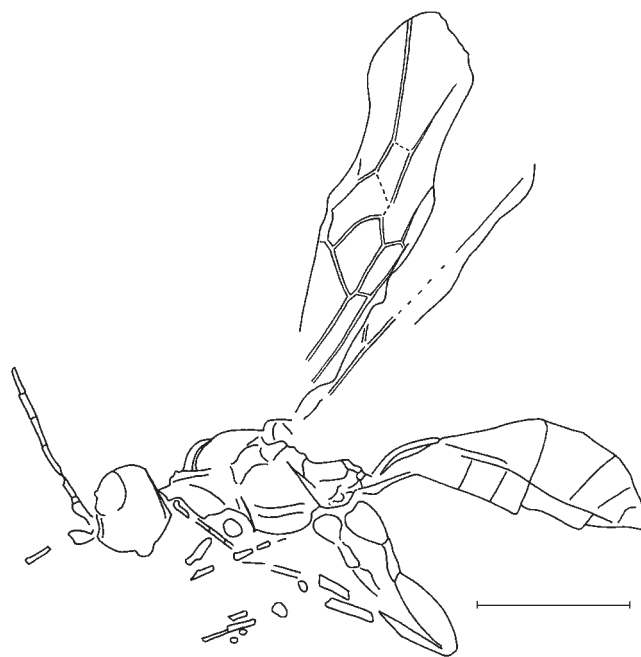


**Text-figure 36** *Ontsira colenutti* (Cockerell), holotype: (A) NHMUK I.9702 (part); (B) NHMUK I.9096, mesosoma and hind leg (countpart). Scale bar = 1 mm.



**Text-figure 37** *Ontsira distracta* (Cockerell), holotype, NHMUK I.9099. Scale bar = 1 mm.

in anterior 0.7 of mesopleuron. Mesopleural suture distinctly crenulated. Metapleura areolate throughout. Propodeum almost entirely finely areolate, with rather small basilateral areas more or less delineated by carinae. Forewing 2.8 times as long as maximum width. Pterostigma subtriangular, rather wide, 2.8 times as long as maximum width. Parastigma rather thick. 1-R1 1.2 times as long as pterostigma, 5.5 times as long as distance from apex of marginal cell to apex of wing; 2-R1 almost absent; r arising near middle of pterostigma, shortly unsclerotised apically (unless damaged there), not perpendicular to pterostigma, 0.75 times as long as maximum pterostigmal width. Marginal cell weakly shortened, pointed apically, 2.6 times as long as maximum width. 3-SR 1.15 times as long as r, 0.25 times as long as regularly and weakly curved SR1, 0.6



**Text-figure 38** *Ontsira cenozoica* sp. nov., holotype, NHMUK I.9601. Scale bar = 1 mm.

times as long as 2-SR; r-m present, faintly sclerotised, 0.55 times as long as 2-SR. Second submarginal cell 2.25 times as long as wide, 1.2 times as long as subdiscal cell. 1-SR+M weakly S-shaped. 2-SR+M very short. M-cu 0.7 times as long as 2-SR, 0.45 times as long as sclerotised part of 2-M. Discal cell distinctly petiolate anteriorly, 1.7 times as long as wide. 1-M about 3.0 times as long as 1-SR; m-cu weakly antefurcal; cu-a distinctly postfurcal. Subdiscal cell narrow. CU1a arising close to anterior side of subdiscal cell, weakly curved basally. Legs: hind coxa possibly with small basiventral tubercle, 1.4 times as long as wide. Hind femur distinctly widened, about 3.0 times as long as wide. Metasoma with no characters to describe. Colour: head possible entirely black; mesosoma black; first metasomal tergite black; laterotergites and sternites of metasoma light brown; hind leg brown; wings distinctly infuscate; pterostigma entirely dark brown, pale basally and apically; veins brown.

**Remarks.** The position of this species in the genus *Ontsira* is tentative, because several important characters (structure of notauli, subdiscal cell, second metasomal tergite, etc.) are not preserved.

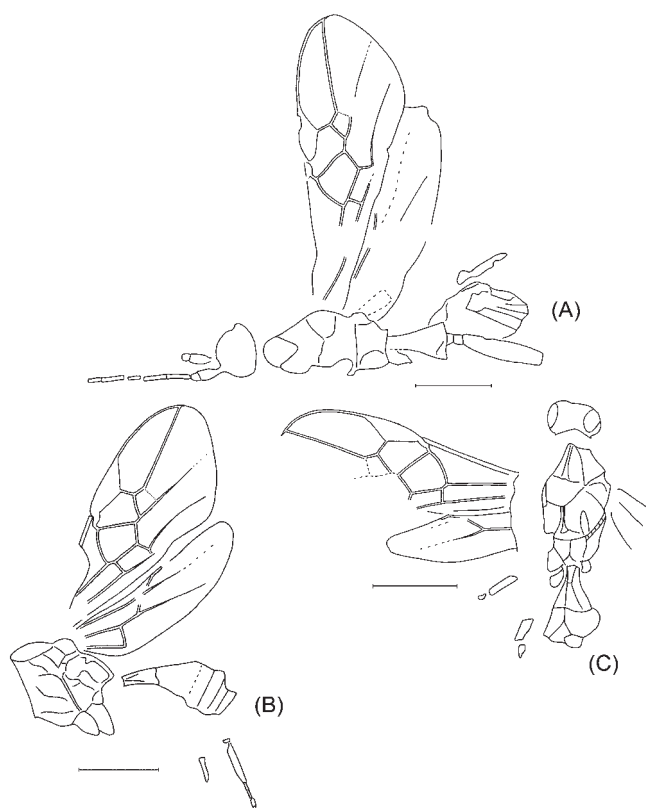
*Ontsira cenozoica* Belokobylskij, sp. nov.

Plate 12, fig. 6; Text-fig. 38

**Etymology.** After Cenozoic.

**Holotype.** NHMUK I.9601, Brodie Collection; insect in side view with incomplete antenna, wings and legs.

**Diagnosis.** Differs from *O. colenutti* (Cockerell) in having mesosoma long, second submarginal cell short and narrow, and 1-SR+M distinctly curved. Of Brues' species of *Doryctes*, the new species is similar to *D. hyalinus* Brues in having long and slender basal antennal segments, but differs in having the temple long, first metasomal tergite not strongly convex above, 3-SR long, m-cu distinctly antefurcal, and 2-SR and r-m not strongly convergent anteriorly. *O. oligocenica* differs from *D. filiformis* Brues in having the first metasomal tergite long and not strongly convex above, the second tergite without a basal area, cu-a less distinctly postfurcal, and m-cu



**Text-figure 39** *Meteorus applanatus* sp. nov.: (A) holotype, NHMUK I.9656; (B) paratype, NHMUK I.8894; (C) paratype, NHMUK I.8674. Scale bars = 1 mm.

distinctly antefurcal. Differs from *Spathius longicornis* Statz in having m-cu distinctly antefurcal and hind femur thick.

**Description.** Length of body 3.6 mm, mesosoma 1.3 mm, metasoma 1.9 mm, forewing 2.6 mm. Head strongly convex dorsally, 1.2 times as high as long in lateral view. Occipital carina present. Temple in lateral view about half as long as eye. Face rather weakly convex. Antenna rather slender. Scape convex, 1.6 times as long as wide, 2.2 times as long as pedicel. First flagellar segment 4.3 times as long as wide apically, 1.2 times as long as second segment. Mesosoma long and rather high, about twice as long as maximum height. Pronotum convex dorsally. Mesoscutum rather highly and roundly elevated above pronotum. Notauli present at least in anterior half. Scutellum convex. Prepectal carina distinctly developed. Sternauli present but shallow. Forewing about 3.5 times as long as maximum width. Pterostigma ill-preserved. 2-R1 almost absent. Marginal cell weakly shortened, pointed apically,  $c.3.5$  times as long as maximum width. 3-SR 1.5 times as long as r, 0.35 times as long as the almost straight SR1, 0.9 times as long as 2-SR; r-m weakly sclerotised, 0.55 times as long as 2-SR. Second submarginal cell rather short, 2.3 times as long as wide, as long as subdiscal cell. 1-SR+M distinctly convex, 4.5 times as long as 2-SR+M. 2-SR+M 0.35 times as long as 2-SR, 0.5 times as long as m-cu, 0.2 times as long as 2-M. Discal cell distinctly petiolate anterior, twice as long as wide; m-cu distinctly antefurcal; cu-a postfurcal, distant from 1-M for 0.7 times cu-a length. Subdiscal cell wide, 3.5 times as long as wide. CU1a arising from posterior 0.25 of distal margin of subdiscal cell, rather distinctly pigmented basally. Legs: hind coxa with basiventral tubercle, without processes dorsally, 1.65 times as long as wide; hind femur wide, 3.3 times as long as wide. Metasoma: first tergite rather long; 0.8 times as long as hind femur; second and following tergites smooth. Colour: body brown or black; forewing faintly infuscate; pterostigma and veins brown or dark brown.

**3.9.6. Subfamily Euphorinae Foerster, 1862.** The subfamily Euphorinae is a diverse group of koinobiont braconid wasps with a wide spectrum of hosts and stages (Tobias 1965, 1966; Shaw 1985). The members of this group are common and almost cosmopolitan.

Several euphorine taxa are recorded as fossils, mainly in amber (Brues 1933, 1939; Carpenter *et al.* 1937; Tobias 1987). Besides the tribe Blacini, which is sometimes considered as a separate subfamily (Achterberg 1982, 1988), species of the tribes Meteorini, Oncometeorini, Perilitini, Centistini, Euphorini, Prosyntretini and Neoneurini are recorded as fossils. Among them are five species of *Meteorus* Haliday, 1835: *M. brevis* Brues, 1933, *M. crassicornis* Brues, 1933, *M. elongatus* Brues, 1933 and *M. interstitialis* Brues, 1933 from the Baltic amber, and "*Meteorus*" *longicornis* Statz, 1938 from Rott, near the Oligocene/Miocene boundary in Germany. The position of the amber species is clear, but the status of *M. longicornis* needs verification. *Fushunobracon orientalis* Hong, 2001 from the Eocene Fushun amber in China might also be a member of the tribe Meteorini.

Two genera of the tribe Centistini are recorded as fossils: *Pygostolus* Haliday, 1833 and *Parasyrrhizus* Brues, 1933. The position of *Pygostolus* from the Canadian and Baltic ambers is not clear, because both described species, *P. clavatus* Brues, 1933 and *P. patriarchicus* Brues, 1937, particularly the last one, differ considerably from extant species of this genus. The forewing venation of *P. patriarchicus*, especially the shape of discal cell, is very similar to the extant genus *Korecnomis* Park & Achterberg, 1994. Described from Baltic amber is the genus *Parasyrrhizus* Brues, 1933 (type species *P. ludens* Brues, 1933), similar to extant *Spathicopsis* Achterberg, 1977 in forewing venation and the presence of notauli, but it apparently differs in having the sternauli absent, the body stout, and the antenna short and stout (shape of the first tergite is unclear). Three species and two genera were described by Brues (1933, 1939) in the tribe Perilitini: *Perilitus* (*Microctonus*) *nanus* (Brues, 1933), comb. nov., and two possibly related species with petiolate first tergite in two monotypic genera *Onychoura* Brues, 1933 (type species *O. petiolata* Brues, 1933) and *Meteorites* Brues, 1939 (type species *M. inopinata* Brues, 1939). A single species, *Euphorus indurescens* Brues, 1910, was described in the tribe Euphorini on the basis of a poorly preserved fossil from Florissant (Brues 1910).

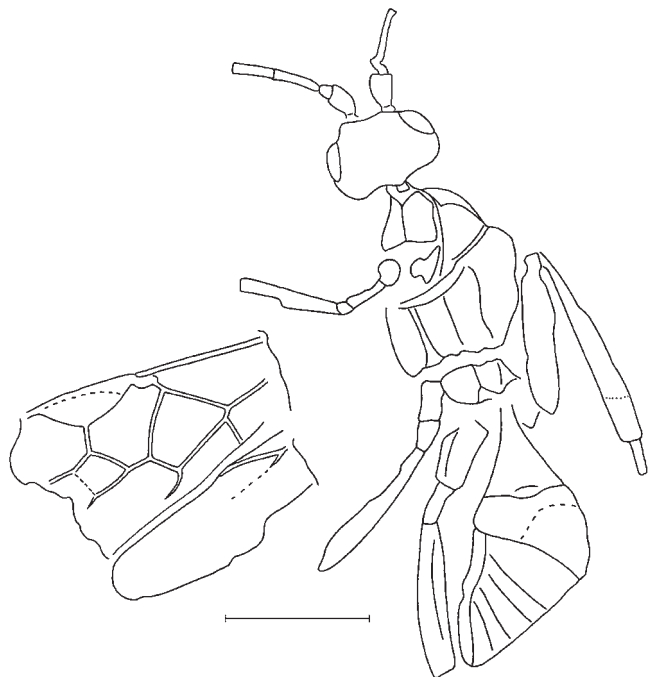
There is an undoubtedly a remarkable discovery in the Baltic amber of the specialised euphorine tribe Neoneurini, represented there by a monotypic extinct genus *Elasmosomites* Brues, 1933 (type species *E. primordialis* Brues, 1933), and the monotypic and exclusively Eocene tribes Oncometeorini Tobias, 1987 (*Oncometeorus aduncus* Tobias, 1987 with abnormally hook-shaped ovipositor) and Prosyntretini Tobias, 1987 (*Prosyntretus calcaratus* Tobias, 1987).

Composition of the euphorine fossils in the Bembridge Marls is much more modest, with only two identifiable species of *Meteorus* described below.

#### Genus *Meteorus* Haliday, 1835

**Remarks.** Described fossil species of *Meteorus* (Brues 1933; Statz 1938) have m-cu distinctly postfurcal (entering the second submarginal cell) or at most interstitial. In contrast, all Bembridge *Meteorus* have m-cu far antefurcal (entering the first submarginal cell well before its apex).

*Meteorus applanatus* Belokobyl'skij, sp. nov.  
Plate 12, figs 7–9; Text-fig. 39



**Text-figure 40** *Meteorus crassitergum* sp. nov., holotype, NHMUK I.9065. Scale bar = 1 mm.

**Etymology.** After *applanatus*, the Latin for flattened, referring to shape of fossil.

**Holotype.** NHMUK I.9656, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; dorsal aspect of insect with each head, meso- and metasoma, antennae, wings and legs more or less incomplete.

**Paratypes.** NHMUK I.8674, lateral aspects of incomplete mesosoma and metasoma with wings and partial legs; NHMUK I.8894 ventral aspect of head, mesosoma, metasomal base, incomplete wings and leg fragments. All from Bembridge Marls, NW Isle of Wight, UK, Brodie Collection.

**Diagnosis.** The new species differs from *M. crassitergum* sp. nov. in having *r* arising weakly behind the middle of the pterostigma, first metasomal tergite less wide apically, and 1-M weakly curved. Comparison with extant species of *Meteorus* is complicated by insufficient preservation.

**Description of holotype.** Body length *c.* 4.2 mm; length of head, mesosoma and first metasomal tergite combined 3.1 mm, mesosoma 1.6 mm, forewing 3.9 mm. Head in lateral view 1.3 times as high as maximum length. Antenna: scape large, 1.6 times as long as maximum width, twice as long as pedicel. First flagellar segment *c.* 5.0 times as long as maximum width, almost as long as second segment. Mesosoma 2.4 times as long as maximum width at level of mesoscutum. Median lobe of mesoscutum distinctly convex anteriorly; notauli present. Forewing 2.6 times as long as maximum width. Pterostigma subtriangular and wide, 2.7 times as long as maximum width; *r* arising slightly beyond middle of pterostigma, 0.8 times as long as maximum pterostigmal width. Marginal cell slightly shortened, 1-R1 about 1.3 times as long as pterostigma. 3-SR 0.9 times as long as *r*, 0.15 times as long as the straight SR1, 0.6 times as long as 2-SR, about as long as *r*-m. Second submarginal cell 1.2 times as long as wide, 0.6 times as long as subdiscal cell. 1-SR+M weakly curved; *m*-cu strongly antefurcal, almost twice as long as 2-SR+M. Discal cell distinctly and shortly petiolate anteriorly, 1.4 times as long as wide. 1-M distinctly convex; *cu*-a postfurcal, distant from 1-M for 0.45 times *cu*-a length. Legs: hind femur 4.0 times as long as

maximum width, with rather wide and long transverse protuberance along lower 0.6. Metasoma: first tergite distinctly and almost linearly widened towards apex, without spiracular tubercle, its length 1.6 times apical width, apical width about 3.5 times its basal width. Colour: propodeum and first metasomal tergite black or dark brown; rest of metasoma possibly brown or pale brown; wings hyaline; pterostigma and veins dark brown.

Additional characters from paratypes. Forewing length 3.1–3.5 mm. Head transverse, about twice as wide as long, evenly roundly narrowed behind eyes. Eye rather large, transverse diameter of eye in dorsal view about twice as long as temple. Occipital carina distinct. Sides of pronotum almost entirely smooth. Prepectal carina distinct. Mesopleuron 1.4–1.5 times as high as long, with more or less distinct, wide and almost smooth sternaulus. Scutellum distinctly convex. Subalar depression rather deep and narrow. Forewing *c.* 2.2 times as long as wide. Pterostigma 3.0–3.3 times as long as maximum width; *r* 0.7–0.8 times as long as maximum width of pterostigma. 1-R1 1.2–1.3 times as long as pterostigma. 3-SR 0.7–0.8 times as long as *r*, 0.13–0.15 times as long as SR1, 0.45–0.5 times as long as 2-SR, 0.7 times as long as *r*-m. Second submarginal cell 1.05–1.2 times as long as width; *m*-cu 2.0–2.5 times as long as 2-SR+M. 1-SR+M almost straight. Discal cell 1.2–1.3 times as long as wide; *cu*-a postfurcal, distant from 1-M for 0.3–0.4 times *cu*-a length. Ventral margins of first tergite possibly separated.

*Meteorus crassitergum* Belokobylskij, sp. nov.

Plate 12, fig. 5; Text-fig. 40

**Etymology.** After *crassus*, the Latin for thick, and tergum, referring to width of the first metasomal tergite.

**Holotype.** NHMUK I.9065, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; subventral aspect of insect with antennae, wings, legs and metasoma incompletely preserved.

**Diagnosis.** Similar to extant *M. oculatus* Ruthe, 1862 (Huddleston 1980) in having hind tibia distinctly thickened, but differs in having the temple long, head behind the eyes rounded and less strongly narrowed, second submarginal cell and 2-SR+M long. Differs from *M. corax* Marshall, 1898 in having the first tergite wide apically, flagellar segments long, hind femur and tibia distinctly wider, and 2-SR+M of forewing long.

**Description.** Length of head, mesosoma and first metasomal tergite combined 2.8 mm, of mesosoma 1.6 mm. Head weakly transverse, 1.6 times as wide as median length. Head behind eyes weakly convex anteriorly and roundly narrowed posteriorly. Transverse diameter of eye in dorsal view about as long as temple. Antenna: Scape large, 1.6 times as long as maximum width. First flagellar segment 3.3 times as long as apical width, almost as long as second segment. Mesosoma 1.8 times as long as wide. Scutellum more or less convex. Prepectal carina present. Sternauli shallow and wide, almost smooth. Forewing: pterostigma subtriangular, wide, 2.8 times as long as maximum width; *r* arising distinctly behind middle of pterostigma, 0.7 times as long as maximum pterostigmal width. 3-SR 0.85 times as long as *r*, 0.55 times as long as 2-SR, 0.7 times as long as *r*-m. Second submarginal cell about as long as wide, 0.6 times as long as subdiscal cell. 1-SR+M weakly curved; *m*-cu strongly antefurcal, almost twice as long as 2-SR+M. Discal cell distinctly and shortly petiolate anteriorly, 1.3 times as long as wide; *cu*-a postfurcal, distant from 1-M for 0.6 times *cu*-a length. Legs: hind femur 4.5 times as long as maximum width; hind tibia distinctly thickened towards apex, 8.6 times as long as wide, its maximum width 0.7 times width of femur.

Metasoma: first tergite distinctly and almost linearly widened towards apex, without spiracular tubercle, its length  $c.1.2$  apical width, apical width 3.5 times its basal width. Colour: head, mesosoma at least in part, and metasoma dark brown to black; wings hyaline; pterostigma and veins brown.

**3.9.7. Subfamily Exothecinae Foerster, 1862, s. l.** Subfamily Exothecinae s. l. is one of the most disputed groups in the Braconidae, and several of its tribes now have the rank of subfamily (Achterberg 1993). The real taxonomic and phylogenetic positions of these taxa require special study using modern morphological, anatomical and molecular data. For the present, I prefer to use the subfamily in the broad sense.

A few exothecine genera and species have been recorded as fossils, although in many cases their taxonomic position needs re-study. Two species are described in the genus *Cantharoctonus* Viereck, 1912 (tribe Rhysipolini): *C. oligocenicus* Brues, 1933 from the Eocene Baltic amber (Brues 1933) and *C. bruesii* Statz, 1936 from near the Oligocene/Miocene boundary (Statz 1936), but their positions are problematic. *C. oligocenicus* was originally included in the Spathiini (Doryctinae) and characterised as having its first tergite petiolate, second metasomal tergite very long and covering almost entirely the following tergites, and CU1a arising near the middle of the distal margin of the closed subdiscal cell. *C. bruesii* was originally described as having the first tergite short and not petiolate, and CU1a arising far behind middle of the distal side of the subdiscal cell. Both descriptions are not enough to identify positions of these species with reasonable confidence. *Colastes abrogatus* (Brues, 1910) (originally *Exothecus abrogatus*: Brues 1910) shows characters unusual for extant species of *Colastes* Haliday, 1833, such as the extremely slender antenna, almost petiolate metasoma, interstitial position of both cu-a and m-cu, and large second submarginal cell distinctly widened towards its apex. *Rhysipolis distinctus* Théobald, 1937 described from the French Oligocene (Théobald 1937) also deserves verification of its generic placement.

Three genera of the tribe Hormiini have been reported from the Taimyr and Baltic ambers. The specialised extinct genus *Parahormius* has been mentioned as found in the Cretaceous Taimyr amber (Zherikhin 1978). *Hormius submersus* (Brues, 1933) was described (as *Hormiellus submersus*) from the Baltic amber (Brues, 1933). It has a venation of the *Hormius* type, but several other characters as described raise some doubts as to this placement: vertex, mesoscutum and scutellum chagrined; maxillary palpi long; pterostigma narrow; cu-a strongly postfurcal. One more fossil from Baltic amber has been described as the extinct monotypic genus *Prochremylus* Brues, 1933 (type species *P. brevicornis* Brues, 1933), also with *Hormius*-type forewing venation, but generically distinct from *Hormius* species in having short and thick antenna, subcircular head, enlarged femora, and indistinct sternaui.

Only one species of this group is recorded from the Bembridge Marls.

Genus *Chremylus* Haliday, 1833

*Chremylus infuscatus* Belokobylskij, sp. nov.

Plate 13, fig. 1; Text-fig. 41

**Etymology.** After *infuscatus*, the Latin for infusate, referring to colouration of the forewing.

**Holotype.** NHMUK I.9066, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; headless body with incomplete forewings and legs in dorsoventral view.

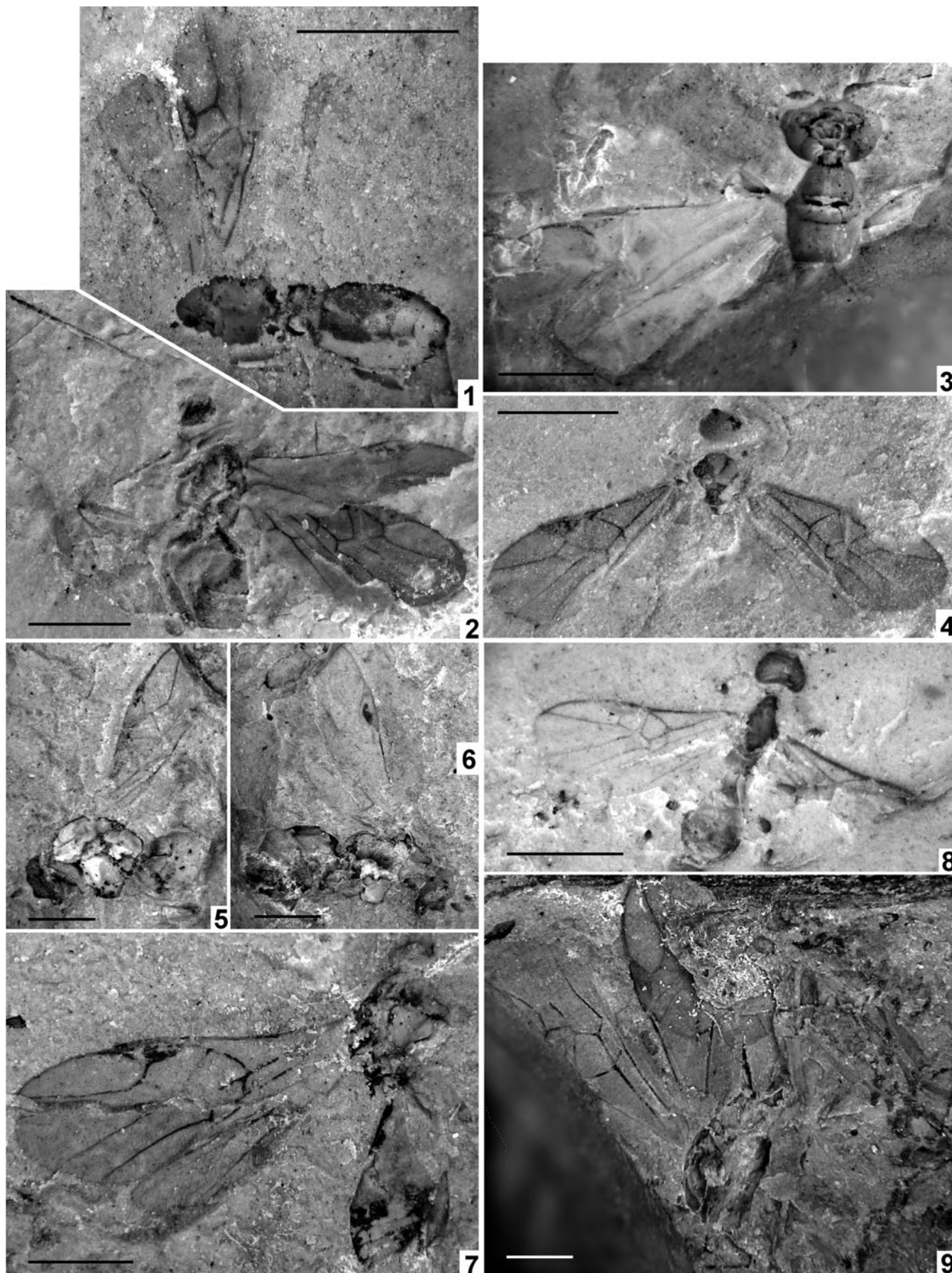
**Diagnosis.** Differs from the type species *C. elaphus* Haliday in having second and third tergites long, m-cu distinctly postfurcal, and forewing distinctly infusate.

**Description.** Length of metasoma 0.9 mm, forewing 1.4 mm. Mesosoma: prepectal carina present. Sternaui narrow, rather deep, situated in basal 0.6 of mesopleuron. Mesopleuron 1.5 times as high as long. Forewing  $c.2.5$  times as long as wide. Pterostigma subtriangular, wide, 3.2 times as long as maximum width; r arising from middle of pterostigma, 1.1 times as long as maximum pterostigmal width. Marginal cell not shortened, 1-R1 1.2 times as long as pterostigma. 3-SR 1.2 times as long as r, 0.4 times as long as the straight SR1, almost as long as 2-SR, 1.4 times as long as r-m. Second submarginal cell 2.2 times as long as width, 0.9 times as long as discal cell; m-cu distinctly postfurcal, 3.0 times as long as 2-SR+M, 0.7 times as long as 2-SR. 1-SR+M weakly S-shaped. Discal cell shortly petiolate anterior, 1.6 times as long as wide. 1-M and m-cu convergent posterior. Subdiscal cell closed posterior, rather narrow, 3.5 times as long as wide. CU1a not interstitial, arising distinctly before middle of distal side of subdiscal cell; cu-a postfurcal, distant from 1-M for about cu-a length. Legs: middle femur rather thick, 4.3 times as long as its width. Metasoma oval. First tergite strongly and almost linearly widened towards apex, its length 0.7 times apical width. Tergites behind first one rather soft, smooth, their combined length 1.5 times maximum width. Second suture indistinct. Median length of second and third tergites combined 1.15 times basal width of second tergite, 0.8 times maximum width of these tergites. Colour: propodeum and possibly all mesosoma dark brown; metasoma brown to dark brown; wings distinctly infusate; pterostigma and veins brown to dark brown.

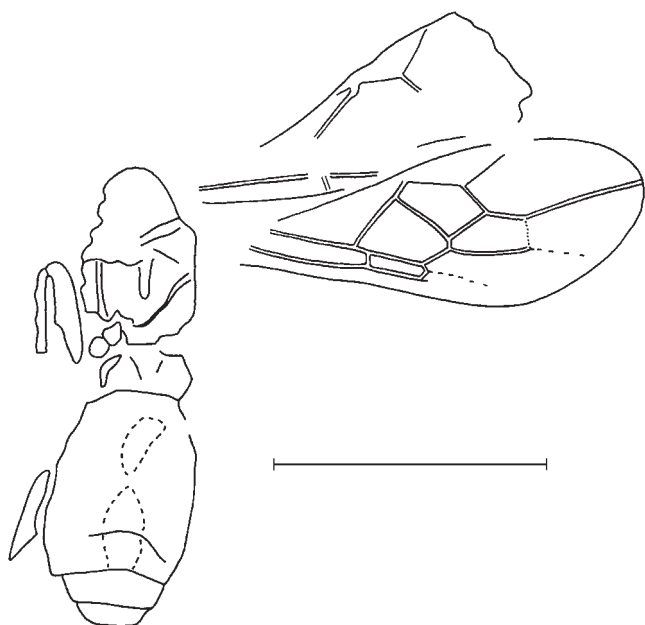
**Remarks.** The position of this species in *Chremylus* is not certain because of insufficient preservation, but seems acceptable for the present. The new species shares the following characters with *Chremylus*: second and third metasomal tergites enlarged; 1-M and m-cu distinctly convergent posteriorly; cu-a distinctly postfurcal; m-cu postfurcal (often almost interstitial in the type species of *Chremylus*); CU1a arising before middle of distal margin of subdiscal cell; sternaui distinct. These characters differentiate the new species from *Prochremylus*.

**3.9.8. Subfamily Helconinae Foerster, 1862.** The subfamily Helconinae is one of the less derived groups of koinobiont parasitoids developing inside of the larvae of xylophagous beetles. The taxa of this subfamily are relatively common in various kinds of fossil assemblages. One of the interesting results of the study of helconine wasps in the Baltic amber was the discovery of *Acampsohelcon rasnitsini* Tobias, 1987, with the carapace-like structure of the metasoma, which has been placed in *Acampsohelconini*, a tribe of its own (Tobias 1987) and later (Achterberg 2000) elevated at the subfamily rank. Wing venation of *Eobracon* Cockerell is similar to that of *Acampsohelcon*, but carapace tergites in this genus are possibly fused and the ovipositor is much longer (see also discussion in Cheloninae). *Sinobracon speciosus* Hong, 1974 described from the Fushun amber (Eocene of China) is possibly also a member of this subfamily.

The largest number of helconine fossil species were described in the genus *Aspicolpus* Wesmael, 1838. Venational characters used in the current taxonomy of Helconinae suggest the fossil *Aspicolpus* should be split into several groups of generic rank. However, this idea is not easily realised because of the incomplete descriptions and insufficiently informative figures, if any. That is why I suggest these fossils form three groups in the genus *Aspicolpus*. One of them includes *A. clarus* Brues, 1933 and *A. perditellus* Brues, 1933, which have short to very short 3-SR (not longer than r) and possibly smooth second and third metasomal tergites (known only for the first species; unknown in the extant species of *Aspicolpus*). *A. fraternus* Brues, 1933 is characterised by the discal cell petiolate anteriorly (unknown in Recent *Aspicolpus*), second submarginal cell distinctly narrowed



**Plate 13** (1) *Chremylus infuscatus* sp. nov., holotype, NHMUK I.9066. (2) *Calyptoides veterinus* Cockerell, holotype, NHMUK I.9722. (3) *Aspicolpus temporalis* sp. nov., holotype, NHMUK I.9935. (4) *Diospilus acourti* Cockerell, holotype, NHMUK I.9071. (5–6) *Diospilus curtithorax* sp. nov., holotype, NHMUK I.9286/I.9025: (5) part; (6) counterpart. (7) *Hellenius* ? *kozlovi* sp. nov., holotype, NHMUK In.17298. (8) *Taphaeus cervicalis* (Cockerell), holotype, NHMUK I.9199. (9) *Homolobus rasnitsyni* sp. nov., holotype, NHMUK In.20545. Scale bars = 1 mm.



**Text-figure 41** *Chremylus infuscatus* sp. nov., holotype, NHMUK I.9066. Scale bar = 1 mm.

distally, and *r* arising distinctly behind middle of the pterostigma. The third group is composed of *A. moniliformis* Brues, 1933 and *A. similis* Brues, 1933, which have a quite short second submarginal cell but, judging from other described features, they are probably the only real members of *Aspicolpus*. Based on wing venation, and particularly shape and size of the second submarginal cell, and because of rather long first metasomal tergite, *Aspicolpus longicornis* Statz, 1936 from near the Oligocene/Miocene boundary of Rott in Germany (Statz 1936) can be transferred to the genus *Taphaeus*. Several species similar to *Aspicolpus* in having rather long and not triangular second submarginal cell species were described by Brues (1933) in the genus *Microtypus* Ratzeburg, 1848. The status of all these taxa needs conformation: it is possible that some of them deserve transfer to Helconinae, Macrocentrinae or Homolobinae.

There is no morphological reason to include *A. minor* Brues, 1933 in *Austrohelcon* Turner, except perhaps for the absence of a deep frontal cavity (Brues 1933). This species, together with the one described as *Gymnoscelis klebsi* Brues, 1933, deserves transfer to the genus *Helcon* Nees, in spite of several special features of wing venation in the latter (second submarginal cell subsquare with almost parallel 2-SR and *r-m*, *cu-a* interstitial, only 2A present). One peculiar species from Baltic amber has been described in the genus *Eumacrocentrus* Ashmead, 1900 (*E. similis* Brues, 1933).

The sole true member of the genus *Diospilus* Haliday, 1833, *D. acourti* Cockerell, 1921, has been described from the Bembridge Marls (Cockerell 1921a). According to the description of the forewing venation, and particularly the shape of the second submarginal cell, *Diospilus repertus* Brues, 1910 from the Florissant (Brues 1910) probably belongs to *Aspicolpus* as well, despite the description lacking several diagnostic characters of this genus. The very peculiar wing venation of *Diospilus allani* Brues, 1937 described from the Cretaceous Canadian amber (Brues 1937) suggests that it belongs to Rhyssalinae rather than Helconinae, even if the shape of the clypeus is unknown and the hind wing is insufficiently described. Similar venation is described in the extant rhyssaline genus *Pseudobathystomus* Belokobylskij, 1986 as well as in some species of *Oncophanes* Foerster, 1862. Lastly, *D. verus* Théobald, 1937

from the Oligocene of France (Théobald 1937) is a quite distinct member of the subfamily Helconinae, but the shape of the second submarginal cell in the forewing indicates its position in the genus *Taphaeus*.

*Taphaeus precox* Brues, 1923 is apparently a true species of that genus from Baltic amber (Brues 1923). *Sigalphus cervicalis* Cockerell, 1921 from the Bembridge Marls (see below) and *Aspicolpus longicornis* Statz (see above) also deserve transfer to *Taphaeus* Wesmael, 1835. Peculiar wing venation similar to *Ascogaster* species is known for three species of the Eocene helconine genus *Electrohelcon* Brues, 1933 (*E. grandis* Brues, 1933, *E. minor* Brues, 1933 and *E. foveatus* Brues, 1933).

The status of *Discoletes soporatus* Brues, 1910 from the Florissant (Brues 1910) is not clear, because it has interstitial *m-cu* and pale yellowish brown pterostigma, which are unknown or rare in extant Helconinae. The taxonomic position of *Calypoides veterinus* Cockerell, 1921 from the Bembridge Marls is discussed below.

#### Genus *Calypoides* Cockerell, 1921

**Type species.** *C. veterinus* Cockerell, 1921

**Diagnosis.** Similar to *Taphaeus* Wesmael, 1835. Differs from that and all other genera of Helconinae in having a long and thick process on the dorsal surface of the hind coxa.

**Description.** Antennae long and slender. Mesosoma short and high, mesoscutum highly and almost vertically elevated above pronotum. Sternauli deep and wide. Prepectal carina probably developed. In forewing, pterostigma subtriangular and rather wide. 1-R1 longer than pterostigma; *r* arising almost from middle of pterostigma. Marginal cell not shortened, pointed apically. 3-SR shorter than *r*; *r-m* present. Second submarginal cell subtrapezoid and rather narrow. Discal cell shortly sessile anteriorly; *m-cu* far antefurcal; *cu-a* distinctly postfurcal. Subdiscal cell closed apico-posteriorly. 2A and *a* indistinct. Sub-basal cell of hind wing large. Legs slender and long. Hind coxa with rather thick, more or less pointed and rearward directed dorso-posterior process. First metasomal tergite as long as second and third tergites combined. Second metasomal suture absent.

#### *Calypoides veterinus* Cockerell, 1921

Plate 13, fig. 2; Text-fig. 42

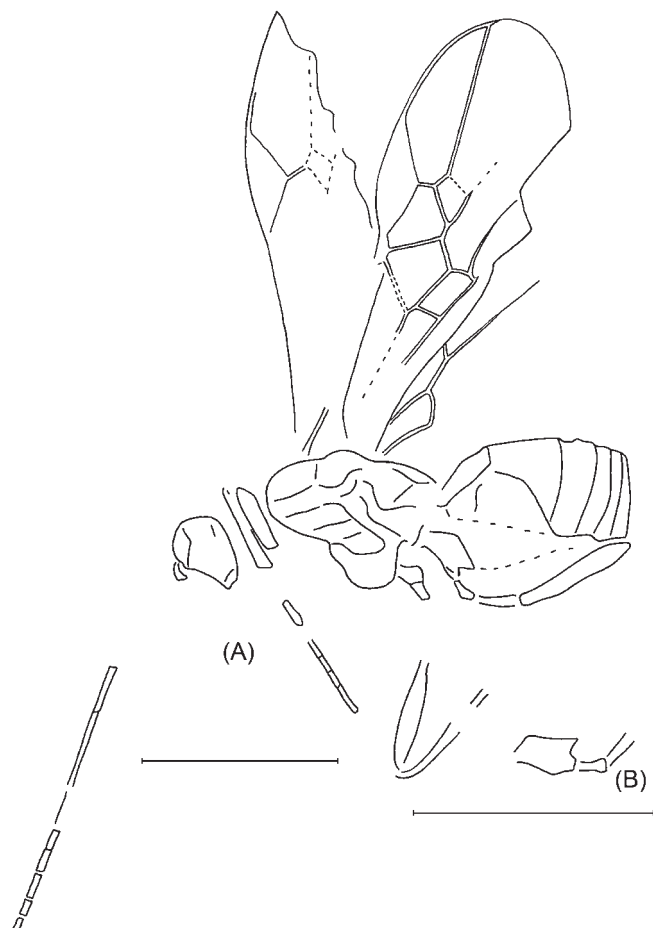
1921a *Calypoides veterinus* Cockerell, p. 13, fig. 13.

1992 *Calypoides veterinus* Cockerell, 1921: Carpenter, p. 478.

**Holotype.** NHMUK I.9722, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; lateral aspect of insect with appendages incompletely preserved.

**Diagnosis.** As for genus.

**Description.** Body length 2.1 mm, mesosoma 0.8 mm, hind femur 0.6 mm, metasoma 0.9 mm, forewing 2.2 mm. Head elongated below, its height 1.5 times maximum width. Antenna rather long, as preserved 0.9 times as long as forewing. Subapical antennal segment *c.* 4.0 times as long as wide. Mesosoma 1.2 times as long as maximum height. Scutellum distinctly convex. Forewing *c.* 2.8 times as long as maximum width. Pterostigma 3.0 times as long as maximum width. 1-R1 1.2 times as long as pterostigma, 2-R1 short. Marginal cell 3.0 times as long as maximum width. R 0.6 times as long as maximum width of pterostigma. 3-SR 0.8 times as long as *r*, 0.1 times as long as weakly curved SR1, 0.4 times as long as 2-SR; *r-m* unsclerotised, 0.7 times as long as 2-SR. Second submarginal cell about as long as wide. 1-SR+M straight, 3.5 times as long as 2-SR+M. 2-SR+M 0.4 times as long as 2-SR, 0.5



**Text-figure 42** *Calyptoides veterinus* Cockerell: (A) holotype, NHMUK I.9722; (B) same, hind coxa. Scale bars = 1 mm.

times as long as m-cu, 0.4 times as long as sclerotised part of 2-M. Discal cell 1.3 times as long as wide. Distance from 1-M for 0.35 times cu-a length. Subdiscal cell wide, 2.3 times as long as wide; CU1b distinct. CU1a arising from posterior 0.3 of distal side of subdiscal cell, rather distinctly sclerotised in about its basal half. Hind wing: M+CU 0.45 times as long as 1-M, 1.3 times as long as m-cu. Legs: middle femur 6.2 times as long as width; hind coxa rather elongate; hind femur 5.5 times as long as width. Metasoma 1.2 times as long as mesosoma. Second and third tergite long; following tergites short and distinctly protruding behind third one. Colour: possibly entirely black head; mesosoma dorsally and first tergite; metasoma behind first tergite brown; wings hyaline; pterostigma and veins brown.

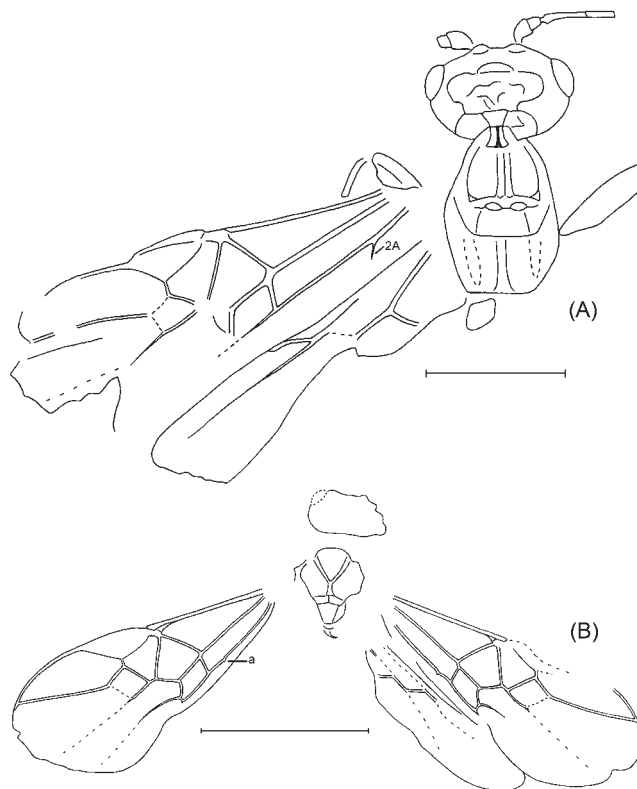
Genus *Aspicolpus* Wesmael, 1838  
*Aspicolpus temporalis* Belokobylskij, sp. nov.  
 Plate 13, fig. 3; Text-fig. 43A

**Etymology.** After *temporalis*, the Latin for temporal, referring to long temples.

**Holotype.** NHMUK I.9935, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; ventral aspect of head with incomplete antennae, pro- and mesothorax with incomplete wings and legs.

**Diagnosis.** Distinctly differs from *A. moniliformis* Brues and *A. similis* Brues (Brues, 1933) in having long 3-SR and second submarginal cell.

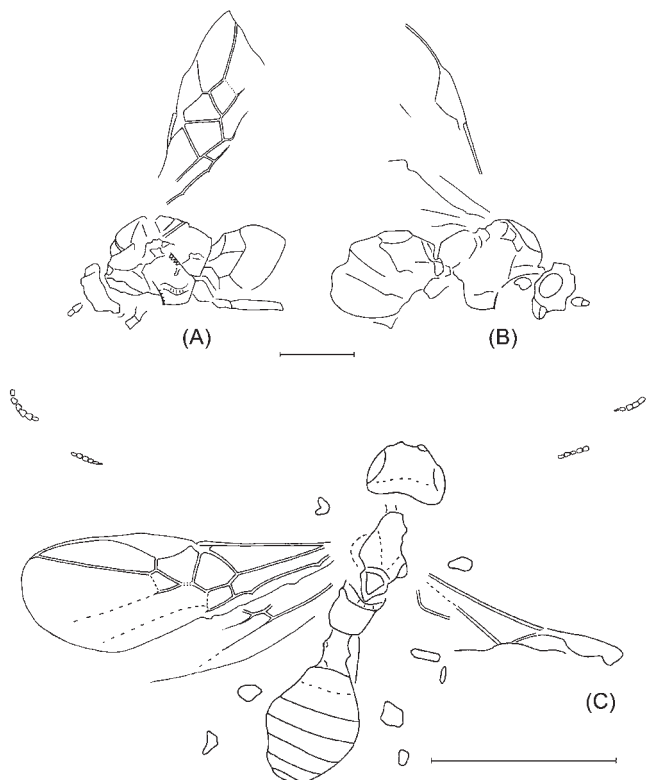
**Description.** Length of head 0.7 mm, pro- and mesothorax combined c.1.0 mm, forewing 3.3 mm. Head distinctly transverse, 1.6 times as wide as maximum length, 1.4 times as wide



**Text-figure 43** (A) *Aspicolpus temporalis* sp. nov., holotype, NHMUK I.9935. (B) *Diospilus acourti* Cockerell, holotype NHMUK I.9071. Scale bars = 1 mm.

as mesosoma. Head behind eyes rather strongly and roundly narrowed, transverse diameter of eye almost as long as temple. Occipital carina distinct, fused below with hypostomal carina distinctly above mandibular base. Malar suture present. Malar space short, 0.4 times as high as basal width of mandible. Clypeus wide and narrow, with weakly and evenly roundly curved posterior margin, 0.7 times as wide as face. Clypeal suture deep laterally and fine above between tentorial pits. Face smooth. Antenna: scape rather short, 1.85 times as long as maximum width, 2.4 times as long as pedicel. First flagellar segment slender, c. 6.0 times as long as wide apically. Mesosoma: propleura convex below. Prepectal carina distinct and rather highly situated laterally. Sternauli very shallow and wide, possibly smooth. Forewing 2.6 times as long as maximum width. Pterostigma partly deformed, subtriangular, rather wide, 2.2 times as long as maximum width; r arising slightly behind middle of pterostigma and almost perpendicular to it, 0.35 times as long as maximum pterostigmal width, forming distinct obtuse angle with 3-SR. 3-SR rather short, 1.4 times as long as r, 0.6 times as long as 2-SR, 0.9 times as long as r-m; r-m distinct, 0.7 times as long as 2-SR. Second submarginal cell c.1.2 times as long as wide, 0.6 times as long as subdiscal cell along posterior margin. 1-SR+M straight. Discal cell narrowly sessile anteriorly; cu-a postfurcal, distant from basal for 0.4 times cu-a length. Subdiscal cell wide, possibly closed antero-posteriorly by CU1b, c.2.5 times as long as maximum width. 2A distinct, a absent. Hind wing: m-cu absent. M+CU about twice as long as 1-M. SR subparallel to wing anterior margin. Basal cell c.5.0 times as long as wide. Legs: fore femur rather wide, 3.6 times as long as wide. Colour: mesosoma possibly dark brown or black; pterostigma and veins brown.

**Remarks.** The position of this species in *Aspicolpus* is supported with only 2A developed, a long second submarginal cell and 3-SR.



**Text-figure 44** (A, B) *Diospilus curtithorax* sp. nov., holotype, NHMUK I.9286/I.9025: (A) part; (B) counterpart; (C) *Taphaeus cervicalis* (Cockerell), holotype, NHMUK I.9199. Scale bars = 1 mm.

Genus *Diospilus* Haliday, 1833  
*Diospilus acourti* Cockerell, 1921  
 Plate 13, fig. 4; Text-fig. 43B

1921a *Diospilus acourti* Cockerell, p. 12, fig. 12.

**Holotype.** NHMUK I.9071/ I.9184 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; head, mesonotum and wings in dorsal view.

**Diagnosis.** Similar to extant *D. oleraceus* Haliday, 1833, the widely distributed Palearctic parasitoid of beetles. Their distinction is difficult based on the limited information available for the fossil.

**Description.** Combined length of head and mesonotum 0.95 mm, forewing 2.0 mm. Head: subcubical, its width 1.4 times maximum length. Head weakly convex behind eyes, temple length 1.3 times transverse diameter of eye. Mesosoma: mesoscutum wide, its maximum width 1.4 times maximum length, median lobe very weakly rounded anteriorly. Notauli distinct, rather narrow and complete, fused before prescutellar depression. Prescutellar depression rather wide, with medial carina. Forewing 2.4 times as long as maximum width. Pterostigma subtriangular, 2.7 times as long as maximum width. 1-R1 1.2 times as long as pterostigma, 2-R1 short; r arising almost from middle of pterostigma, 0.3 times as long as maximum pterostigmal width. Marginal cell slightly shortened, pointed apically, 2.6 times as long as maximum width. 3-SR 2.8 times as long as r, 0.3 times as long as the straight SR1, 0.7 times as long as 2-SR; r-m present, faintly pigmented, 0.8 times as long as 2-SR. Second submarginal cell weakly widened anteriorly, 1.1 times as long as wide. 1-SR+M straight, 6.5 times as long as 2-SR+M. 2-SR+M 0.2 times as long as 2-SR, 0.3 times as long as m-cu, 0.2 times as long as 2-M. Discal cell shortly sessile anteriorly, as long as wide; m-cu strongly ante-furcal; cu-a shortly postfurcal. Subdiscal cell wide, closed



**Text-figure 45** *Hellenius ? kozlovi* sp. nov., holotype, NHMUK In.17298. Scale bar = 1 mm.

apico-posteriorly by CU1b, 2.5 times as long as width. CU1a arising from posterior 0.3 of distal margin of subdiscal cell, rather distinctly sclerotised in basal c.0.3. 3-CU1 distinctly curved. a present, but very indistinct. Hind wing: sub basal cell large; M+CU c. 0.5 times as long as 1-M and 1.8 times as long as m-cu. Colour: head, almost entirely, and mesonotum black; wings hyaline; pterostigma and veins brown.

*Diospilus curtithorax* Belokobylskij, sp. nov.  
 Plate 13, figs 5, 6; Text-fig. 44A, B

**Etymology.** From *curtus*, the Latin for short, and thorax, referring to form of mesosoma.

**Holotype.** NHMUK I.9286/ I.9025 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; lateral aspect of insect with incomplete appendages and metasoma.

**Diagnosis.** Very similar to *D. acourti*, but differs by having the second submarginal cell not widened anteriorly, 2-SR+M short and the mesosoma very short and high.

**Description.** Length of mesosoma 1.4–1.5 mm, forewing c. 3.0 mm. Head: occipital carina distinct. Eye medium size, 1.4 times as high as broad. Malar space short, 0.2 times as high as longitudinal diameter of eye, 0.7 times as high as basal width of mandible. Antenna: scape about as long as wide. Pedicel rather long. Subapical segments 1.4–1.7 times as long as maximum width. Mesosoma short and high, 1.1–1.15 times as long as high. Mesoscutum highly and roundly elevated above pronotum, with deep notauli at least anteriorly. Scutellum weakly convex. Mesopleuron about twice as high as long, with deep, curved, rather narrow, long and crenulated sternaulus. Prepectal carina present. Mesopleural suture distinctly crenulated anteriorly. Propodeum largely irregularly areolate at least in posterior half. Forewing: pterostigma triangular, wide and short, 2.5 times as long as maximum width. Marginal cell shortened; r arising slightly behind middle of pterostigma, 0.25 times as long as maximum pterostigmal width,

forming very obtuse angle with 3-SR. 3-SR 4.0 times as long as r, 0.7 times as long as 2-SR; r-m present, faintly pigmented, almost parallel to 2-SR, 0.75 times as long as 2-SR, almost as long as 3-SR. Second submarginal cell not widened anteriorly, subparallel-sided, 1.3 times as long as wide, rather narrow. 1-SR+M almost straight, 8.5 times as long as 2-SR+M. 2-SR+M 0.15 times as long as 2-SR, 0.2 times as long as 2-M. Discal cell shortly but distinctly sessile anteriorly, almost as long as wide; m-cu antefurcal; cu-a shortly postfurcal, distant from 1-M for 0.3 times cu-a length. Subdiscal cell closed antero-posteriorly. 2A and a present, but very indistinct. Legs: Middle coxa rather large. Metasoma wide. Colour: head possibly black; mesosoma brown to black; metasoma brown; wings hyaline; pterostigma and veins dark brown.

Genus *Hellenius* Tobias, 1982

**Remarks.** The recently described Palaearctic genus *Hellenius* (Tobias 1982) includes three extant species. The hosts of this genus are unknown yet, but according to their position in Helconinae they are possibly beetle larvae. This is first fossil record of the genus.

*Hellenius* ? *kozlovi* Belokobylskij, sp. nov.  
Plate 13, fig. 7; Text-fig. 45

**Etymology.** After the late Professor Mikhail A. Kozlov, distinguished Russian hymenopterist.

**Holotype.** NHMUK In.17298, Bembridge Marls, NW Isle of Wight, UK, Smith Collection; lateral view of insect lacking antennae and most of head and legs, and with somewhat incomplete wings.

**Diagnosis.** Differs from extant ones in having marginal cell not shortened and with straight SR1, and rather long pterostigma.

**Description.** Length of mesosoma 1.2 mm, meso- and metasoma combined 2.8 mm, forewing 3.1 mm. Mesosoma about 1.5 times as long as high. Mesoscutum distinctly and roundly elevated above pronotum. Notauli deep and possibly complete. Subalar depression rather deep and wide. Sternauli deep and wide. Prepectal carina possibly present. Propodeum at least partly with coarse areolate-reticulate sculpture. Forewing 2.6 times as long as maximum width. Pterostigma subtriangular, rather wide, 3.3 times as long as maximum width; r arising distinctly behind middle of pterostigma, oblique in respect to it, 0.65 times as long as maximum pterostigmal width, forming very obtuse angle with 3-SR. 3-SR rather short, almost as long as r, 0.15 times as long as SR1, 0.65 times as long as 2-SR, almost as long as r-m; r-m distinct, 0.65 times as long as 2-SR. Second submarginal cell small, 1.6 times as long as wide. 1-SR+M almost straight. Discal cell long petiolate anterior, 1.7 times as long as maximum width; m-cu strongly antefurcal, almost as long as 2-SR+M; cu-a postfurcal, distant from 1-M for half cu-a length. Subdiscal cell wide, closed antero-posteriorly by CU1b. 2A unsclerotised but rather distinct. Hind wing: 1r-m as long as 2-SC+R. Marginal cell narrow; SR subparallel to anterior margin of wing. Hind femur rather narrow. Metasoma: first tergite rather long, about 0.9 times as long as second and third tergites combined, following segments rather weakly protruding behind third tergite. Colour: mesosoma almost entirely black; metasoma black dorsally, brown ventrally; forewing faintly infusate; pterostigma and veins dark brown.

**Remarks.** Wing venation is practically the same as in extant *Hellenius*, including petiolate discal cell, small second sub-

marginal cell, and strongly antefurcal m-cu. Unfortunately, the comparison in structure of meso- and metasoma is not possible because of poor preservation.

Genus *Taphaeus* Wesmael, 1835

*Taphaeus cervicalis* (Cockerell, 1921), comb. nov.

Plate 13 fig. 8; Text-fig. 44C

1921a *Sigalphus* (?) *cervicalis* Cockerell, p. 15, fig. 15

**Holotype.** NHMUK I.9199, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; dorsal aspect of insect with incomplete antennae, right wings and fragmentary legs.

**Diagnosis.** The species is similar to extant *T. hiator* (Thunberg, 1822) but differs in having discal cell widely sessile, 3-SR short, cu-a distinctly postfurcal, and first tergite narrow and long. Differs from *T. praecox* Brues, 1923 from the Baltic amber in having forewing faintly infusate, temple short, and r arising from middle of pterostigma. Differs from *T. longicornis* (Statz, 1936), comb. nov. from Rott in having discal cell widely sessile, marginal cell weakly shortened, and m-cu distinctly antefurcal.

**Description.** Length of body 1.8 mm, mesosoma 0.65 mm, metasoma 0.75 mm, forewing 1.6 mm. Head weakly transverse, distinctly widened behind eyes, 1.3 times as wide as long medially, 1.35 times as wide as maximum length. Eye 2.2 times as long as temple in dorsal view, measured along straight line. Mesosoma rather long and slender, c. 3.0 times as long as maximum width. Notauli present, complete, rather narrow. Forewing 2.7 times as long as maximum width. Pterostigma subtriangular, 3.3 times as long as maximum width. 1-R1 about as long as pterostigma, 5.2 times as long as 2-R1, 3.5 times as long as distance from apex of marginal cell to apex of wing; r arising before middle of pterostigma (internal distance of pterostigma between parastigma and r 0.85 times internal distance between r and apex of pterostigma), not strictly perpendicular to pterostigma, 0.55 times as long as maximum pterostigmal width. Marginal cell shortened, pointed apically, 4.0 times as long as maximum width. 3-SR short, 0.4 times as long as r, 0.05 times as long as the straight SR1, 0.25 times as long as 2-SR; r-m present, strongly desclerotised, 0.8 times as long as 2-SR. Second submarginal cell about as long as wide, 0.7 times as long as subdiscal cell. 1-SR+M straight. 2-SR+M almost as long as r, 0.6 times as long as 2-SR, 0.7 times as long as m-cu, 0.6 times as long as sclerotised part of 2-M; m-cu 0.7 times as long as 2-SR, 0.85 times as long as sclerotised part of 2-M. Discal cell distinctly sessile anterior, 1.1 times as long as wide; m-cu strongly antefurcal; cu-a distinctly postfurcal, distant from 1-M for 0.65 times cu-a length. Subdiscal cell wide, closed antero-posteriorly by CU1b. CU1a arising from posterior 0.3 of distal side of subdiscal cell. Hind wing: m-cu absent; M+CU about twice as long as 1-M. Metasoma egg-shaped, rather wide, 1.7 times as long as maximum width. First tergite narrow, with distinct spiracular tubercles in basal 0.3, 1.6 times as long as wide at level of spiracular tubercles, 2.3 times as long as minimum width before tubercles. Colour: head and mesosoma brown; metasoma light brown; wings faintly infusate; pterostigma entirely pale brown; veins brown.

**3.9.9. Subfamily Homolobinae Achterberg, 1979.** The koinobiont braconid wasps of the subfamily Homolobinae, parasitoids of lepidopteran larvae, have not been previously recorded as fossils (Achterberg 1979). Members of this subfamily are absent even from the well known Baltic amber assemblage.



**Text-figure 46** *Homolobus rasnitsyni* sp. nov., holotype, NHMUK In.20545. Scale bar = 1 mm.

Genus *Homolobus* Foerster, 1862  
*Homolobus rasnitsyni* Belokobylskij, sp. nov.  
 Plate 13, fig. 9; Text-fig. 46

**Etymology.** In honor of A.P. Rasnitsyn, Russian palaeontologist.

**Holotype.** NHMUK In.20545, Bembridge Marls, NW Isle of Wight, UK, Smith Collection; lateral aspect of partial head with antennal bases, mesosoma with incomplete forewings, mid and hind legs, and metasomal base.

**Diagnosis.** Characteristic of the species are eyes and possibly ocelli enlarged, discal cell shortly petiolate, *r* arising distinctly behind middle of pterostigma, *cu-a* strongly proclined, and hind tibia distinctly widened apically.

**Description.** Mesosoma length 2.3 mm, forewing 4.9 mm. Head: eye large. Possibly enlarged ocelli present. Scape rather large. Mesosoma: length 1.7 times height. Oblique depression on sides of pronotum rather narrow and crenulated. Notauli very deep and complete. Scutellum almost flat. Subalar depression deep and narrow. Propodeum distinctly rugose-reticulate at least in posterior half. Wing: pterostigma subtriangular, wide, rather long, 3.3 times as long as maximum width. Marginal cell weakly shortened, 2.4 times as long as maximum width. 1-R1 1.2 times as long as pterostigma; *r* arising distinctly behind middle of pterostigma (internal distance of pterostigma between parastigma and *r* 1.5 times internal distance between *r* and apex of pterostigma), forming very obtuse angle with 3-SR, 0.75 times as long as maximum pterostigmal width. 3-SR 1.3 times as long as *r*, 0.25 times as long as SR1, 0.85 times as long as 2-SR, 1.3 times as long as *r-m*. Second submarginal cell short, narrowed posteriorly, 1.2 times as long as maximum width, about 0.7 times as long as subdiscal cell; *m-cu* distinctly antefurcal, almost twice as long as 2-SR+M, almost as long as 2-SR. 1-SR+M weakly S-shaped. Discal cell very shortly petiolate anteriorly, 1.2 times as long as width. 1-M and *m-cu* more or less distinctly convergent

posteriorly. Subdiscal cell wide, closed posteriorly, its length along posterior margin 2.3 times maximum width. CU1a almost entirely sclerotised, arising from posterior 0.2 of distal side of subdiscal cell; *cu-a* strongly inclined posteroapically, subparallel to 3-CU1, postfurcal, distance from 1-M is 0.3 times *cu-a* length. Basal part of 1-1A straight. 2A present. Legs: middle coxa of medium size, weakly elongate; hind femur rather wide; middle and hind tibiae widened posteriorly, with long and pointed spurs; hind basitarsus thickened. Metasoma: first tergite long, with more or less distinct spiracular tubercle in basal 0.2, with sharp lateral border. Colour: mesosoma light reddish brown, propodeum darker; first metasomal tergite dark brown; pterostigma possibly brown, veins dark brown.

**Remarks.** Subgeneric position of the new species is not clear because of insufficient preservation: structure of hind wing, tarsal claws, antennal segments and ovipositor is unknown, as well as some details concerning the head, mesosoma and metasoma.

**3.9.10. Subfamily Microgastrinae Foerster, 1862.** The subfamily Microgastrinae is one of the most specialised braconid groups. The extant members are exclusively koinobiont parasitoids of larvae of various Lepidoptera. Mason (1981) has made a reclassification of the former large genus *Apanteles* Foerster, 1862 and erected several additional genera. Unfortunately the characters used for their diagnoses are rarely available for rock fossils, so I have used the old nomenclature from Nixon (1965).

Only a few fossil microgastrine taxa are described so far, although they are among the most abundant parasitoids in Recent biocenoses. Two fossil species of *Apanteles* are described by Statz (1938). The attribution of *A. macrophthalmus* Statz, 1938 looks convincing, unlike that of another species *A. concinna* Statz, 1938 which shows a distinct *r-m* on forewing and so should be transferred elsewhere, possibly to *Microplitis* Foerster, 1862. The latter genus has two described fossil species, *Microplitis elegans* Timon-David, 1944 from the Oligocene of Camoins in France (Timon-David 1944) and *M. vesperus* Brues, 1910 from the Florissant. *M. vesperus* has a distinctly enlarged second submarginal cell (cf. Brues 1910, p. 102, fig. 85) suggesting a different generic position (possibly in *Eocardiophiles* Brues, 1933 or *Semionis* Nixon, 1965). The status of *Microgaster primordialis* Brues, 1906 from the Florissant also needs verification, because of the peculiar shape and size of its second submarginal cell.

Several described fossil genera have distinctly or strongly elongate 3-SR and correspondingly enlarged second submarginal cell. Described in Cardiochilinae is *Eocardiophiles* Brues, 1933 (with type species *E. fritschii* Brues, 1933) later transferred to Microgastrinae (Mason 1981). This genus, together with *Semionis* Nixon (including Eocene *Semionis nixonii* Tobias, 1987) and New World *Pelicope* Mason, 1981, now form the tribe Semionini (Tobias 1987). The similar shape of the second submarginal cell has been also indicated for *Oligoneuroides* Brues, 1910 (type species *O. destructus* Brues), described in Microgastrinae from the latest Eocene of the Florissant (Brues 1910). However, the number of antennal segments (25 or 26) recorded for this genus strongly testifies against Microgastrinae and suggests rather in favour of the subfamily Cardiochilinae. The genus *Miracoides* Brues, 1933 (type species *M. proteus* Brues, 1933), described from the Baltic amber in Microgastrinae as well, is very similar to the Recent *Mirax* Haliday, 1833 and should be included in the subfamily Miracinae.

The species *Dacnusites sepultus* Cockerell, 1921 and *D. reductus* Cockerell, 1921 were originally described in the new genus *Dacnusites* Cockerell, 1921 as a member of the subfamily Alysiinae (Dacnusiini) (Cockerell 1921a). The present

study reveals their position as members of the subfamily Microgastrinae (see below).

Three genera and four species of Microgasterinae are currently recorded in the Bembridge Marls of the Isle of Wight.

#### Genus *Dacnuses* Cockerell, 1921

**Type species.** *D. sepultus* Cockerell, 1921 (by original designation).

**Remarks.** This genus was described in the subfamily Alysiinae (Dacnusiini). Re-study of forewings of both constituent species, *D. sepultus* and *D. reductus* Cockerell, 1921, undoubtedly indicates its position in Microgastrinae rather than in Alysiinae.

The enlarged second submarginal cell with distinctly developed 3-SR and closed by r-m, together with large hind coxa, are known in extant genera *Microgaster* Latreille, 1804, *Hygroplitis* Thomson, 1895 and *Paroplitis* Mason, 1981. Unfortunately, the diagnostic characters of these genera (venation of the hind wing, shape and sculpture of three basal metasomal tergites, length and shape of ovipositor sheaths) cannot be seen on the fossils at hand. Therefore I have to retain *Dacnuses* as a morphogenus of Microgastrinae with the above diagnostic characters, until better preserved fossils are available for study.

#### *Dacnuses sepultus* Cockerell, 1921

Plate 14, fig. 1; Text-fig. 47B

1921a *Dacnuses sepultus* Cockerell, p. 19, fig. 21.

1992 *Dacnuses sepultus* Cockerell, 1921: Carpenter, p. 478.

**Holotype.** NHMUK In.17176, Bembridge Marls, NW Isle of Wight, UK, Smith Collection; lateral view of body with incomplete antennae and one forewing.

**Diagnosis.** Very similar to *D. reductus* Cockerell, differs in having 1-SR short and in having discal cell respectively lower.

**Description.** Length of mesosoma 1.05 mm, metasoma c. 1.0 mm, forewing 2.3 mm. Head rather small. Antennae thick, flagellar segments divided medially in two parts by rather distinct constriction; sub-basal segments 2.0–2.2 times as long as maximum width. Mesosoma short and high, about 1.2 times as long as high. Notauli absent. Sternauli present, rather wide, situated in anterior 0.7 of mesopleuron. Forewing 2.2 times as long as maximum width. Pterostigma subtriangular, 2.5 times as long as maximum width. C+SC+R thick. Sclerotised part of 1-R1 almost as long as pterostigma. Marginal cell not shortened; r arising almost from middle of pterostigma, almost perpendicular to it, 0.7 times as long as maximum pterostigmal width, almost twice as long as sclerotised 3-SR, as long as 2-SR. SR1 strongly desclerotised, weakly visible; r-m present, desclerotised, enclosing small second submarginal cell. Second submarginal cell almost as wide as long, subtriangular. 1-SR+M weakly curved, 3.5 times as long as 2-SR+M. 2-SR+M 0.7 times as long as r, 0.7 times as long as 2-SR, 0.65 times as long as m-cu and sclerotised part of 2-M. Discal cell rather shortly petiolate anteriorly, almost as long as wide. 1-M weakly curved, 5.5 times as long as 1-SR; m-cu strongly antefurcal; cu-a strongly postfurcal, distant from 1-M for cu-a length. Subdiscal cell wide, almost twice as long as wide; CU1b indistinct; subdiscal cell open apico-posteriorly because 2-1A desclerotised distally. 2A indistinct. Legs: hind coxa rather large and long, about twice as long as maximum width; hind femur rather wide. Metasoma without carapace. Colour: head, antennae, mesosoma, middle and hind coxa and hind femur black; metasoma brown; forewing almost hyaline; pterostigma almost black; veins brown.

#### *Dacnuses reductus* Cockerell, 1921

Plate 14, fig. 2; Text-fig. 47A

1921a *Dacnuses reductus* Cockerell, p. 19, fig. 22.

**Holotype.** NHMUK In.24338, Bembridge Marls, NW Isle of Wight, UK, Hooley Collection; isolated forewing.

**Diagnosis.** Very similar to the type species, differs in having longer 1-SR and higher discal cell.

**Description.** Forewing 2.0 mm long, almost twice as long as maximum width. Pterostigma subtriangular, 2.6 times as long as maximum width. C+SC+R thick. Sclerotised part of 1-R1 almost as long as pterostigma. Marginal cell not shortened; r arising almost from middle of pterostigma, almost perpendicular to pterostigma, 0.7 times as long as maximum pterostigmal width, 2.5 times as long as sclerotised 3-SR, 0.9 times as long as 2-SR. SR1 much desclerotised but visible; r-m present, not sclerotised, enclosing small second submarginal cell. Second submarginal cell almost as wide as long, subtriangular. 1-SR+M weakly curved, 4.0 times as long as 2-SR+M. 2-SR+M 0.6 times as long as r, 0.5 times as long as 2-SR, 0.55 times as long as m-cu and sclerotised part of 2-M. Discal cell rather long and petiolate anteriorly, 1.1 times as long as wide. 1-M almost straight, 3.2 times as long as 1-SR; m-cu strongly antefurcal; cu-a strongly postfurcal, distance from 1-M 1.2 times cu-a length. Subdiscal cell wide, 1.7 times as long as wide; CU1b present; subdiscal cell open apico-posteriorly because 2-1A desclerotised distally. 2A and a indistinct. Colour: forewing almost hyaline; pterostigma light brown; veins light brown.

#### Genus *Palaeomicrogaster* Belokobylskij, gen. nov.

**Etymology.** After *palaios*, the Greek for ancient, and genus *Microgaster*. Gender masculine.

**Type species.** *Palaeomicrogaster oculatus* sp. nov., by present designation and monotypy.

**Diagnosis.** Similar to *Microgaster* Latreille and related genera, but differs in having eyes very long, distinctly narrowed below and almost reaching mandibular base, and hind wing SR arising from 1r-m. The latter character appears in a few microgastrine taxa including species of *Choeras* Mason; the new genus distinctly differs from all such Microgastrinae in the peculiar eye shape.

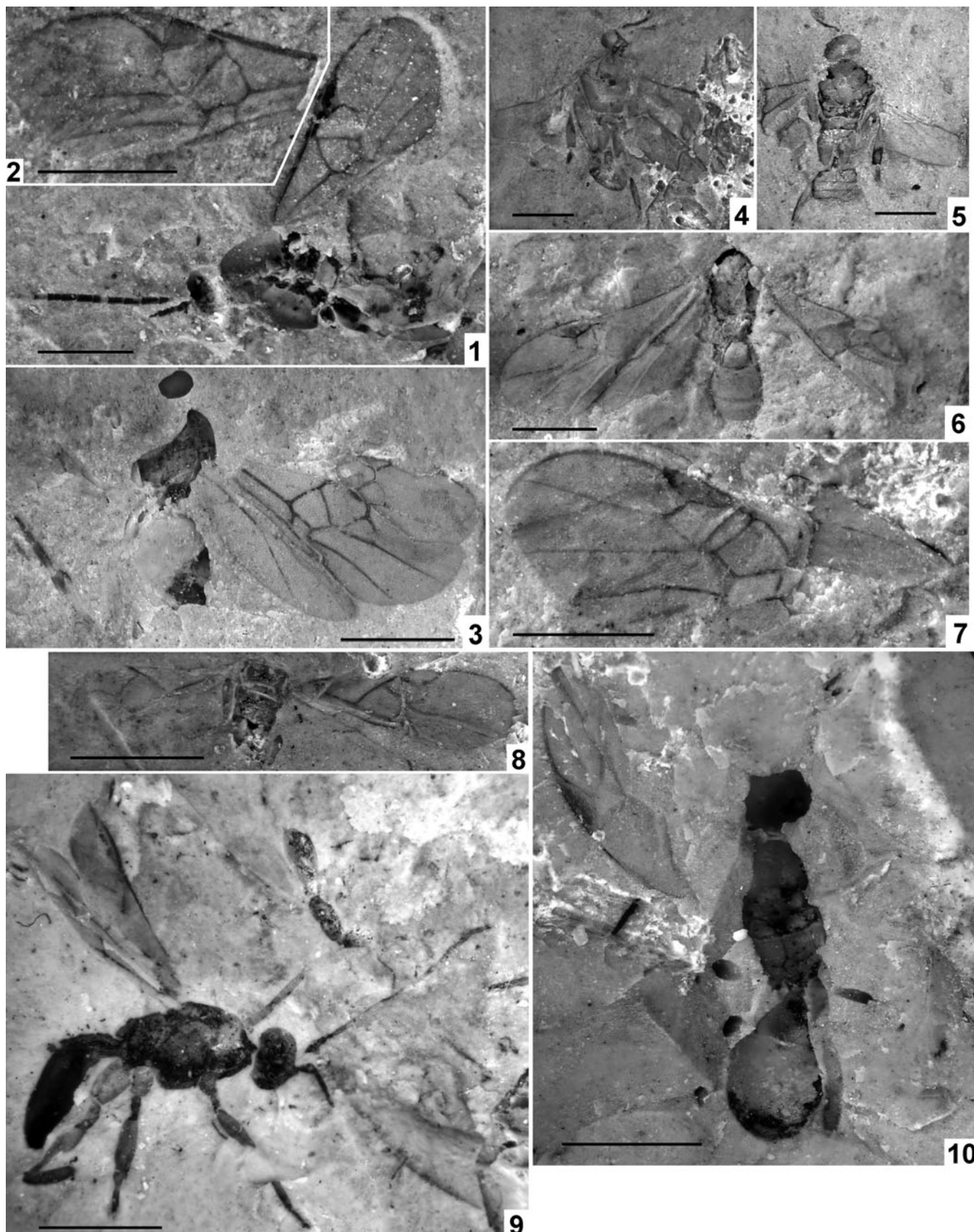
**Description.** Head more or less transverse. Eyes very long, distinctly and regularly narrowed downward and possibly reaching base of mandible. Occipital carina absent. Frons distinctly concave. Ocelli enlarged. Notauli, prepectal carina and sternauli absent. Propodeum without delineated areas; r-m of forewing present, enclosing rather small second submarginal cell. Discal cell petiolate anteriorly; cu-a strongly postfurcal and evenly curved. In hind wing, SR arising from 1r-m, distinctly convex in anterior half and almost straight in posterior half. Marginal cell with distinct desclerotised submedial cross-vein; r-m present enclosing long and rather narrow submarginal cell. Hind coxa large. Hind femur wide; hind tibia slender basally and distinctly widened towards apex. Metasoma short and wide, more or less depressed. First tergite possibly long.

#### *Palaeomicrogaster oculatus* Belokobylskij, sp. nov.

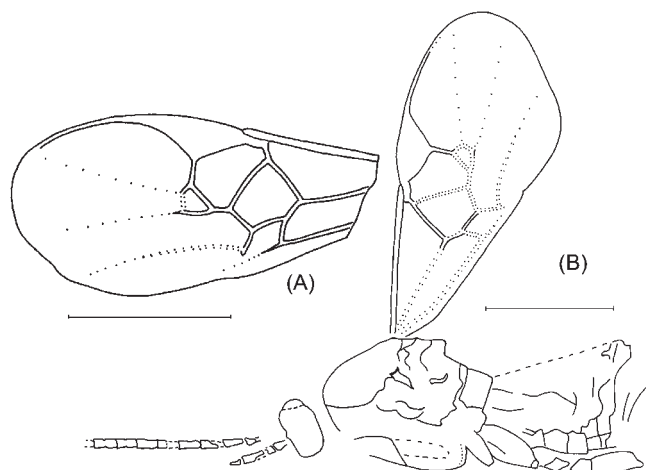
Plate 14, figs 4, 5; Text-fig. 48

**Etymology.** From *oculatus*, the Latin for 'having an eye', referring to peculiar shape of eyes.

**Holotype.** NHMUK I. 9853/I. 10187 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Brodie



**Plate 14** (1) *Dacnusa sepultus* Cockerell, holotype, NHMUK In.17176. (2) *Dacnusa reductus* Cockerell, holotype, NHMUK In.24338. (3) *Semionis wightensis* sp. nov., holotype, NHMUK In.17081. (4–5) *Palaeomicrogaster oculatus* sp. nov., holotype, NHMUK I.9853/I.10187: (4) part; (5) counterpart. (6) *Biosteres peritus* (Cockerell), holotype, NHMUK I.9671. (7) *Microtypus hooleyi* (Cockerell), holotype, NHMUK In.24337. (8) *Oncophanes andrewrossi* sp. nov., holotype, NHMUK I.10252. (9) *Rhyssalus bruesi* sp. nov., holotype, NHMUK I.9468. (10) *Dolopsidea ? intermedia* sp. nov., holotype, NHMUK I.9615. Scale bars = 1 mm.



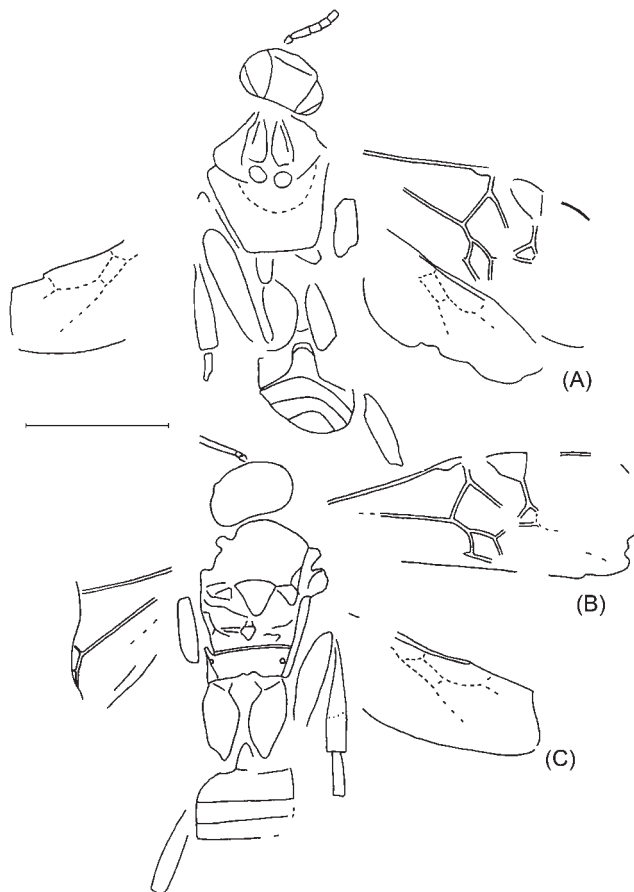
**Text-figure 47** (A) *Dacnusites reductus* Cockerell, holotype, NHMUK In.24338. (B) *D. sepultus* Cockerell, holotype, NHMUK In.17176. Scale bars = 1 mm.

Collection; dorso-ventral aspect of insect with antennae, wings and legs incomplete.

**Description.** Body length 2.7 mm, mesosoma 1.05 mm, metasoma 1.1 mm, forewing *c.* 2.5 mm. Head 2.2 times as broad as median length. Frons with shallow but distinct median longitudinal furrow. Head distinctly and roundly narrowed behind eyes. Transverse diameter of eye in dorsal view about as long as temple. Ocelli subcircular, arranged in triangle with base 1.2 times its side; postocellar line equal to ocellar diameter. Antennae rather thick. Mesosoma short and wide, 1.4 times as long as wide. Lateral sides of pronotum with shallow vertical furrow. Mesosternal suture absent. Mesoscutum 0.8 times as long as maximum width. Prescutellar depression distinct and narrow. Scutellum with no subposterior transverse furrow. Metanotum medially with rather large subcircular area delineated by carinae laterally and distinct convex protuberance posteriorly. Propodeum smooth, with distinct lateral longitudinal carinae and rather large spiracles in basal 0.3–0.4. Forewing 2.2 times as long as maximum width; *r* arising possibly behind middle of pterostigma and almost perpendicular to it, about 0.9 times as long as maximum pterostigmal width, 2.5 times as long as sclerotised 3-SR, 1.2 times as long as 2-SR; *r*–*m* distinctly pigmented. Small second submarginal cell 1.25 times as long as maximum width, rather narrow. 1-SR+M weakly curved. Discal cell rather long and petiolate anteriorly, 1.1 times as long as wide. 1-M weakly curved; *m*–*cu* antefurcal; *cu*–*a* strongly postfurcal, distant from 1-M for 0.9 times *cu*–*a* length, 0.65 times distance from *cu*–*a* to *m*–*cu*. Subdiscal cell wide, 1.6 times as long as wide. Hind wing wide, *c.* 3.0 times as long as maximum width. Submarginal cell 3.2 times as long as wide. Legs: hind coxa about twice as long as wide, 0.7 times as long as hind femur; hind femur 3.4 times as long as maximum width; maximum width of hind tibia 0.7 times maximum width of hind femur. Metasoma without carapace, visible tergites (behind hind coxae) rather short and strongly transverse. Colour: head, antennae, mesosoma and metasoma black; hind tibia black (at least on widened part and base of hind tarsus); hind femur brown; forewing almost hyaline; pterostigma pale with distinctly darkened margins; veins pale brown.

Genus *Semionis* Nixon, 1965

**Remarks.** Wing venation with large second submarginal cell is recorded in three genera of the tribe Semionini Tobias, 1987: extant South African and fossil *Semionis*; extant New



**Text-figure 48** *Palaeomicrogaster oculatus* sp. nov., holotype NHMUK I 9853/I.10187: (A) part; (B) left forewing; (C) counterpart. Scale bar = 1 mm.

World *Pelicope* Mason, 1981; and fossil *Eocardiochiles* Brues, 1933 (Mason 1981; Tobias 1987). Additionally characteristic of these genera is the absence of the hind wing 2*r*–*m*, one of the unique microgastrine features.

A combination of the absence of the hind wing 2*r*–*m* with the enlarged forewing submarginal cell is diagnostic of the worldwide Cardiochilinae Ashmead, 1900 and even African Khoikhoinae Mason, 1983. Unfortunately, the unique fossil under description below does not display several important morphological features (antennae, metasomal tergites, etc.) which could help improve understanding of its taxonomic position.

*Semionis wightensis* Belokobylskij, sp. nov.

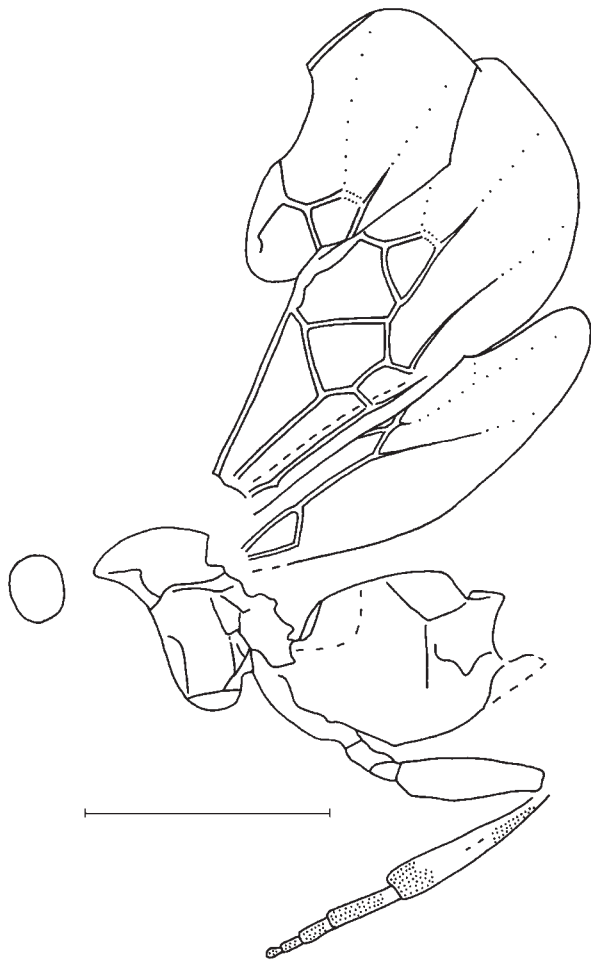
Plate 14, fig. 3; Text-fig. 49

**Etymology.** After the Isle of Wight.

**Holotype.** NHMUK In.17081, Bembridge Marls, NW Isle of Wight, UK, Smith Collection; lateral aspect of head fragment, incomplete mesosoma and wings, hind leg and metasoma.

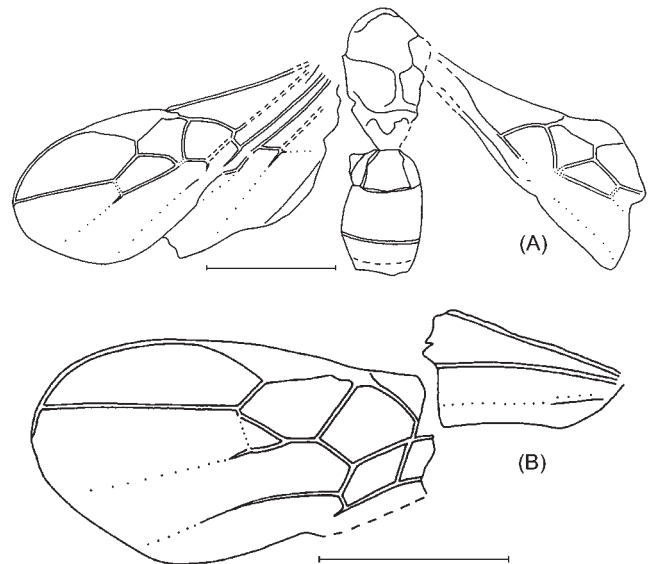
**Diagnosis.** Similar to *S. nixonii* Tobias, 1987 from the Baltic amber, but differs in having *cu*–*a* much more postfurcal, 1-SR long, 3-M almost straight, SR of hind wing subparallel to anterior margin of wing, 3-SR short, and the hind tibia with wide, pale coloration.

**Description.** Length of body *c.* 2.0 mm, mesosoma 0.8 mm, metasoma 0.9 mm, forewing 2.2 mm. Mesosoma 1.2 times as long as high. Mesoscutum highly and roundly elevated above pronotum. Notauli absent. Prescutellar depression very narrow and rather deep. Sternauli shallow and wide. Prepectal carina possibly present. Propodeum possibly entirely with coarse



**Text-figure 49** *Semionis wightensis* sp. nov., holotype, NHMUK In.17081. Scale bar = 1 mm.

areolate-reticulate sculpture. Forewing 2.4 times as long as maximum width. Pterostigma subtriangular, rather wide. Sclerotised part of 1-R1 long, protruding almost to apex of wing; r arising distinctly behind middle of pterostigma and almost perpendicular to it. 3-SR long, 1.4 times as long as r, 0.75 times as long as 2-SR. SR1 not at all sclerotised but visible and distinctly curved; r-m present, desclerotised, enclosing rather large second submarginal cell. Second submarginal cell 1.5 times as long as wide. 1-SR+M straight, 4.5 times as long as 2-SR+M; m-cu almost twice as long as 2-SR+M, 0.7 times as long as 2-SR, 1.3 times as long as r. Discal cell long petiolate anterior, 1.3 times as long as wide. 1-M very weakly curved, 3.5 times as long as 1-SR; m-cu strongly antefurcal. Cu-a strongly postfurcal, distant from 1-M for almost cu-a length, 0.6 times distance from cu-a to m-cu. Subdiscal cell wide, 1.7 times as long as width; CU1b absent. 2A present but very strongly desclerotised. CU1a and 3-M distinctly pigmented. Hind wing 4.0 times as long as wide. Basal cell narrow. Sub basal cell rather large; M+CU almost equal to 1-M; cu-a weakly S-shaped. 2-M and SR distinctly divergent, without 2r-m. SR subparallel to anterior margin of wing; marginal cell with indistinct oblique cross-vein submedially. Legs: hind coxa of medium size; hind femur 3.8 times as long as wide; hind tibia distinctly widened, c.5.0 times as long as wide, 1.2 times as long as hind femur. Metasoma wide and rather short. Colour: head, most part of mesosoma and metasoma, at least in posterior third, almost black; hind leg pale; hind tibia distinctly infusate in apical 0.25; hind tarsus dark brown; forewing faintly infusate; pterostigma entirely brown; veins dark brown.



**Text-figure 50** (A) *Biosteres peritus* (Cockerell), holotype, NHMUK I.9671. (B) *Microtypus hooleyi* (Cockerell), holotype, NHMUK In.24337. Scale bars = 1 mm.

**3.9.11. Subfamily Opiinae Foerster, 1862.** The members of Alysinae and Opiinae, parasitoids of various families of Diptera, are very rare as fossils. Nine species of four genera (*Alysia exigua* Brues, 1910, *A. latifrons* Statz, 1938, *A. meunieri* Théobald, 1937, *A. petrina* Brues, 1910, *A. ruskii* Cockerell, 1913, *Pentapleura* (?) *filicornis* Statz, 1938, *P. longipennis* Statz, 1938, *Tanycarpa interstitialis* Statz, 1938, and *Aspilota* (?) *stigmatalineata* Statz, 1938) were described only as fossils from Florissant, Rott and Aix-en-Provence (Brues 1910; Cockerell 1913; Théobald, 1937, Statz 1938). Unfortunately, the above alysiine fail to show the main alysiine character – shape and position of mandibles. Identification of Alysinae based on other characters is difficult and deserve verification. Two species of *Dacnusites* described as a member of the tribe Dacnusiini (Alysinae) (Cockerell 1921a) are found to belong to the subfamily Microgastrinae (see above).

Subfamily Opiinae has the only fossil record *Bathystomus* (?) *peritus* Cockerell, 1921. Although several important opiine characters cannot be seen in the unique fossil, evaluation of those available (absence of prepectal carina and presence of hind wing m-cu, as well as some details of forewing venation) would agree with its position in Opiinae but in a different genus, *Biosteres* Foerster, 1862.

Genus *Biosteres* Foerster, 1862

*Biosteres peritus* (Cockerell, 1921), comb. nov.

Plate 14, fig. 6; Text-fig. 50A

1921a *Bathystomus peritus* Cockerell, p. 16, fig. 17.

**Holotype.** NHMUK I.9671, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; dorsal view of headless body with no appendages but partially incomplete wings.

**Diagnosis.** Characteristic of the new species are the second submarginal cell rather long and narrowed towards apex, not shortened marginal cell, antefurcal m-cu, wide second metasomal suture, and perhaps the entirely smooth three anterior metasomal tergites.

**Description.** Length of mesosoma and metasoma combined 1.9 mm, mesosoma 0.95 mm, metasoma 0.95 mm, forewing 2.4 mm. Mesosoma 1.7 times as long as maximum width. Prepectal carina absent. Forewing 2.5 times as long as maximum

width. Pterostigma subtriangular, wide, almost 3.0 times as long as maximum width. 1-R1 1.3 times as long as pterostigma. Marginal cell not shortened, pointed apically, 2.5 times as long as maximum width; r arising behind middle of pterostigma (internal distance of pterostigma between parastigma and r 0.6 times internal distance between r and apex of pterostigma), 0.7 times as long as maximum pterostigmal width. 3-SR twice as long as r, 0.4 times as long as weakly S-shaped SR1, 0.8 times as long as 2-SR; r-m present and weakly sclerotised, 0.3 times as long as 2-SR. Second submarginal cell 2.3 times as long as wide, 1.6 times as long as subdiscal cell. 1-SR+M distinctly curved; m-cu 0.6 times as long as 2-SR and 1-SR+M. Discal cell distinctly petiolate anterior, 1.4 times as long as width; m-cu antefurcal; cu-a postfurcal. Subdiscal cell rather wide, about 3.0 times as long as wide. 2A and a probably absent. Basal cell of hind wing distinctly widened distally; m-cu present, weakly postfurcal, almost straight. Metasoma: first tergite distinctly and slightly roundly widened towards apex, with clear spiracular tubercle in basal 0.3; length of tergite about equal to its apical width; apical width c.2.5 times its basal width. Second tergite 0.8 times as long as basal width, 1.4 times as long as visible third tergite. Combined median length of second and third tergites almost equal to its maximum width, 1.3 times basal width of second tergite. Colour: mesosoma brown; metasoma yellow; wings almost hyaline; pterostigma brown, pale basally and possibly apically; veins brown.

**Remarks.** Position of this species in the genus *Biosteres* is not definite, and additional material might help to understand the real status of this taxon. It is not easy to find related species from extant taxa of this genus.

**3.9.12. Subfamily Orgilinae Ashmead, 1900 (= Microtypinae Szepilgeti, 1908).** Subfamily Orgilinae (including tribe Microtypini which some authors consider as a separate subfamily, Achterberg 1993) is a small group of koinobiont braconids, endoparasitoids of lepidopteran larvae, widely distributed on almost all continents.

Only three species of *Orgilus* Haliday, 1833, recently transferred to this genus from *Blacus* Nees, 1818 (Achterberg 1982), are known as fossils: *O. ashmeadii* (Brues 1933), *O. grandior* (Brues 1933) and *O. longicornis* (Brues 1933). The majority of the fossil Orgilinae were described in *Microtypus* Ratzeburg, 1848 (Brues 1933, 1939). Because the morphological limits of this genus in Brues' view were very broad, only three species (*M. brevicornis* Brues, 1933, *M. triangulifer* Brues, 1933 and *M. verticalis* Brues, 1933) out of ten described by him from Baltic amber had the real *Microtypus*-type triangular second submarginal cell and only two radial abscissae r and SR1+3-SR (see also discussion for subfamily Helconinae).

Cockerell described the new genus and species *Diospiloides hooleyi* Cockerell, 1921 from the Bembridge Marls and considered it as possibly related to Diospilini (Helconinae). Re-study of the type has shown it belongs to the genus *Microtypus* (see below). Thus, *Diospiloides* Cockerell, 1921 is a junior synonym of *Microtypus* Ratzeburg, 1848.

Only one genus and species of the subfamily Orgilinae is recorded in the Bembridge Marls of the Isle of Wight.

Genus *Microtypus* Ratzeburg, 1848

= *Diospiloides* Cockerell (type species *D. hooleyi* Cockerell, 1921), syn. nov.

*Microtypus hooleyi* (Cockerell, 1921), comb. nov.

Plate 14, fig. 7; Text-fig. 50B

1921a *Diospiloides hooleyi* Cockerell, p. 14, fig. 14.

1992 *Diospiloides hooleyi* Cockerell, 1921: Carpenter, p. 478.

**Holotype.** NHMUK In.24337, Bembridge Marls, NW Isle of Wight, UK, Hooley Collection; isolated forewing.

**Diagnosis.** Very similar to *M. brevicornis* Brues, 1933 but differs in having r arising distinctly behind middle of pterostigma, m-cu distinctly longer than 2-SR+M, and cu-a less strongly postfurcal.

**Description.** Forewing length c.3.0 mm; 2.8 times as long as maximum width. Pterostigma subtriangular, 3.5 times as long as maximum width. 1-R1 1.75 times as long as pterostigma, 8.2 times as long as 2-R1. Marginal cell not shortened, pointed apically, 3.6 times as long as maximum width; r arising distinctly behind middle of pterostigma (internal distance of pterostigma between parastigma and r 1.4 times internal distance between r and apex of pterostigma), 1.1 times as long as maximum pterostigmal width, 0.2 times as long as 3-SR, 0.65 times as long as 2-SR; r-m present, strongly desclerotised, enclosing second submarginal cell. Second submarginal cell 1.5 times as wide as long, narrow, triangular, shortly petiolate anterior. 1-SR+M very weakly S-shaped, about twice as long as 2-SR+M. 2-SR+M 0.9 times as long as r, 0.6 times as long as 2-SR, 0.7 times as long as m-cu, 0.6 times as long as sclerotised part of 2-M. Discal cell shortly sessile anteriorly, 1.4 times as long as wide; m-cu strongly antefurcal; cu-a very shortly postfurcal. Subdiscal cell wide, 2.3 times as long as wide; CU1b distinct, closing subdiscal cell. CU1a arising from posterior 0.3 of distal margin of subdiscal cell, distinctly sclerotised on about basal half of its length. 2A and a indistinct. Colour: forewing hyaline; pterostigma very pale, narrowly brown marginally and almost black shortly anteriorly; veins mostly pale brown and partly brown.

**3.9.13. Subfamily Rhyssalinae Foerster, 1862.** The members of subfamily Rhyssalinae were for a long time included in other subfamilies, such as Rogadinae and Exothecinae s.l. This subfamily was re-instated by Quicke & Achterberg (1990), after a special study of the inner structures of the ovipositor and venom apparatus, and now includes the genera *Rhyssalus* Haliday, 1833, *Oncophanes* Foerster, 1862, *Dolopsidea* Hincks, 1944, *Pseudobathystomus* Belokobylskij, 1986 and possibly several subtropical and tropical genera (*Doryctomorpha* Ashmead, 1900, *Caenopachyella* Szepilgeti, 1908, *Metaspathius* Brues, 1922).

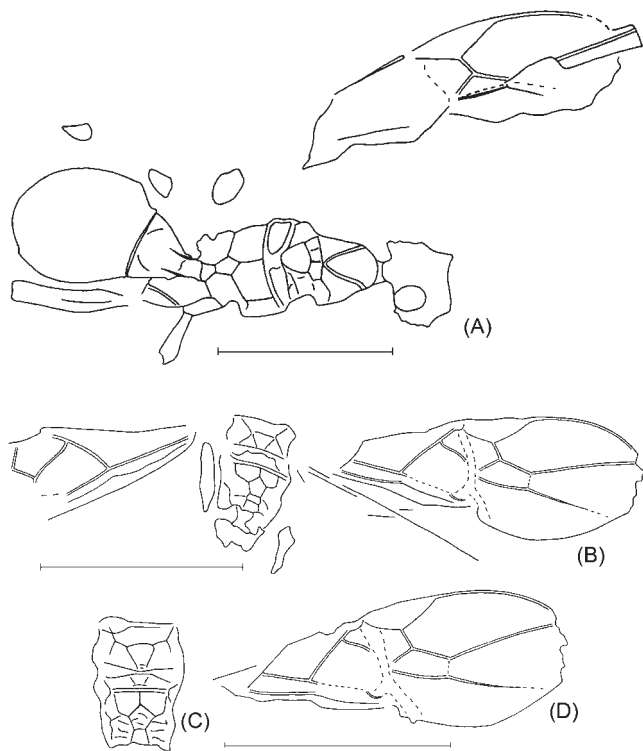
Two genera of this subfamily were described from the Baltic amber as Rogadinae (Brues 1933). Descriptions and figures of both species of *Rhyssalus* (*R. brevicornis* Brues, 1933 and *R. rugosus* Brues, 1933) show their position to be unequivocally not of this genus. This opinion is supported by the sculptured second and third metasomal tergites, the granulate sculpture of mesoscutum, the reduced number of antennal segments and, for the second species only, not clavate male hind tibia. If really cyclostomate braconids, these species are possibly members Exothecinae (because of areolate propodeum). The same taxonomic position should also include *Palaeorhyssalus* Brues, 1933 (with *P. dubitosus* Brues, 1933) from the Baltic amber. At the same time, *Diospilus allani* Brues, 1937 from the Upper Cretaceous Canadian amber is very similar venationally to the extant *Pseudobathystomus* Belokobylskij, or some species of *Oncophanes* Foerster (see discussion above under Helconinae).

New species of *Dolopsidea*, *Rhyssalus* and *Oncophanes* described below are the first reliable members of Rhyssalinae in the fossil record.

Genus *Dolopsidea* Hincks, 1944

*Dolopsidea ? intermedia* Belokobylskij, sp. nov.

Plate 14, fig. 10; Text-fig. 51A



**Text-figure 51** (A) *Dolopsidea* ? *intermedia* sp. nov., holotype, NHMUK I. 9615. (B–D) *Oncophanes andrewrossi* sp. nov., holotype, NHMUK I.10252. Scale bars = 1 mm.

**Etymology.** After *intermedius*, the Latin for intermediate, referring to a position of this species between several subfamilies.

**Holotype.** NHMUK I.9615, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; dorsal aspect of incomplete head, meso- and metasoma with deformed forewing and fragmentary legs.

**Diagnosis.** Differs from the extant species *D. indagator* (Haliday, 1836) in having the propodeum smooth between carinae, the first metasomal tergite short and wide, and the mesoscutum distinctly rounded anteriorly.

**Description.** Length of body 2.5 mm, mesosoma 1.0 mm, metasoma 1.1 mm, forewing *c.* 2.0 mm. Head rather transverse, distinctly and roundly narrowed behind eyes. Eyes rather large. Ocelli small, arranged in triangle with base 1.3 times its sides. Distance between antennal sockets almost equal to distance between socket and eye. Frons more or less distinctly concave, without medial furrow or keel. Occipital carina complete dorsally. Mesosoma 1.8 times as long as maximum width. Median lobe of mesoscutum weakly protruding forwards and rounded anterior. Notauli complete, shallow particularly posteriorly, fused with each other on border of prescutellar depression. Prescutellar depression wide, deep, without medial carina, weakly sculptured. Scutellum convex, rather short, without transverse depression posteriorly. Sternauli shallow and wide, almost smooth, situated in anterior 0.6 of mesopleuron. Metanotum with fine medial carina. Propodeum more or less short, almost entirely smooth, with rather distinct and wide lateral tubercles, with distinct areas delineated by high carinae, basal carina 1.7 times as long as anterior fork, areola 1.6 times as long as width, petiolate area rather long and separated from areola by distinct carina. Forewing: pterostigma 2.6 times as long as maximum width. 1-R1 1.3 times as long as pterostigma. Marginal cell not shortened, pointed apically, 2.7 times as long as maximum width; r arising from middle of pterostigma, 0.8 times as long as maximum pterostigmal width. 3-SR 1.7 times as long

as r, 0.3 times as long as SR1, about 1.5 times as long as 2-SR; m-cu shortly antefurcal. Legs: hind coxa without dorsal processes; hind femur rather wide. Metasoma wide, 1.8 times as long as maximum width, 1.1 times as long as mesosoma. First tergite moveably connected with second tergite, short and wide, distinctly convex and rugulose-striate mostly on wide and subcircular area, regularly and strongly widened towards apex, with high and weakly convergent dorsal carinae in basal 0.3 of tergite, dorsope small; length of tergite 0.9 times its apical width, apical width about 3.0 times its basal width. Second suture absent. Second and third tergites strongly sclerotised, smooth, their combined median length about 1.8 times basal width of second tergite and 1.25 times their maximum width. Colour: body entirely black; hind femur possibly brown; forewing faintly infusate; pterostigma and veins brown.

**Remarks.** With the limited information available, the species is tentatively included in the genus *Dolopsidea* Hincks, based on the forewing with the r-m and second submarginal cell, mesoscutum with notauli, propodeum with distinct peculiar areolation, short first tergite with basal carinae, and absence of the second metasomal suture.

Genus *Oncophanes* Foerster, 1862

*Oncophanes andrewrossi* Belokobylskij, sp. nov.

Plate 14, fig. 8; Text-fig. 51B–D

**Etymology.** In honour of Dr Andrew Ross, palaeontologist at National Museums Scotland, Edinburgh.

**Holotype.** NHMUK I.10252, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; dorsal aspect of incomplete mesosoma and wings with leg fragments.

**Diagnosis.** Differs from the type species *O. minutus* (Wesmael, 1838) in having 1-R1 much longer than pterostigma, forewing infusate, propodeal areola less narrowed posteriorly, and possibly the m-cu postfurcal.

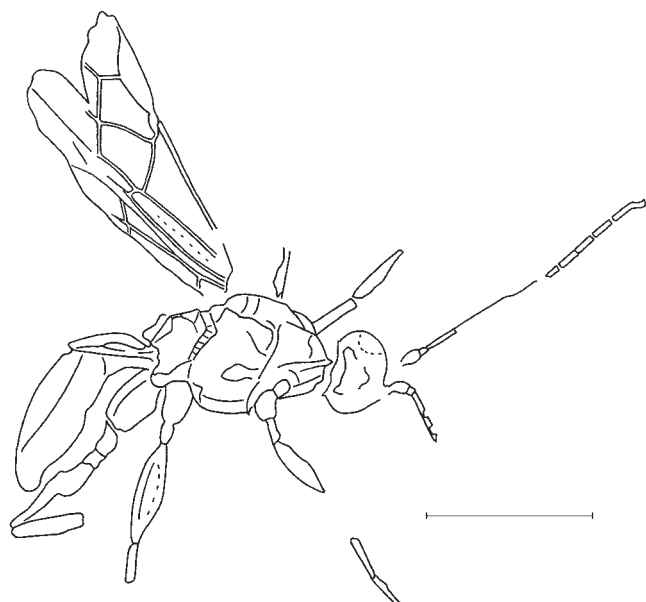
**Description.** Length of forewing 1.6 mm. Mesosoma: scutellum about as wide as median length. Metanotum medially with two carinae convergent posteriorly and fused with transverse posterior protuberance. Propodeum distinctly areolated with carinae. Areola wide, pentagonal, short, 1.2 times as long as wide, 1.6 times as long as petiolate area, distinctly separated from wide and long petiolate area by carina, basilateral areas rather short and almost smooth, other parts of propodeum rugose-reticulate. Wings: forewing 2.8 times as long as wide. Pterostigma subtriangular, wide, small, about 3.0 times as long as maximum width. Marginal cell not shortened, 1-R1 1.6 times as long as pterostigma; r long, almost as long as maximum width of pterostigma, arising almost from middle of pterostigma. 3-SR 1.5 times as long as r, 0.3 times as long as the straight SR1, about 1.5 times as long as 2-SR and r-m. Second submarginal cell rather wide, 2.4 times as long as wide, almost as long as subdiscal cell; m-cu possibly postfurcal, 3.7 times as long as 2-SR+M, 1.3 times as long as 2-SR. 1-SR+M weakly S-shaped. Discal cell distinctly petiolate anteriorly, 1.6 times as long as wide. 1-M and m-cu subparallel, weakly convergent posteriorly. Subdiscal cell narrow. CU1a not interstitial. Colour: mesosoma dark brown to black; wings faintly infusate; pterostigma (possibly) and veins brown.

**Remarks.** Propodeal areolation with long and distinctly delineated petiolate area and forewing venation confirm position of the species in *Oncophanes* despite the poor preservation of the holotype.

Genus *Rhyssalus* Haliday, 1833

*Rhyssalus bruesi* Belokobylskij, sp. nov.

Plate 14, figure 9; Text-fig. 52



**Text-figure 52** *Rhyssalus bruesi* sp. nov., holotype, NHMUK I.9468. Scale bar = 1 mm.

**Etymology.** After C.T. Brues, well-known hymenopterist and researcher of the hymenopteran fauna of Baltic amber.

**Holotype.** NHMUK I.9468, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection; male, lateral aspect of insect with antennae, wings and legs incomplete.

**Diagnosis.** Differs from all extant congeners in having the fore and middle femora thick, the first tergite elongate, *r* arising almost from the middle of pterostigma, and *m-cu* a little antefurcal.

**Description.** Body length 2.6 mm, mesosoma 1.1 mm. Head 1.65 times as high as maximum length. Occipital carina present. Antennae slender, rather long. Scape 1.8 times as long as maximum width, 2.2 times as long as pedicel. First flagellar segment about 5.0 times as long as apical width, 1.1 times as long as second segment. Subapical segment about 3.0 times as long as wide. Mesosoma: length 1.6 times height. Notauli present at least anteriorly and deep. Prepectal carina present. Sternauli rather distinct, more or less short, rather wide. Subalar depression deep and wide. Mesopleural suture crenulated. Basilateral areas of propodeum delineated by carinae. Wings: pterostigma subtriangular, wide, rather long, 3.6 times as long as maximum width; *r* arising almost from middle of pterostigma, forming almost straight line with 3-SR, almost as long as maximum pterostigmal width, about 0.5 times as long as 2-SR; *m-cu* shortly antefurcal, 0.7 times as long as 2-SR. 1-SR+M distinctly S-shaped. Discal cell distinctly petiolate anteriorly, almost twice as long as wide. 1-M and *m-cu* weakly convergent posteriorly. Subdiscal cell rather wide, possibly closed posteriorly, 3.5 times as long as wide. CU1a not interstitial; *cu-a* postfurcal, distant from 1-M almost for *cu-a* length. Hind wing: sub-basal cell shortened; M+CU shorter than 1-M; *m-cu* present and weakly antefurcal. Legs rather robust. Legs: middle femur wide; hind coxa without basiventral corner, 1.4 times as long as wide; visible basal part of hind tibia thickened. Metasoma: first tergite rather long. Colour: head, mesosoma and at least first metasomal tergite almost black; pterostigma and veins dark brown.

**Remarks.** The position of this species in *Rhyssalus* is somewhat unreliable, because of the lack of information about the hypoclypeal cavity and incomplete wing venation. Nevertheless, the thickened hind tibia, areolated propodeum, thick

femora, short sub basal cell of hind wing, and developed *m-cu* support the attribution of this species to *Rhyssalus*.

**3.9.14. Discussion.** The studied material from the Bembridge Marls includes members of many subfamilies previously recorded as fossils. Undoubtedly interesting is the first discovery from this locality of the subfamily Homolobinae.

Living members of most of braconid taxa recorded in the Bembridge Marls, and particularly Agathidinae, Cheloninae, Exothecinae, Homolobinae, Microgastrinae, Orgilinae, partly Rhyssalinae, and possibly Euphorinae (*Meteorus*), are known as parasitoids of various Lepidoptera. This information is interesting, because their lepidopteran hosts are very rare in the Bembridge Marls fossil assemblage. The abundance of lepidopteran parasitoids, both in the Bembridge and Baltic amber assemblages, suggests the same abundance for moths since at least the late Eocene, in spite of their poor fossil record which is apparently due to taphonomic processes. Interesting is the presence, in both assemblages (see Brues 1933 and Tobias 1987 for Baltic amber), of plenty of *Ascogaster*, a specialised egg-larval parasitoid of micro-Lepidoptera, and particularly the species of very small size indicative of their development at the expense of leaf miners (for example, *A. pinicola* Brues or *A. pygmaea* sp. nov.). Microlepidopteran hosts, mainly of the families Tortricidae, Gelechiidae and Pyralidae, are known also for *Bassus*, *Microtypus*, *Chremylus* (as well as some Coleoptera: Shenefelt 1975), *Oncophanes* and possibly Microgastrinae genera. However, the latter group is known to prefer to parasitise gregarious macrolepidopteran caterpillars. Equally *Homolobus* and some *Meteorus* are often dependent on macrolepidopterans, mainly the families Geometridae, Noctuidae and Lasiocampidae.

Several groups of Bembridge Marls braconids belong to subfamilies today displaying larval or egg-larval parasitism of beetles, the supposed groundplan host group of Braconidae (Telenga 1952). These are mostly the plesiomorphic subfamilies Rhyssalinae, Doryctinae, Helconinae and Brachistinae, with the exception of Braconinae, which includes quite specialised genera (*Bracon*, *Cyanopterus* and especially *Bembracon*). Larval parasitism is known for the rhyssaline *Rhyssalus* and possibly *Dolopsidea*, the doryctine *Ontsira* (one from the less derived genus in subfamily Doryctinae) and braconine genera. Some of the recorded genera of Helconinae (*Taphaeus* and *Diospilus*) and Brachistinae (*Eubazus*) are known to include egg-larval parasitoids of beetles (Anobiidae, Cerambycidae, Nitidulidae, Curculionidae etc.). Worth mentioning is that the most primitive species of the euphorine genus *Meteorus* (for example, *M. corax* species group) are also parasitoids of beetle larvae, and it is possible that the Bembridge Marls species of this genus might be a coleopteran and not lepidopteran entomophage.

The fossil record of the subfamilies Alysiinae and Opiinae, parasitoids of flies, is doubtful because of the insufficient preservation of the respective fossils (see discussion under Opiinae above). In that connection, of importance is the first reliable record of Opiinae (*Biosteres peritus* (Cockerell)) in the Bembridge Marls. At the same time, it is surprising that not a single specimen of Aphidiidae (or Aphidiinae in Braconidae: Achterberg 1993), the obligate parasitoids of aphids, is found here (as well as in Florissant) in contrast to Oligocene and Miocene fossil sites (Stary 1973). Noteworthy is the absence of Euphorinae, specialised (often nymphal or imaginal) as parasitoids of various insects; these genera are described from Baltic amber and Florissant.

The majority of genera recorded in the Bembridge Marls (*Bassus*, *Eubazus*, *Ascogaster*, *Diospilus*, *Taphaeus*, *Homolobus*, *Biosteres*, *Microtypus* etc.) are very common and usually abundant today in the northern hemisphere temperate zone, with

species richness undoubtedly increasing in its southern parts. Conversely, the morphologically peculiar taxa such as *Bembracecon* gen. nov. with a number of original metasomal apomorphies, *Semionis* (of South African distribution now), species of *Bracon* and *Cyanopterus* with the forewing venation unusual for boreal taxa, *Ontsira distracta* (Cockerell) and *Chremylus infuscatus* sp. nov. with deviant wing venation and body structures, are more characteristic of at least a subtropical fauna. Thus the climatic inferences from Braconidae are somewhat contradictory, but indicate a warmer and possibly less humid past climate compared with today.

Composition of the braconid taxa in the Bembridge Marls and their host preferences implies the presence of large open and possibly rather wet grassland, or possibly wetlands, as the source environment. A number of specialised beetle parasitoids indicates the presence of at least isolated trees over these open lands. Alternatively, broadleaf forests might exist nearby, allowing the braconid parasitoids of beetles to enter the grasslands actively (for additional feeding) or passively (by wind), to reach a target water body for burial.

The most diverse Cenozoic braconid faunas are described from the Baltic amber (Brues 1923, 1933, 1939; Tobias, 1987) and the Florissant beds (Cockerell 1913; Brues 1906, 1910). The Baltic fauna is the best studied in respect to the hymenopterans. Despite taxonomic problems with several taxa, 18 subfamilies are recorded, including the endemic Diospilinae Tobias, 1987 (*Acampsohelconinae* Tobias, 1987 is now recorded as living: Achterberg 2002), embracing 52 genera and 101 species. All species and 19 genera were described from this amber, and two genera (*Bracon* and *Chelonus*) are reported, based on unidentified species. Compared with that of the Baltic amber, the Bembridge Marls braconid fauna is less rich, displaying only 13 subfamilies (lacking Diospilinae, *Acampsohelconinae*, Ichneutinae, Macrocentrinae, Miracinae and Aphidiidae, but with the addition of Homolobinae and Opiinae). Out of 23 Bembridge genera, only 11 are the same as in the Baltic amber and four endemic taxa, and only 39 species were identified. Of interest is that the subfamily Bracoinae is more diverse in the Bembridge Marls yielding three genera and six species, vs. one genus with no identified species in the Baltic amber. In contrast, the diversity of Euphorinae (including Blacini) is much restricted here (one genus and two species vs. ten genera and 17 species in the Baltic assemblage); the same holds true for Doryctinae (one genus, three species vs. four genera and about ten species) and Orgilinae (one genus, one species vs. two genera and 13 species, some of which are of obscure taxonomic position).

The fauna of the Florissant beds is much less studied, and includes only ten subfamilies, 17 genera (with one endemic, *Oligoneuroides* Brues, 1910) and 24 species. Investigated almost a century ago, its taxonomic composition is problematic now in respect of several taxa (see discussions under subfamilies). Nevertheless, it is possible to conclude that the Florissant fauna includes the subfamily Alysiinae, which is absent from the Bembridge Marls. Several genera recorded from the Florissant (*Agathis*, *Cremonops*, *Iphiaulax*, *Chelonus*, *Discoletes* etc.) are unknown in the Bembridge Marls, but their identity is not certain because of imperfection of either preservation or description. A reliable comparison of the Bembridge Marls and Florissant braconid assemblages requires a taxonomic revision of the latter.

### 3.10. Aculeata Scopoli, 1763

[By Alexander V. Antropov]

Eleven specimens of aculeate wasps and bees were studied. Of them, three represent the type material of *Polybia* ? *anglica*

Cockerell, 1921, *Polybia* ? *oblita* Cockerell, 1921 and *Sceliphron* (?) *brevior* Cockerell, 1921, and six more fossils are described herein for the first time. Two other fossils are attributed only to the family level, because of their poor preservation and lack of data necessary for more precise determination. The twelfth fossil, named *Philoponites clarus* Cockerell, 1915 is redescribed below based on a photograph of the holotype. The thirteenth fossil, *Mesitius*? *rectinervis* Cockerell, 1921, was not re-studied and is included herein for the sake of completeness.

Morphological terminology of the wing venation in general follows a traditional approach, with several additions concerning separate segments of longitudinal veins M and Cu.

#### 3.10.1. Superfamily Chrysidoidea Latreille, 1802

Family Bethyridae Haliday, 1839

Genus *Mesitius* Spinola, 1853

*Mesitius*? *rectinervis* Cockerell, 1921.

Plate 15, fig. 1

1921a *Mesitius*? *rectinervis* Cockerell, 1921, p. 21, fig. 25.

**Holotype.** NHMUK I.9352, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection.

**Remarks.** The type was not re-studied and its taxonomic position is not confirmed.

#### 3.10.2. Superfamily Vespoidea Laicharting, 1781

Family Vespidae Laicharting, 1781

**Remarks.** Four fossils considered below are attributed to Vespidae because of their very long and narrow forewing basal cell (m), discal cell I (1mcu) much longer than subdiscal cell (2cu<sub>a</sub>) in three samples, deeply emarginate inner eye orbits and the pronotum exceeding the tegula posteriorly in the last one.

##### 3.10.2.1. Subfamily Polistinae Lepeletier, 1836

Genus *Palaepolybia* Antropov, gen. nov.

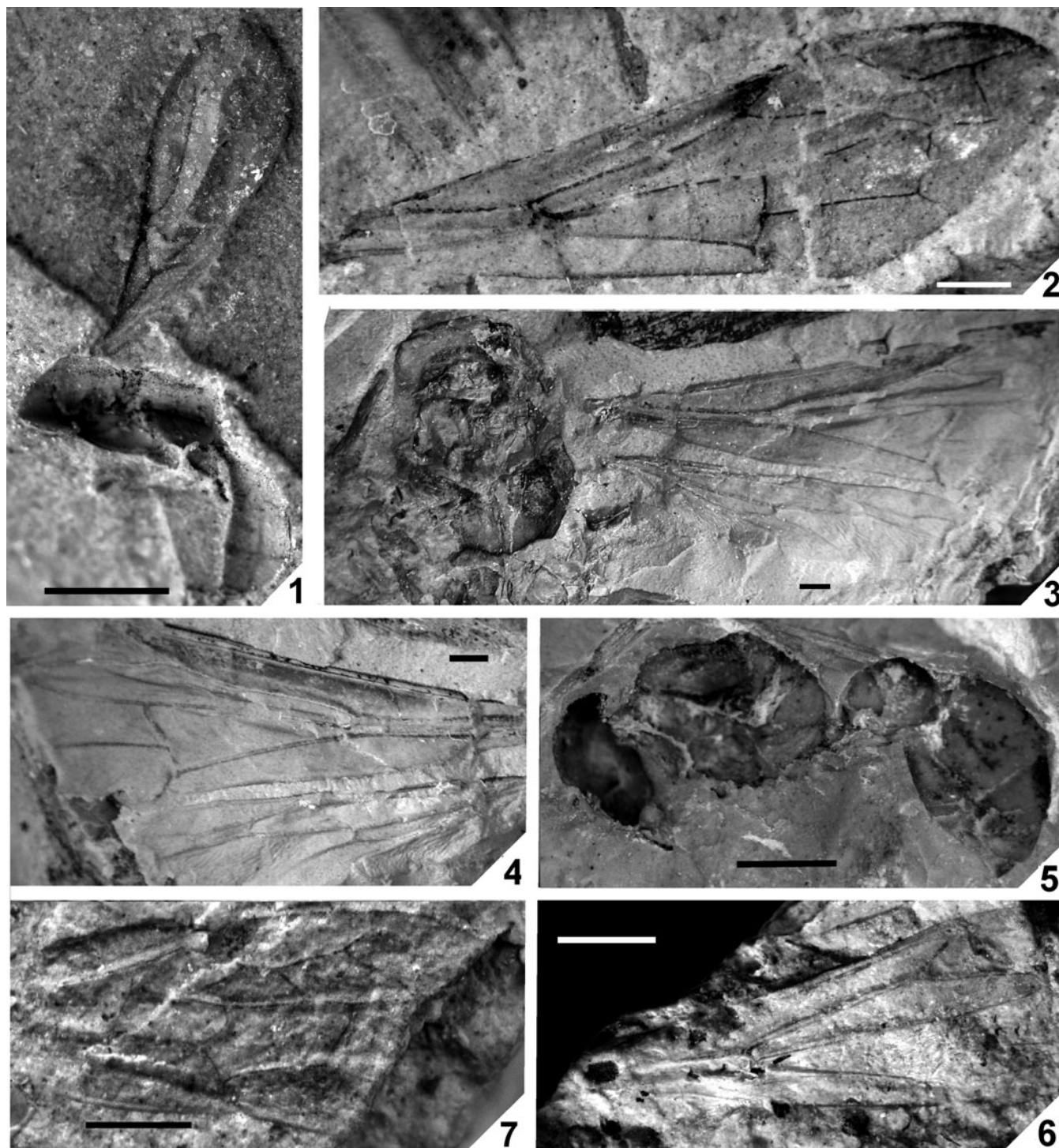
**Etymology.** After *palaesus*, the Greek for old, and genus *Polybia*. Gender feminine.

**Type species.** *Polybia*? *anglica* Cockerell, 1921; by present designation and monotypy.

**Diagnosis** (based on the forewing venation). Forewing with prStg less than half as long as ptStg and only slightly shorter than 1RS which is 0.21 times as long as M and 0.37 times as long as RS+M, 2RS S-like bent, 3RS much shorter than 4RS, 4RS somewhat longer than 3M, 2r-m practically straight, recurrent veins 1m-cu and 2m-cu joining 2rm near its posterior corners, 2rm wider than long and than 2RS, 2M1 and 2M3 almost equal; cu-a strongly postfurcal, half as long as 2M+Cu, 1Cu1 and 1Cu2 joined at right angle.

**Remarks.** The type and only species was described as a questionable *Polybia* by Cockerell (1921a) who stated that the specimen "agrees in general with *Polybia*". He also compared it with the genus *Apoica*. This viewpoint was discussed by Carpenter & Grimaldi (1997), who concluded that illustrations of the forewing venation of "*Polybia*" *anglica* showed "no particular resemblance to either genus – or to any polistine".

This new study of the part and counterpart holotype shows that its forewing marginal cell (r) touches apically the anterior margin of the wing (more or less separated from the wing margin in Masarinae and Eumeninae). There is a comparatively short pterostigma (ptStg) (comparatively long in Eumeninae



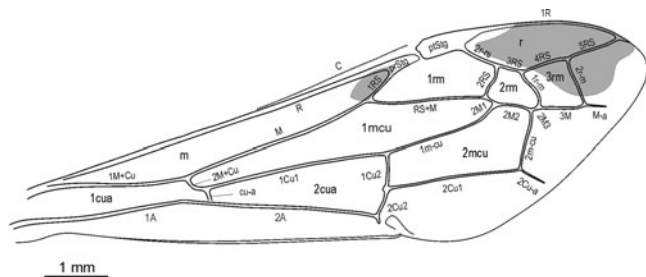
**Plate 15** (1) *Mesitius? rectinervis* Cockerell, holotype, NHMUK I.9352. (2) *Palaeopolybia anglica* (Cockerell), holotype, forewing, NHMUK In.24339. (3–4) *Protopolistes oblitus* (Cockerell), holotype, NHMUK In.20530/In.17166: (3) head, thorax and wings, part; (4) wings, counterpart. (5) *Sphaerancistrocerus petiolatus* sp. nov., holotype, body, NHMUK In.8967. (6) Subfamily Eumeninae Leach, *incertae sedis*, forewing, NHMUK In.43416. (7) *Philoponites clarus* Cockerell, holotype, forewing, USNM No 61409. Scale bars = 1 mm.

and Stenogastrinae), both recurrent veins join 2rm (2m–cu joining 3rm in Priorivespinae and some Euparagiinae). There is a right angle between veins 2M1 and 2RS (very obtuse in Vespinae), strongly postfurcal vein cu–a (antefurcal in Masarinae; interstitial or slightly postfurcal in Euparagiinae, Priorivespinae and Stenogastrinae; slightly postfurcal in Vespinae), and also short vein cu–a and unmodified subdiscal cell (2cu–a) (cu–a long and 2mcu narrowed and projecting apically in Euparagiinae). These features place this species

in the subfamily Polistinae (Carpenter & Rasnitsyn 1990; Brothers & Finnamore, 1993).

**Diagnosis.** In comparison with fossil genera, the forewing radial area of *Palaeopolybia* is most similar to that of *Palaeovespa* Cockerell, 1906 (Vespinae) in the relatively long prestigma, both recurrent veins joining the 2rm near its posterior corners, and 2rm wider than long.

However, *Palaeopolybia* differs from *Palaeovespa* in the following features (mainly in comparison with *P. socialis* Poinar,



**Text-figure 53** *Palaeopolybia anglica* (Cockerell), line drawing of holotype forewing, NHMUK In.24339.

2005, in which forewing venation has been described in detail): prStg somewhat shorter than ptStg (prStg almost equal to ptStg in *Palaeovespa*); 1RS 0.21 times as long as M (0.38 times in *Palaeovespa*); 1RS 0.37 times as long as RS+M (1.37 times in *Palaeovespa*); 2RS S-shaped (straight in *Palaeovespa*); 3RS much shorter than 4RS (almost equal to 4RS in *Palaeovespa*); 4RS somewhat longer than vein 3M (much shorter than 3M in *Palaeovespa*); 2r-m practically straight (strongly S-shaped in *Palaeovespa*); 2M1 almost equal to 2M3 (shorter than 2M3 in *Palaeovespa*); cu-a strongly postfurcal (slightly postfurcal in *Palaeovespa*); cu-a half as long as 2M+Cu (almost equal to 2M+Cu in *Palaeovespa*); angle between 1Cu1 and 1Cu2 right (acute in *Palaeovespa*); angle between 2Cu1 and 2m-cu obtuse (right in *Palaeovespa*).

In comparison with extant genera, no similarity in the basic features is found between *Palaeopolybia* and *Polybia* Lepeletier, 1836. The two genera differ in the following features: r roundly acute apically (strongly acute in *Polybia*); prStg shorter than ptStg and almost as long as 1RS (1RS at least 2–3 times as long as ptStg, which is still longer comparing prStg in *Polybia*); 2r-rs equal to 2RS (1.2–1.5 times as long as 2RS in *Polybia*); 1RS c. 0.2 times as long as M (0.3–0.37 times as long as 1M in *Polybia*); 2RS S-shaped (2RS straight or slightly curved outward in *Polybia*); 2RS and 2M1 join at right angle (at obtuse angle in *Polybia*); 3RS approximately half as long as 4RS (3–4 times shorter than 4RS in *Polybia*); 4RS almost equal to 5RS (1.5–2 times shorter than 5RS in *Polybia*); 4RS slightly longer than 3M (distinctly shorter than 3M in *Polybia*); 5RS approximately equal to 2r-m (c. 1.5 times as long as 2r-m in *Polybia*); 2rm wider than long (distinctly longer than wide in *Polybia*); 2M2 almost three times as long as 2M1 and 2M3 (subequal to 2M1 and 2M3 in *Polybia*); 3rm longer than wide (distinctly wider than long in *Polybia*); cu-a approximately half as long as 2M+Cu (0.12–0.14 times as long as 2M+Cu in *Polybia*); 1Cu2 not curved (S-shaped in *Polybia*); 1Cu2 approximately equal to 2Cu2 (distinctly longer than 2Cu2 in *Polybia*).

Instead, *Palaeopolybia* shows some resemblance to *Apoica* Lepeletier, 1836, for example, in that: prStg is longer than wide, 2rm not longer than wide, 2rm and 3rm of comparable width; 5RS as long as 2r-m; 1Cu2 straight. At the same time, the two genera are different in a number of important features: marginal cell roundly acute apically (distinctly acute in *Apoica*); ptStg distinctly wider (very narrow in *Apoica*); prStg only slightly shorter than 1RS (less than two thirds as long as 1RS in *Apoica*); 2r-rs equal to 2RS (twice as long as 2RS in *Apoica*); 2r-rs hardly longer than 3RS (almost 2.5 times as long as 3RS in *Apoica*); 2r-rs shorter than 5RS (2r-rs more than 1.5 times as long as 5RS in *Apoica*); 3RS c. 0.5 times as long as 4RS (only slightly shorter than 4RS in *Apoica*); 3RS 2 times longer than 2M3 (3RS shorter than 2M3 in *Apoica*); 4RS approximately equal to 5RS (distinctly shorter than 5RS in *Apoica*); 4RS slightly longer than 3M (distinctly shorter than 3M in

*Apoica*); 1r-m S-shaped (straight in *Apoica*); 2M2 almost three times as long as 2M1 and 2M3 (0.67 times as long as 2M1 and 0.4 times as long as 2M3 in *Apoica*); M only slightly longer than vein 1Cu1 (almost 1.5 times as long as 1Cu1 in *Apoica*); RS+M only slightly shorter than vein 1m-cu (RS+M c. 0.7 times as long as 1m-cu in *Apoica*); 2m-cu straight (curved outward in *Apoica*); 2m-cu c. half as long as 2Cu1 (almost one third as long as 2Cu1 in *Apoica*); 1m-cu c. 6 times as long as its maximum width (almost eight times as long as its maximum width in *Apoica*); cu-a c. half as long as 2M+Cu (one sixth to one seventh as long as 2M+Cu in *Apoica*); 1cu-a not shorter than subdiscal cell (distinctly shorter than subdiscal cell in *Apoica*).

The above comparison justifies generic level distinction of the fossil under consideration.

The combination of other venational features of *Palaeopolybia* is otherwise unknown in Polistinae, in which prestigma (prStg) is several times shorter than ptStg (slightly shorter only in *Apoica* Lepeletier, 1836), 2rm is not wider than long nor wider than 3rm, both recurrent veins (1m-cu and 2m-cu) never joining 2rm near its posterior corners, and vein cu-a is much shorter than vein 2M+Cu, often almost nonexistent.

*Palaeopolybia anglica* (Cockerell, 1921), new combination  
Plate 15, fig. 2; Text-fig. 53

1921a *Polybia* ? *anglica* Cockerell, p. 20, fig. 23.

1997 *Polybia* ? *anglica* Cockerell: Carpenter & Grimaldi, p. 2.

**Holotype.** NHMUK In. 24339; Bembridge Marls, NW Isle of Wight, UK, Hooley Collection. A piece of rock 43.8 × 28.6 mm with almost complete impression of the forewing lacking its base, apex of prestigma and base of pterostigma near its widest side. Convex veins and concave membrane of the discal cell I indicate that the fossil is the obverse impression (the part as opposed to the counterpart) of the right forewing.

**Description** (based on the forewing venation). Forewing with prStg less than half as long as ptStg and only slightly shorter than 1RS, which is 0.21 times as long as M and 0.37 times as long as RS+M; 2RS S-like bent; 3RS much shorter than 4RS; 4RS somewhat longer than 3M; 2r-m practically straight; recurrent veins 1m-cu and 2m-cu joining 2rm near its posterior corners; 2rm wider and longer than 2RS; 2M1 and 2M3 almost equal; cu-a strongly postfurcal, half as long as 2M+Cu; 1Cu1 and 1Cu2 joined at right angle. Two darkened spots are present: the larger one covering almost whole marginal cell and also anteriodistal half of 3rm and anterior half of the free cell corresponding to 4rm, and the smaller spot in the distal corner of basal cell.

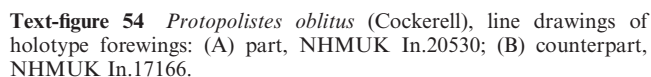
Vein measurements in mm: R1 1.63; 2r-rs 0.53; 1RS 0.62; 2RS 0.48; 3RS 0.38; 4RS 0.72; 5RS 0.78; 2M+Cu 0.46; M 2.98; RS+M 1.75; 2M1 0.18; 2M2 0.55; 2M3 0.15; 3M 0.68; M-a 0.60; 1Cu1 2.65; 1Cu2 0.50; 2Cu2 0.44; 1m-cu 1.78; 2m-cu 0.89; 1r-m 0.58; 2r-m 0.73; cu-a 0.22; 2Cu1 2.03; 2Cu-a 0.50; 2A 2.38. Total forewing length c. 9.6 mm.

Genus *Protopolistes* Antropov, gen. nov.

**Etymology.** After *protos*, the Greek for previous, and genus *Polistes*. Gender masculine.

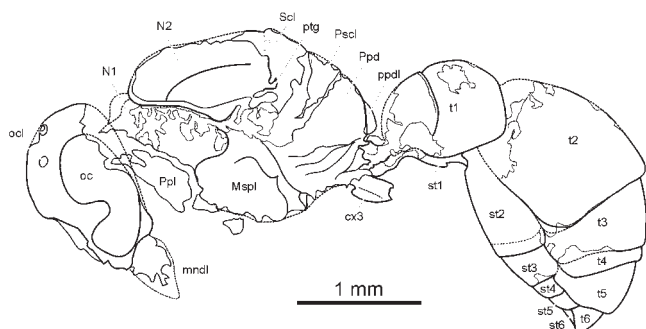
**Type species.** *Polybia oblita* Cockerell, 1921, by present designation and monotypy.

**Diagnosis** (based on wing venation). Forewing with prStg only slightly shorter than 1RS, M c. 3 times as long as 1RS and practically equal to 1Cu1; 1m-cu at least twice as long as



In comparison with extant genera, among recent Vespidae the weakly postfurcal position of cu-a and its similar length subequal to  $2M+Cu$  are characteristic of only a limited number of taxa, including *Vespa* and *Provespa* Ashmead, 1903 (Vespi-

**Type species.** *Sphaerancistrocerus petiolatus* sp. nov.; by present designation and monotypy.



**Text-figure 55** *Sphaerancistrocerus petiolatus* sp. nov., line drawing of holotype body, NHMUK In.8967. Body parts: cx3 = metacoxa; mndl = mandible; Msp = mesopleuron; N1 = pronotum; N2 = mesonotum; oc = compound eye; ocl = ocelli; Ppd = propodeum; ppdl = propodeal lamella; Psc = postscutellum; ptg = parategula; Scl = scutellum; st1–6 = metasomal sternites I–VI (abdominal sterna II–VII); t1–6 = metasomal tergites I–VI (abdominal terga II–VII).

**Diagnosis.** Several eumenine genera with a transverse carina on metasomal tergite I, including fossils *Ancistrocerus berlandi* Piton, 1840, *Ancistrocerus eocenicus* Piton, 1840 and *Symmorphus senex* Carpenter, 2000 are known. They differ from *Sphaerancistrocerus* gen. nov. in having a distinct bend dividing anterior and posterior parts of tergite I. The only known eumenine genus with a uniformly rounded first metasomal segment is the fossil *Rotundipetiolus longiantennus* Zhang, 1989, which distinctly differs from *Sphaerancistrocerus* gen. nov. in its much larger body size and the absence of a transverse carina on tergite I.

*Sphaerancistrocerus petiolatus* Antropov, sp. nov.

Plate 15, fig. 5; Text-fig. 55

**Etymology.** After *petiolus*, the Latin for petiole.

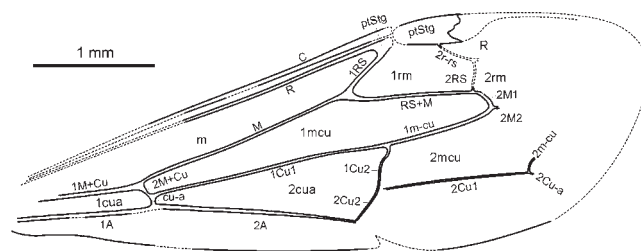
**Holotype.** NHMUK I.8967, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection. Female. The part is a piece of rock 41.5 × 37.0 mm with impression of the partially preserved head, thorax, propodeum, hind coxa, and metasoma. Antennae, legs and wings not preserved.

**Description** (based on body structure). Head with front significantly convex, noticeably stronger than occiput; vertex noticeably convex; ocelli in at least right-angled triangle; pronotal collar moderately roundly convex, without transverse carina; scutum convex anteriorly, with rest surface flattened; adlateral lines almost reaching scutal posterior margin; scutellum posteriorly and postscutellum flattened; metacoxa with outer longitudinal carina; propodeum uniformly rounded, without anterior teeth, with oval apical lamellae; 1st metasomal segment with short petiole not longer than metacoxa; dorsal surface of metasomal tergite I uniformly convex, with transverse carina but without distinct bend between anterior and posterior parts; metasomal sternite I with broad transverse apical depression; metasomal tergite II 1.25 as long as metasomal tergite I, uniformly convex, without isolated border.

Length of body parts in mm: head 0.83; scutum 1.18; scutellum 0.38; postscutellum 0.29; mesosoma 2.06; petiolus 0.15; metasomal tergite I 1.11; anterior part of metasomal tergite I 0.49; posterior part of metasomal tergite I 0.54; metasomal tergite II 1.21; metasomal tergites II–VI 2.07. Total body length c.6.0 mm.

### 3.10.2.3. Subfamily Eumeninae Leach, 1815, incertae sedis

Plate 15, fig. 6; Text-fig. 56



**Text-figure 56** Subfamily Eumeninae Leach, *incertae sedis*, line drawing of forewing, NHMUK In.43416.

**Material studied.** NHMUK In.43416, Bembridge Marls, NW Isle of Wight, UK, Hooley Collection. Sex unknown. The piece of rock 34.8 × 26.7 mm with an impression of the partially preserved forewing. Distal part of the wing including marginal and submarginal cells, and discal cell II not preserved.

**Description.** Vein measurements in mm: prStg 0.17, ptStg 0.79, 2M+Cu 0.13; M 1.86; 1RS 0.50; RS+M 1.05; 2M1 0.23; cu-a 0.08; 1Cu1 1.78; 1Cu2 0.36; 2Cu2 0.33; 1m-cu 1.15; 2Cu1 1.27; 2A 1.68. Total forewing length ? 5.3 mm.

**Remarks.** Though the form of basal, discal I, and subdiscal cells and enlarged pterostigma attribute the specimen to the subfamily Eumeninae, absence of the most important cells of the submarginal sector prevents its placement even to genus.

### 3.10.3. Family Tiphidae Leach, 1815

#### 3.10.3.1. Subfamily Tiphinae Leach, 1815

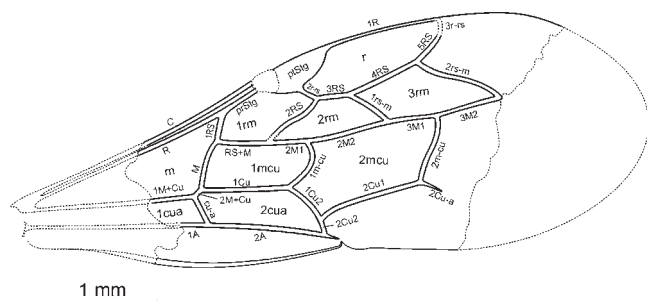
Genus *Philoponites* Cockerell, 1915

**Type species.** *Philoponites clarus* Cockerell, 1915; by monotypy.

**Diagnosis** (based on forewing venation). Forewing with complete marginal (r), three submarginal (1rm, 2rm, and 3rm), two discal (1m-cu and 2m-cu), basal (m), sub-basal (1cua), and subdiscal (2cua) cells. Marginal cell not strongly elongate, probably rounded or truncate apically; 2rm equal to 1rm; 3rm more than 1.3 times as long as 2rm; 2m-cu 1.5 times as long as 1m-cu; 2cua longer than 1m-cu; ptStg enlarged, oval, somewhat longer than prStg; 2r-m at least twice as long as 5RS; M almost straight; 1m-cu strongly curved outwards, received by 2rm; 2m-cu almost straight, received by 3rm; cu-a antefurcal, oblique, longer than 2M+Cu; 2Cu-a short, not reaching wing margin.

**Remarks.** *Philoponites* was described by Cockerell (1915) as a member of “Philanthidae” and compared venationally with *Philoponus* Kohl, 1889 (junior synonym of *Pseudoscolia* Radoszkowski, 1876). I had no opportunity to study the holotype kept at the USNM (No 61409); however, I did study a photograph, which unfortunately does not display all necessary details. For example, not clearly visible are the apex of the marginal cell and posterior part of 2RS near RS+M. The form and the relative sizes of ptStg, 1m-cu, 2m-cu, and 1rm of the type species do not correspond to those of Philanthidae *sensu* Cockerell (1915), nor Philanthinae *sensu* Bohart & Menke (1976). Particularly, 1m-cu strongly curved outwards does not occur in Crabronidae or generally in digger wasps. On the contrary, the forewing venation of *Philoponites clarus* is similar to that of some tiphid wasps of the subfamily Tiphinae. Furthermore, if the posterior part of 2RS of *Philoponites clarus* was not reaching RS+M, its venation becomes practically identical to that of *Paratiphia praefecta* Cockerell, 1907 (Upper Eocene shales of Florissant).

However, both of these species essentially differ from extant representatives of the genus *Paratiphia* Saussure & Sichel,



**Text-figure 57** *Philoponites clarus* Cockerell, line drawing of holotype forewing, USNM No 61409.

1864 in the form of the marginal cell, with its simple narrowed apex touching the wing margin; whereas *Paratiphia* species have the marginal cell either broadly rounded apically and not reaching the wing margin, or entirely opened anteriorly. In any case, the marginal cell of *Paratiphia* has a short apical vein Rs-a, which is not present in either *Paratiphia praefecta* or *Philoponites clarus*. As a result, I consider, that *P. praefecta* Cockerell, 1907 should be transferred from the genus *Paratiphia* to *Philoponites*, thus becoming *Philoponites praefectus* (Cockerell), comb. nov., and the genus *Philoponites* should be included in the family Tiphidae.

*Philoponites clarus* Cockerell, 1915  
Plate 15, fig. 7; Text-fig. 57

1915 *Philoponites clarus* Cockerell, pp 482, 499, plate 64, figure 2.

1976 *Philoponites clarus* Cockerell: Bohart & Menke, p. 31.

1992 *Philoponites clarus* Cockerell: Carpenter, p. 496.

**Holotype.** USNM No 61409; sex unknown. Lacoe Coll. 7620. Incomplete impression of the right forewing missing distal part of outer membrane and proximal parts of wing and of pterostigma.

**Description** (based on forewing venation). Forewing with complete marginal (r), three submarginal (1rm, 2rm, and 3rm), two discal (1mcu and 2mcu), basal (m), sub-basal (1cua), and subdiscal (2cua) cells. Marginal cell not strongly elongate, probably rounded or truncate apically; 2rm equal to 1rm; 3rm more than 1.3 times as long as 2rm; 2mcu 1.5 times as long as 1mcu; 2cua longer than 1mcu; ptStg enlarged, oval, somewhat longer than prStg; 2r-m at least twice as long as 5RS; M almost straight; 1m-cu strongly curved outwards, received by 2rm; 2m-cu almost straight, received by 3rm; cu-a antefurcal, oblique, longer than 2M+Cu; 2Cu-a short, not reaching wing margin.

Vein measurements in mm: ptStg 0.66; 1R 0.98; 3r-rs 0.15; 2r-rs 0.18; 1RS 0.28; 2RS 0.60; 3RS 0.28; 4RS 0.68; 5RS 0.30; 1r-m 0.43; 2r-m 0.80; RS+M 0.44; M 0.45; 2M1 0.35; 2M2 0.75; 3M1 0.44; 3M2 0.58; 2M+Cu 0.08; cu-a 0.28; 1Cu1 0.76; 2Cu1 0.78; 1Cu2 0.41; 2Cu2 0.14; 1m-cu 0.45; 2m-cu 0.60; 2Cu-a 0.18; 2A 1.00. Total forewing length c. 5.8 mm.

### 3.10.3.2. Subfamily Dryophiinae Antropov, nov.

**Type genus.** *Dryophia* Antropov, gen. nov.; by present designation and monotypy.

**Diagnosis.** Subfamily Dryophiinae nov. somewhat resembles females of Dryinidae (Chrysidoidea), differing from them in having frons flattened and head strongly narrowed and elongate behind the eyes, and by the non-specialised mesosoma and more complete forewing venation.

Subfamily Dryophiinae is more similar to Tiphidae (Vespoidea) in having the pronotum deeply U-like excised and reaching the tegulae posteriorly, and in the structure of the mesosoma, contiguous metacoxae, and sessile metasoma.

Among Tiphidae, the new subfamily is most similar to Myzininae in the presence of a frontal tubercle, comparatively large eyes and structure of the scutellum, postscutellum and propodeum, but differs in its head elongate and narrowed behind the eyes, narrow pronotal collar, deeply emarginate pronotum, tegulae ending posteriorly at transscutal furrow, absence of scutal adlateral lines, hind femora not widened, and in absence of metasomal intersegmental constrictions and a pygidial plate.

Genus *Dryophia* Antropov, gen. nov.

**Etymology.** After generic names *Dryinus* and *Tiphia*. Gender feminine.

**Type species.** *Dryophia oculata* Antropov, sp. nov.; by present designation and monotypy.

**Diagnosis.** As for subfamily.

*Dryophia oculata* Antropov, sp. nov.  
Plate 16, fig. 1; Text-fig. 58

**Etymology.** After *oculatus*, the Latin for goggle-eyed.

**Holotype.** NHMUK In.17086, Bembridge Marls, NW Isle of Wight, UK, Smith Collection. Female. The holotype is a piece of rock 40.0 × 39.7 mm with impressions of the head, mesosoma, metasoma and partially preserved left femur, tibia and left forewing. Antennae, other legs, right wings and left hind wings not preserved.

**Description.** Compound eyes very large and convex, head behind eyes strongly narrowed and elongate; frons flat, not broader than eye, with indistinct tubercle below; vertex weakly convex; ocelli small, round, in equilateral triangle between eyes; occipital carina distinct at least dorsally; pronotal collar short, weakly concave anteriorly, with strong lateral corners; pronotum deeply emarginate, U-shaped posterior and reaching tegulae posterolaterally; notauli almost reaching transscutal furrow; adlateral lines absent; scutellum large and flat, rounded posteriorly; tegulae semicircular, not enlarged, but ending at transscutal furrow posteriorly; forewings somewhat shortened, probably scarcely reaching metasomal middle; 1cua only scarcely narrower than 1rm; hind coxae not enlarged; hind femora and tibiae not widened; propodeum somewhat elongate, oval, without dorsal enclosure and lateral and posterior carinae or furrows; metasoma sessile, without elongate petiole, and constrictions between tergites; apical tergite without pygidial plate.

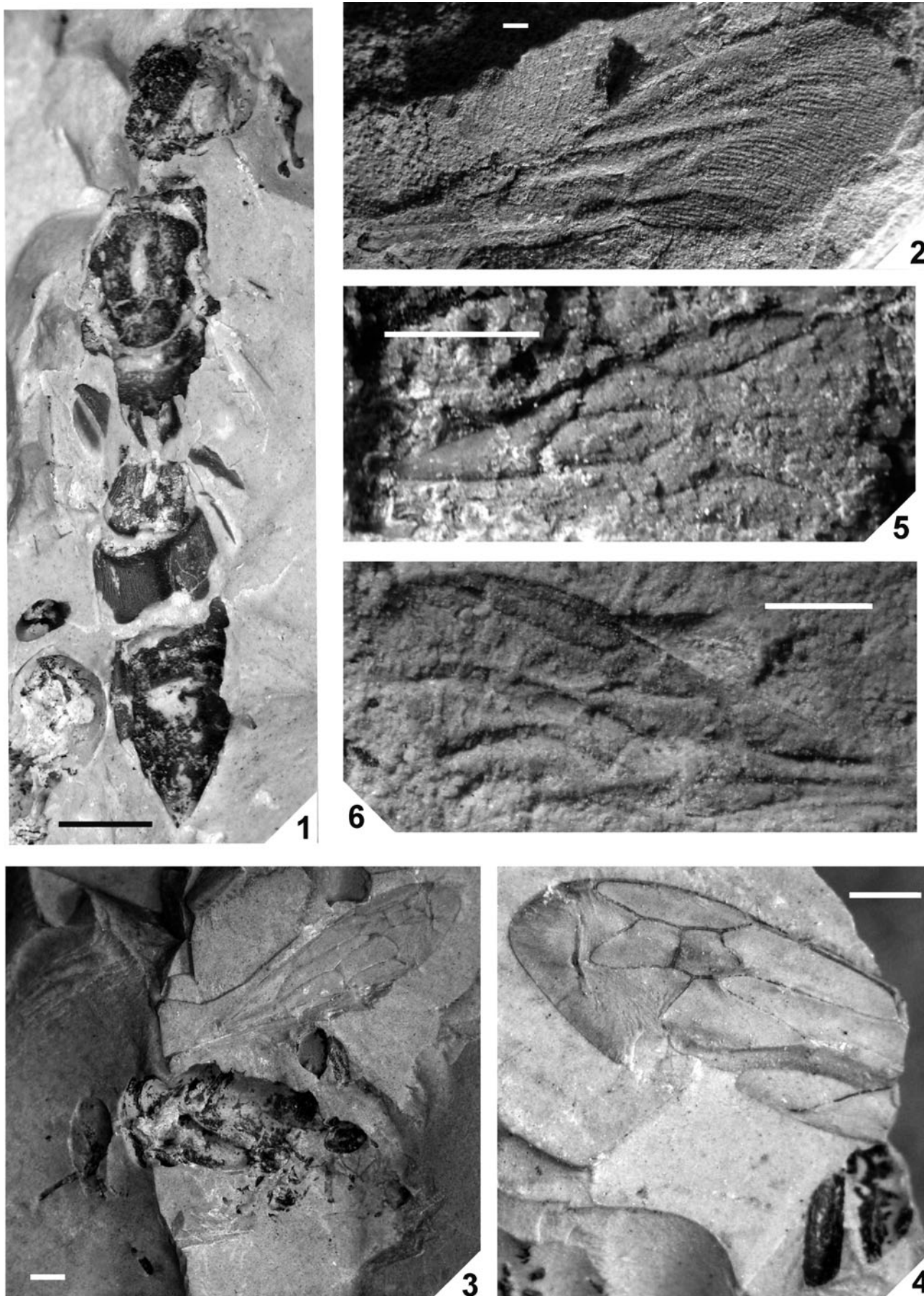
Body parts measurements in mm: head length 1.67; eye length 0.68; eye height 0.99; mesomal length 2.81; pronotal collar length 0.21; pronotal length 0.93; pronotal width 1.25; scutal length 0.93; scutellar length 0.56; postscutellar length 0.17; propodeal length 0.82; pronotal collar width 0.96; scutal width 1.13; scutellar width 0.85; metasomal width 1.53; metasomal length 3.99; 1st metasomal segment 0.90; 2nd metasomal segment 0.79. Total body length c. 8.54 mm. Forewing veins measurements in mm: 2M+Cu 0.04; cu-a 0.21; M 0.36; RS+M 0.29; 1Cu1 0.37; 1m-cu 0.22.

### 3.10.4. Family Scolidae Leach, 1815

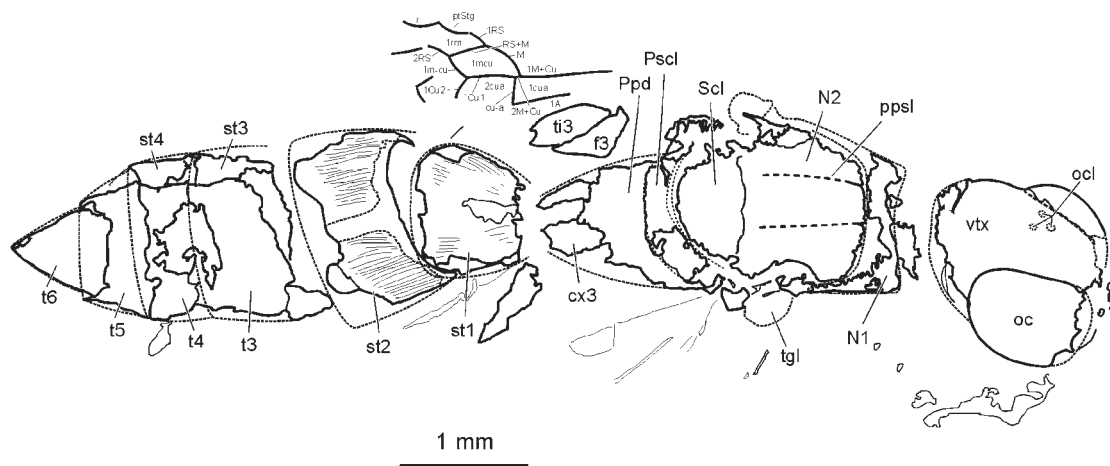
#### 3.10.4.1. Subfamily Palaeoscoliinae Antropov, nov.

**Type genus.** *Palaeoscolia* gen. nov., by present designation and monotypy.

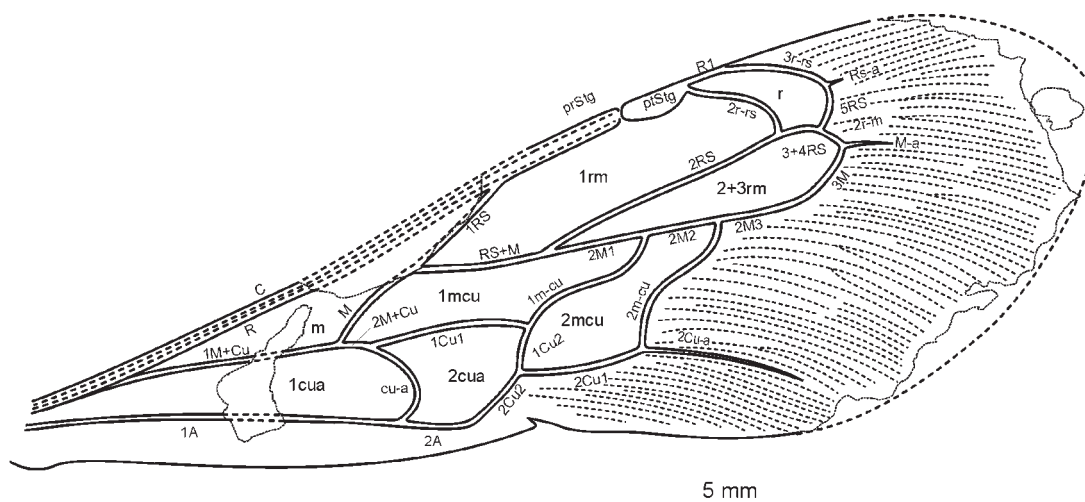
**Diagnosis.** Subfamily Palaeoscoliinae differs from Archaeoscoliinae Rasnitsyn, 1993 in its broad and corrugated outer



**Plate 16** (1) *Dryophia oculata* sp. nov., holotype, body, NHMUK In.17086. (2) *Palaeoscolia relictia* sp. nov., holotype, forewing, NHMUK In.24527. (3–4) *Protosceliphron brevior* (Cockerell), holotype, NHMUK In.17472/In.24340: (3) head, thorax, wings, and anterior part of metasoma, part; (4) apical parts of wings and hind femur and tibia, counterpart. (5) *Tythopsen nanus* sp. nov., holotype, forewing, NHMUK In.25568. (6) *Plisomena gigantea* sp. nov., holotype, forewing, NHMUK In.24789. Scale bars = 1 mm.



**Text-figure 58** *Dryophia oculata* sp. nov., line drawing of holotype body, NHMUK In.17086. Body parts: cx3 = metacoxa; f3 = hind femur; N1 = pronotum; N2 = mesonotum; oc = compound eye; ocl = ocelli; Ppd = propodeum; ppsl = parapsidal line; Psc = postscutellum; Scl = scutellum; st1–4 = metasomal sternites I–IV (abdominal sterna II–V); t3–6 = metasomal tergites III–VI (abdominal terga IV–VII); ti3 = hind tibia; tgl = tegula; vtx = vertex. Wing cells: 1cu-a = sub-basal; r = marginal; 1rm = submarginal I; 1m-cu = discal I; 2cu-a = subdiscal.



**Text-figure 59** *Palaeoscolia relictata* sp. nov., line drawing of holotype forewing, NHMUK In.24527.

forewing membrane, not elongate ptStg half as long as prStg, short marginal cell, two closed submarginal cells (2+3rm formed by merged 2rm and 3rm), and distinctly postfurcal and curved distal cu-a. Palaeoscoliinae also differs from Proscoliinae Rasnitsyn, 1977 in having ptStg not strongly enlarged (half as long as prStg), 2RS straight, bearing no inner appendicular vein, 2RS and 2r-m non-convergent anteriorly, 1m-cu shorter than 2+3rm and twice as long as 2cu-a, 1m-cu and 2m-cu S-shaped, and cu-a strongly postfurcal.

Subfamily Palaeoscoliinae is more similar to Scoliinae Latreille, 1802 and particularly to the genera *Campsoscolia* Betrem, 1933 and *Campsomeris* Guérin, 1838, but differs in having a wide and comparatively short costal prStg, a notch separating a distinct and not strongly elongate ptStg from prStg, and S-curved 1m-cu.

Genus *Palaeoscolia* Antropov, gen. nov.

**Etymology.** After *palaeus*, the Greek for old, and genus *Scolia*. Gender feminine.

**Type species.** *Palaeoscolia relictata* Antropov, sp. nov.; by present designation and monotypy.

**Diagnosis.** As for the subfamily.

*Palaeoscolia relictata* Antropov, sp. nov.

Plate 16, fig. 2; Text-fig. 59

**Holotype.** NHMUK In.24527; Bembridge Marls, NW Isle of Wight, UK, Hooley Collection. A piece of rock 44.8 × 28.6 mm with almost complete impression of the forewing lacking its base, middle part of anterior margin and distal margin of membrane. Preservation rather poor because of salt crystals covering rock surface (strong oblique illumination can somewhat improve visibility of the fossil).

**Description** (based on forewing venation). Forewing with marginal (rR), two submarginal (1rm and 2+3rm), two discal (1m-cu and 2m-cu), basal (m), sub-basal (1cu-a), and subdiscal cells (2cu-a); ptStg distinct, oval, half as long as prStg; costal notch between ptStg and prStg distinct; marginal cell strongly shortened, rounded apically; second and third submarginal cells united; 1rm strongly elongate, longer and broader than 2+3rm; 2RS straight, without inner appendix vein, not strongly convergent with 2rs-m anteriorly; 5RS 1.6 times as long as 2rs-m; M almost straight, 0.8 times as long as RS+M; 2M1 1.3 times as long as 2M2, 1.2 times as long as 2M3, 1.4 times as long as 3M; 1m-cu twice as long as 2cu-a; 1m-cu and 2m-cu strongly S-shaped, both received by 2+3rm; cu-a postfurcal,

roundly curved distally, 2.5 times as long as 2M+Cu; 2A 0.8 times as long as cu-a, 1.4 times as long as 1Cu2, and 0.9 times as long as 2Cu2; 2Cu-a long, but not reaching wing margin.

Vein measurements in mm: prStg 2.62; ptStg 1.27; 1R 0.78; 3r-rs 1.86; 2r-rs 1.90; 1RS 2.13; 2RS 4.75; 3+4RS 0.77; 5RS 0.80; 2r-m 0.50; RS+M 2.44; M 1.97; 2M1 1.84; 2M2 1.37; 2M3 1.53; 3M 1.26; 2M+Cu 0.61; cu-a 1.46; 1Cu1 2.90; 2Cu1 2.25; 1Cu2 0.84; 2Cu2 1.40; 1m-cu 2.73; 2m-cu 2.74; 2A 1.20; Rs-a 0.31; M-a 0.90; 2Cu-a 3.01. Total forewing length *c.* 20.8 mm.

### 3.10.5. Superfamily Apoidea Latreille, 1802

#### 3.10.5.1. Family Sphecidae Latreille, 1802

Subfamily Sceliphrinae Ashmead, 1899

Tribe Protosceliphirini Antropov, nov.

**Type genus.** *Protosceliphron* gen. nov., by present designation and monotypy.

**Diagnosis.** Protosceliphirini tribe nov. resembles Sceliphirini in the forewing venation, but differs from all Sphecidae in the postfurcal hind wing cu-a and particularly in the metasomal segment I lacking a petiole.

The unique species of the type genus has been described as *Sceliphron* (?) *brevior* Cockerell, 1921 based mainly on the forewing venation. Forewing venation and structure of the thorax and propodeum (mesosoma), as preserved, do generally correspond to those of extant representatives of the genus *Sceliphron* Klug, 1801, except that 5RS is uniformly curved outwards in the new tribe and not S-shaped as usually in *Sceliphron*, and that 1Cu1 is more curved forwards than in *Sceliphron*. Hind wing shows a greater difference in the postfurcal position of cu-a, which is interstitial in Sceliphirini.

Most striking is, however, structure of the metasomal segment I of the new tribe, which lacks a long petiole of entirely sternal nature which is otherwise a universal feature of Sphecidae.

The visible metasomal segment I of all known members of Sceliphirini have a more or less elongate tubular petiole, formed by metasomal sternite I, while metasomal tergite I is divided into a larger convex posterior part, embracing laterally the posterior end of sternite I, and a broadly separated minute acrotergite I. The last forms, above the anterior end of the sternal petiole, a dorsal part of the metasomal joining plate connecting the metasoma with the propodeum. Some extant genera of Sphecinae (Stangeellini: *Stangeella* Menke, 1962) or related to Chloriontinae (*Chlorion* Latreille, 1809) have a comparatively short petiole, which is not longer than the hind coxa, but with the posterior part of metasomal tergite I separated from its acrotergite. *Chalybion mortuum* Cockerell, 1907 (Sceliphirinae, Sceliphirini) from the Upper Eocene shales of Florissant, Colorado has a short but distinct petiole. *Hoplisidia kohliana* Cockerell, 1906 (Sceliphirinae, Sceliphirini) also from Upper Eocene shales of Florissant, Colorado described in "Nyssonidae" and attributed to Tiphiidae (Evans 1966) and later to Sceliphirini (Menke & Rasnitsyn 1987) also have a short petiole, though its precise structure is not visible in the type specimen.

The holotype of the only known species in the new tribe has a sclerite lying directly behind the propodeal apex which, judging from its form and size, represents metasomal tergite I or at least the main fragment of its anterior part. It has a distinct rectangular prominence at the anterior margin which obviously represents its acrotergite forming a joining plate, but not separated from the posterior part of the tergite. Thus, the metasoma of *Protosceliphron* is sessile and has no elongate sternal petiole, as characteristic of the subfamily Sceliphirinae and the family Sphecidae in general.

Genus *Protosceliphron* Antropov, gen. nov.

**Etymology.** After *protos*, the Greek for previous, and genus *Sceliphron*. Gender neuter.

**Type species.** *Sceliphron* (?) *brevior* Cockerell, 1921, by present designation and monotypy.

**Diagnosis.** As for the tribe.

*Protosceliphron brevior* (Cockerell, 1921), new combination

Plate 16, figs 3, 4; Text-fig. 60

1921a *Sceliphron* (?) *brevior* Cockerell, pp 20–21, fig. 24.

1976 *Sceliphron* (?) *brevior* Cockerell: Bohart & Menke, p. 31, 106.

**Holotype.** NHMUK In.17472/In. 24340 (part and counterpart), Bembridge Marls, NW Isle of Wight, UK, Smith/Hooley Collections. The part is a piece of rock 57.6 × 41.4 mm with impression of the partially preserved head, thorax, metasomal base, fragments of antennae, hind femur and tibia, almost complete right forewing, and base of right hind wing. The counterpart is a piece of rock 24.4 × 18.7 mm with the partially preserved hind femur and tibia, and apical half of the right forewing.

**Description.** Head flattened, with vertex strongly broadened; mandibles long, almost straight, acute apically; palpal segments elongate; hind femora stout; forewing venation almost like that of *Sceliphron*, with complete basal, marginal, three submarginal, two discal, sub-basal, and subdiscal cells; forewing 1Cu1 distinctly curved forwards; hind wing cu-a postfurcal; metasomal tergite I complete, with acrotergite not separated from posterior part; sternal petiole not developed.

Vein measurements in mm: Forewing: prStg 1.78; ptStg 0.94; R1 1.34; 1RS 0.40; 2RS 0.70; 3RS 0.55; 4RS 0.75; 5RS 0.68; RS+M 2.61; M 1.18; 2M1 0.40; 2M2 0.41; 2M3 0.36; 3M 1.33; 2M+Cu 0.16, 1Cu1 2.30; 2Cu1 1.90, 1Cu2 0.66; 2Cu2 0.18, 2A 2.53; 2r-rs 0.48, 3r-rs 0.29; 1r-m 0.60, 2r-m 1.00; 1m-cu 1.65; 2m-cu 0.93; cu-a 0.25; 2Cu-a 0.47. Hindwing: Cu 0.21; cu-a 0.28. Total forewing length *c.* 10.1 mm, hindwing 6.8 mm.

**Remark.** Judging from the length of the left basal and right apical antennomeres and the relative length of antennae, the studied specimen is a female.

#### 3.10.5.2. Family Crabronidae Latreille, 1802

Subfamily Pemphredoninae Dahlbom, 1835

Tribe Psenini A. Costa, 1858

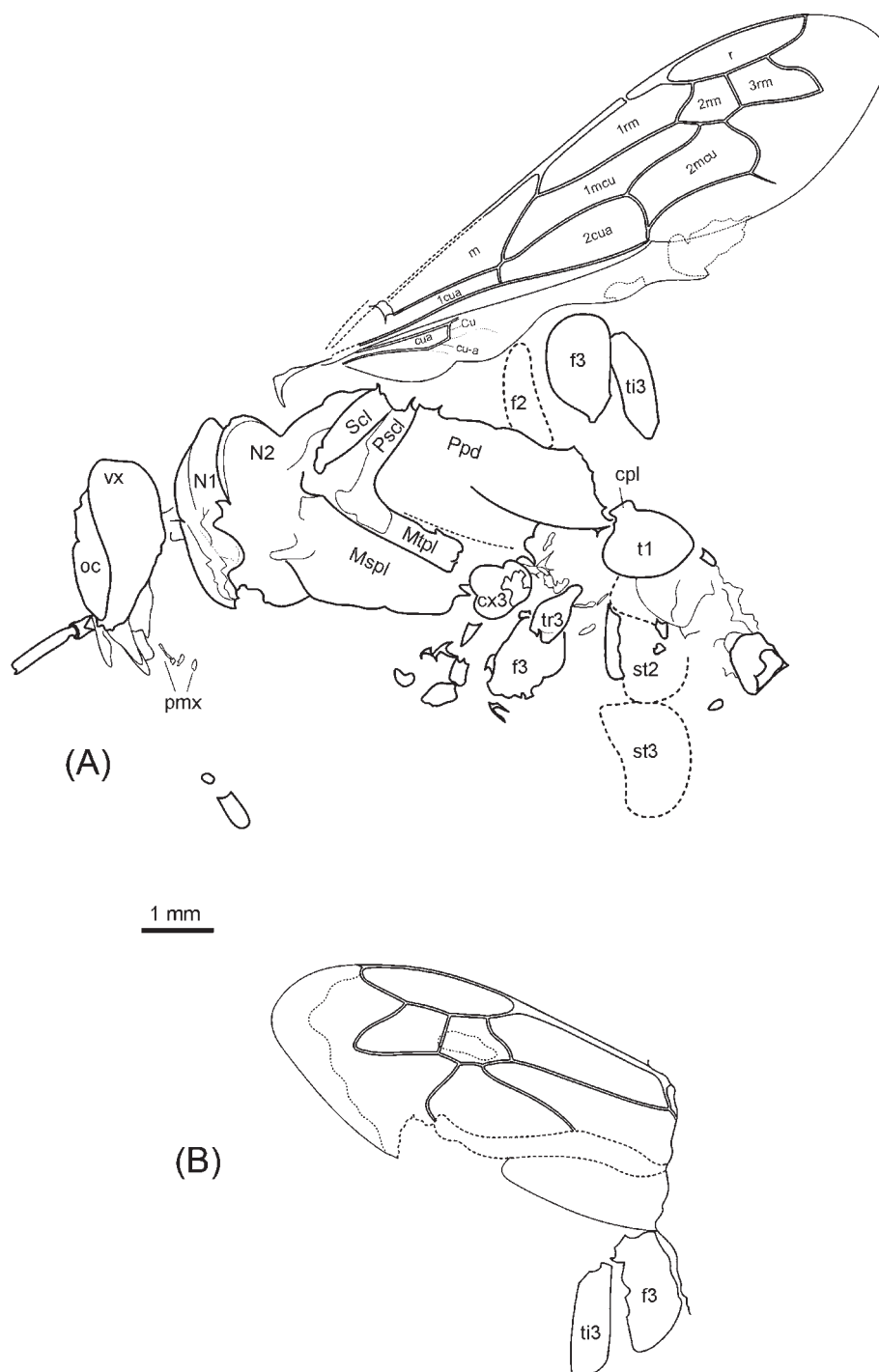
Genus *Tytthopsen* Antropov, gen. nov.

**Etymology.** After *tytthos*, the Greek for smallest, and genus *Psen*. Gender masculine.

**Type species.** *Tytthopsen nanus* sp. nov., by present designation and monotypy.

**Diagnosis.** The complete forewing venation of the type genus, with the elongate marginal cell which touches the wing margin with its acuminate apex, indicates it probably belongs to the crabronid subfamily Pemphredoninae.

In comparison with fossil genera, only two species of fossil pemphredonines are known to have three submarginal cells: *Palanga succinea* Budrys, 1993 and *Eomimesa rasnitsyni* Budrys, 1993, both from the Upper Eocene Baltic amber. Of them, *Palanga* differs from *Tytthopsen* in the following features: prStg at most one third as long as ptStg; 2r-rs received by outer third of ptStg and 1.5 times as long as 4RS; M and 1m-cu practically straight; 1m-cu received by 1rm, 2m-cu



**Text-figure 60** *Protosceliphron brevior* (Cockerell), line drawing of holotype NHMUK In.17472/In. 24340: (A) part; (B) counterpart. Body parts: cpl = connecting plate; cx3 = metacoxa; f2 = mid femur; f3 = hind femur; Msp = mesopleuron; Mtp = metapleuron; N1 = pronotum; N2 = mesonotum; oc = compound eye; pmx = maxillary palpus; Ppd = propodeum; Psc = postscutellum; Scl = scutellum; st2 = sternite II; st3 = sternite III; t1 = metasomal tergite I (abdominal tergum II); ti3 = hind tibia; vx = vertex. Wing cells: m = basal; 1cua = sub-basal; r = arginal; 1rm = submarginal I; 2rm = submarginal II; 2, 3rm = submarginal III; 1m-cu = discal I; 2m-cu = discal II; 1cua = subdiscal I; 2cua = subdiscal 2.

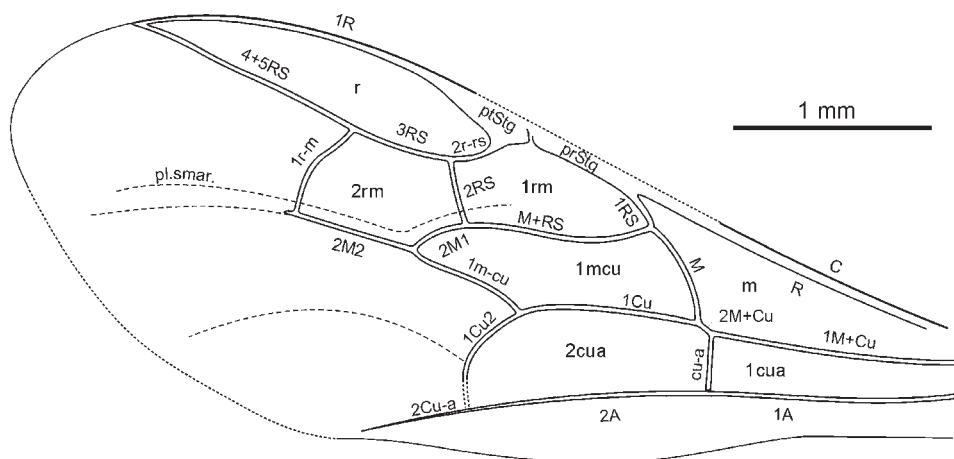
by 3rm; 1rm  $c.0.67$  times as long as marginal cell, more than four times as long as 2rm and at least three times as long 3rm.

*Eomimesa* differs from *Tytthopsen* in having prStg at most 0.25 times as long as ptStg; 2r-rs received by outer third of ptStg and almost half as long as 4RS; M and 1m-cu practically straight; 1m-cu and 2m-cu received by 2rm near its angles; M-a and 2Cu-a reaching outer wing margin; 1rm approximately 0.67 times as long as marginal cell, more than three times as long as 2rm and approximately 1.5 times as long as 3rm; 1m-cu  $c.4$  times as long as its maximum width.

In comparison with extant aculeates, the following living aculeates are similar to *Tytthopsen* in combining an apically acuminate marginal cell touching the wing margin, three submarginal cells, 1m-cu received by 2rm, and 2m-cu received by 3rm:

1. Male *Myrmosa* Latreille, 1796 (Mutillidae: Myrmosinae) differ from *Tytthopsen* in distinctly enlarged ptStg, marginal cell not strikingly acuminate apically and elongate (5RS not longer than 2r-m); in M-a and 2Cu-a reaching





Text-figure 62 *Plisomena gigantea* sp. nov., line drawing of holotype forewing, NHMUK In.24789.

differ from *Plisomena* in having 1m-cu received by 1rm, which is markedly larger than 2rm.

In comparison with extant genera, wing venation of *Plisomena* is almost typical for the subtribes Spilomenina and Stigmina. Among them only several species of the genus *Spilomena* Shuckard, 1838 (Spilomenina) have 1m-cu received by 2rm: *S. argentina* Antropov, 1992; *S. catamarca* Antropov, 1992; *S. willinki* Antropov, 1992; *S. kimseyae* Antropov, 1993; *S. nasuta* Antropov, 1993; *S. schlingeri* Antropov, 1993; and *S. tungurachua* Antropov, 1993. However, unlike *Plisomena*, all these species have a distinct angle between RS+M and 2M1. Additionally, the type species of *Plisomena* has its forewing more than 5 mm long, that is at least twice as long as even in the largest species of *Spilomena*.

*Plisomena gigantea* Antropov, sp. nov.

Plate 16, fig. 6; Text-fig. 62

**Etymology.** After *giganteus*, the Latin for giant.

**Holotype.** NHMUK In.24789, Bembridge Marls, NW Isle of Wight, UK, Hooley Collection. Sex unknown. The holotype is a piece of rock 28.3 × 31.9 mm, with an almost complete impression of the forewing. Judging by concave veins and convex submarginal wrinkle (Platesmar.), the fossil is a counterpart of the right forewing.

**Description. Diagnosis.** Forewing with single marginal, two submarginal and single discal cells; marginal cell with apex acuminate and touching wing margin; 1rm almost half as long as marginal cell and slightly longer than 2rm; 1mcu approximately twice as long as its maximum width and 1.5 times as long as 1rm; RS+M and 2M1 smoothly joining (not angulated); 1m-cu received by 2rm approximately 1/3 its length from 2RS; cu-a antefurcal.

Vein measurements in mm: prStg 0.60; ptStg 0.60; 1R 1.50; 2r-rs 0.27; 1RS 0.22; 2RS 0.35; 3RS 0.52; 4+5RS 1.23; 2M+Cu 0.10; M 0.52; RS+M 0.93; 2M1 0.28; 2M2 0.62; 1Cu1 0.90; 1Cu2 0.36; 2Cu2 0.23; 1m-cu 0.63; 1r-m 0.51; cu-a 0.30; 2Cu-a 0.53; 2A 1.22. Total length of forewing c.5.2 mm.

### 3.10.5.3. Family Apidae Latreille, 1802

Subfamily Apinae Latreille, 1802

Tribe ?Bombini Latreille, 1802

Genus *Oligobombus* Antropov, gen. nov.

**Etymology.** After Oligocene and genus name *Bombus*. Gender masculine.

**Type species.** *Oligobombus cuspidatus* sp. nov., by present designation and monotypy.

**Diagnosis.** Forewing distinctly pointed apically with complete venation, including marginal cell (r), three submarginal cells (1rm, 2rm, 3rm), basal (m), sub-basal (1cua), subdiscal (2cua), and two discal cells (1mcu, 2mcu); marginal cell elongate, distinctly longer than distance between its apex and wing tip, with non-parallel veins 5RS and 1R, 5RS uniformly curved backwards, and roundly truncate apex bearing appendicular vein Rs-a; forewing distal membrane distinctly papillate; ptStg comparatively short, with straight margin within marginal cell, 4.0 times as long as prStg, slightly longer than 3RS, but shorter than 4RS; 2r-rs arising from distal part of ptStg after its midpoint, as long as 3RS and 0.77 times as long as 4RS; 3RS shorter than 4RS; 4RS half as long as 5RS; 2RS practically straight; 1r-m 0.45 times as long as 3M1; RS+M as long as 2M1; angle between 1r-m and 2M2 acute; angle between 1r-m and 3M1 obtuse; 2M1 slightly shorter than 2M2; 2M2 almost 1.5 times shorter than 3M1; 3M1 almost 1.5 times as long as 4RS, almost 7.0 times as long as 3M2; 1rm with an oblique translucent vein rs and not wider than 2rm; 2rm shorter than 3rm, the widest submarginal cell; 1m-cu slightly curved anteriorly; 2m-cu uniformly curved outwards; distance between anterior ends of 1m-cu and 2m-cu distinctly exceeding their length; 1Cu2 twice as long as 2Cu2; 2Cu1 uniformly curved inwards; cu-a postfurcal, vertical and longer than 2M+Cu.

**Diagnosis.** Forewing venation complete, including marginal cell (r), three submarginal cells of approximately equal sizes (1rm, 2rm, 3rm), basal (m), sub-basal (1cua), subdiscal (2cua), and two discal cells (1mcu, 2mcu); marginal cell elongate, distinctly longer than distance between its apex and wing tip, with non-parallel veins 1R and 5RS, with 5RS uniformly curved backwards, and with roundly truncate apex bearing appendix Rs-a; ptStg comparatively short, with a straight margin within marginal cell; ptStg with straight margin within marginal cell, 4.0 times as long as prStg; 3RS shorter than 4RS; 1rm with an oblique translucent vein rs; 2RS straight basally; angle between 1r-m and 2M2 acute; 2rm shorter than 3rm; 1m-cu slightly curved anteriorly; distance between anterior ends of 1m-cu and 2m-cu distinctly exceeding their length; 2m-cu uniformly curved outwards; forewing distal membrane distinctly papillate.

**Remarks on the tribal position.** The complete forewing venation (including closed elongate marginal cell narrowed and not touching anterior wing margin apically, three submarginal

cells of approximately equal size, basal, sub-basal, subdiscal, and two discal cells) with comparatively short pterostigma and distance between anterior ends of recurrent veins distinctly exceeding their length allows the attribution of the studied specimen to the apid subfamily Apinae Latreille, 1802.

According to the last revision (Engel 2001), apines are divided into four extant tribes: Euglossini Latreille, 1802, Bombini Latreille, 1802, Apini Latreille, 1802 and Meliponini Lepeletier, 1836; and three fossil tribes: Electrobombini Engel, 2001, Electrapini Engel, 2001 and Melikertini Engel, 2001. The tribe Meliponini should be excluded because of its reduced venation and usually opened apex of the marginal cell. The tribe Apini should be also excluded because of its strongly elongate and parallel-sided marginal cell, strongly narrowed and obliquely elongate 3rm, and antefurcal cu-a.

The distinctly papillate forewing distal membrane of the studied specimen is characteristic of Electrobombini, Bombini and Euglossini, but not of Electrapini or Melikertini.

The marginal cell vein 5RS of the studied specimen is slightly curved outwards, as it is characteristic of Electrobombini, some Electrapini (*Thaumastobombus* Engel, 2001), and Melikertini (*Melikertes* Engel, 1998), but not of other Electrapini (some *Protobombus* Cockerell, 1908), Melikertini (*Succinapis* Engel, 2001, *Melissites* Engel, 2001) and the extant tribes Euglossini (5RS curved inwards) and Bombini (5RS curved inwards or sometimes almost straight).

An oblique translucent vein rs dividing 2RS of the studied specimen is characteristic of Electrobombini, Electrapini and Bombini, but not of Melikertini or Euglossini.

First transverse submarginal vein (2RS) of the studied specimen is practically straight as in Melikertini (*Succinapis* and *Melikertes*) and Euglossini, while it is S-curved basally in *Melissites* (Melikertini), Electrapini, Electrobombini, and all Bombini (2RS is even angular in many subgenera).

Submarginal cell II (2rm) of the studied specimen is distinctly narrower than submarginal cell III (3rm) as in Melikertini, Electrapini, Electrobombini and Euglossini, but not in Bombini, characterised by 2rm wider than 3rm.

The recurrent vein I (1m-cu) of the studied specimen is only slightly curved outwards preapically, as in Electrapini and Electrobombini, while it is strongly curved roundly in Electrapini (some *Protobombus*), Melikertini (*Succinapis* Engel, 2001, *Melissites* Engel, 2001) and some Euglossini and Bombini (for example, in subgenera *Bombias* Robertson, 1903, *Kallobombus* Dalla Torre, 1880, or *Thoracobombus* Dalla Torre, 1880), or distinctly angular in some Melikertini (*Melikertes* Engel, 1998), Euglossini and Bombini (for example, in subgenera *Alpinobombus* Skorikov, 1914, *Bombus* Latreille, 1802, *Exilobombus* Skorikov, 1922, *Megabombus* Dalla Torre, 1880, *Melano-bombus* Dalla Torre, 1880, *Pyrobombus* Dalla Torre, 1880, or *Subterraneibombus* Vogt, 1911).

The recurrent vein II (2m-cu) of the studied specimen is uniformly curved outwards, as in Electrapini, Melikertini and Euglossini, while it is S-shape in Electrobombini and almost all Bombini (excluding subgenera *Alpinobombus* or *Kallobombus*).

The studied specimen also differs from Electrobombini in 1rm not wider than 2rm, ptStg only slightly longer than 3RS, 2r-rs arising from distal part of ptStg after its midpoint, Rs-a present, 2Cu1 uniformly curved anterior, and cu-a longer than 2M+Cu.

The studied specimen differs from Bombini (characters in brackets) in marginal cell distinctly longer than distance between its apex and wing tip (as long as, or somewhat longer), with 5RS uniformly curved backwards (usually curved forwards), ptStg 4.0 times as long as prStg (shorter to slightly longer than prStg), 2RS almost straight (curved or angulate basally), 3RS shorter

than 4RS (as long as, or longer than 4RS), 1m-cu weakly curved anteriorly (roundly curved or distinctly angulate), and 2m-cu uniformly curved outwards (S-shaped).

Finally, an angle between 1r-m and 2M2 in the studied specimen is acute, while it is right or obtuse in all other apine tribes; excluding Apini, which distinctly differs in the form of strongly elongate marginal cell and submarginal cell II and strongly narrowed and obliquely elongate submarginal cell III. It is very likely that acute outer angles of 2rm in these two tribes have separate origin.

Thus, the forewing venation of the studied specimen is characterised by a set of features which, taken separately, can be found in different fossil and extant tribes of the apine bees, but taken together do not occur in any of them. At the same time, though the forewing venation of the studied specimen is generally similar to that of the extant tribe Bombini, absence of the most important features of the head, legs and mesosoma prevents identification of its exact tribal position.

**Diagnosis.** In comparison with fossil genera, the forewing venation of *Oligobombus* gen. nov. is similar to that of the monotypical genus *Electrobombus* Engel, 2001 from the Baltic amber (Electrobombini) in the papillate distal membrane, short ptStg with a straight margin within marginal cell, 2M1 almost equal to or slightly shorter than 2M2, and also in non-parallel veins 1R and 5RS.

*Oligobombus* gen. nov. differs from *Electrobombus* (characters in brackets) in having forewing longer than 13 mm (10.50 mm in *Electrobombus samlandensis* Engel, 2001, which is the largest known bee species preserved in amber of any deposit); marginal cell roundly truncated apically (acutely rounded); ptStg slightly longer than 3RS but shorter than 4RS (ptStg appreciably longer than 3RS and somewhat longer than 4RS); 2r-rs arising from distal part of ptStg after its midpoint, equal to 3RS and 0.77 times as long as 4RS (2r-rs arising from proximal part of ptStg before its midpoint, distinctly longer than 3RS and equal to 4RS); 4RS half as long as 5RS (c. 0.33 times); Rs-a present (absent); 1rm not wider than 2rm (distinctly wider); angle between 1r-m and 2M2 acute (obtuse); RS+M as long as 2M1 (1.5 times longer); 2M1 slightly shorter than 2M2 (as long as 2M2); 3M1 almost 1.5 times as long as 4RS (c. 2.0 times); 1m-cu 1.7 times as long as 2M1 (more than twice so); 2m-cu uniformly curved outwards (S-shaped); 1Cu2 twice as long as 2Cu2 (at least 4.0 times); 2Cu1 uniformly curved anterior (2Cu1 straight); and cu-a longer than 2M+Cu (the veins are equal).

*Oligobombus* gen. nov. is also similar to *Protobombus* Cockerell, 1909, *Electrapis* Cockerell, 1909 and *Thaumastobombus* Engel, 2001 (Electrapini), all from the Late Eocene Baltic amber, in the roundly truncate apex of marginal cell bearing an appendix Rs-a and approximately equal veins RS+M and 2M1.

*Oligobombus* differs from *Protobombus* (in brackets) in the forewing longer than 13 mm (3.9–7.4 mm); ptStg with straight margin within marginal cell (convex); ptStg slightly longer than 3RS and shorter than 4RS (at least twice as long as 3RS and distinctly longer than 4RS); 2r-rs arising from distal part of ptStg after its midpoint (from the midpoint of ptStg); 2r-rs equal in length to 3RS (distinctly longer than 3RS); 2r-rs 0.77 times as long as 4RS (almost equal in length to 4RS); 2RS almost straight (S-shaped); 4RS half as long as 5RS (c. 0.33–0.4 so); 1rm not wider than 2rm (distinctly wider); angle between 1r-m and 2M2 acute (right or obtuse); 2M1 somewhat shorter than 2M2 (distinctly longer); 3M1 almost 1.5 times as long as 4RS (c. twice as long as); 1m-cu 1.7 times as long as 2M1 (not more than 1.3 times so); 1m-cu slightly curved preapically (angulate preapically); 2m-cu uniformly curved outwards (angulate preapically); 1Cu2 twice

as long as 2Cu2 (2.5–3.0 times longer); and cu-a longer than 2M+Cu and postfurcal (cu-a still longer or interstitial).

*Oligobombus* gen. nov. also differs from *Electrapis* and *Thaumastobombus* (in brackets) in having forewing longer than 13 mm (6.38–9.17 mm in *Electrapis* and 4.45 mm in *Thaumastobombus*); small ptStg (enlarged) with straight margin within marginal cell (with convex margin within marginal cell) and longer than 3RS and shorter than 4RS (slightly longer than 3RS and 4RS); 4RS half as long as 5RS (at least 0.4 times as long as 5RS); 2r-rs equal in length to 3RS (distinctly shorter than 3RS in *Electrapis* and somewhat longer than 3RS in *Thaumastobombus*); 1rm with an oblique translucent vein rs (1rm without rs); 1rm not wider than 2rm (slightly wider); 2RS almost straight (more or less S-shaped in *Electrapis* and angulate inwards in *Thaumastobombus*); 2M1 somewhat shorter than 2M2 (distinctly longer); angle between 1r-m and 2M2 acute (obtuse); 1m-cu 1.7 times as long as 2M1 (not more than 1.2 times so); 2m-cu uniformly curved outwards (sometimes straight in *Thaumastobombus*); 1Cu2 twice as long as 2Cu2 (2.5–3.0 times so in *Electrapis*); and cu-a vertical (posterior end of cu-a moved distally in *Electrapis* and distinctly moved proximally in *Thaumastobombus*).

Finally, *Oligobombus* gen. nov. also differs from *Succinapis*, *Melikertes* and *Roussyana* (Manning, 1961) (all Melikertini) in the not narrowed anteriorly submarginal cell II (2r-rs as long as 3RS).

In comparison with extant genera, the forewing venation of *Oligobombus* gen. nov. is generally similar to that of extant members of the genus *Bombus* Latreille, 1802 in papillate distal membrane; in short ptStg with a straight margin within R; in 2r-rs arising distally from the midpoint of ptStg; and in abscissa 2M1 almost equal in length or only slightly shorter than 2M2. *Oligobombus* gen. nov. differs from *Bombus* (in brackets) in marginal cell distinctly longer than distance from its apex to the wing tip, wider and with non-parallel 1R and 5RS, with 5RS almost straight or uniformly curved backwards, and roundly truncate apex (marginal cell narrow, somewhat longer than distance from its apex to the wing tip, with almost parallel 1R and 5RS, which is curved forwards basally, or sometimes almost straight, and rounded apically); ptStg almost four times longer than prStg (ptStg little if any longer than prStg); 2r-rs equal in length to 3RS (2r-rs distinctly shorter than 3RS); 2RS almost straight (curved or angulate basally); 3RS only slightly shorter than 4RS (3RS 0.67 times as long as 4RS); 4RS half as long as 5RS (0.26–0.29 times as long as 5RS); 1r-m 0.45 times as long as 3M1 (practically equal in length); 2rm narrower than 3rm (2rm mainly wider than 3rm), 3rm less narrowed anterior and the widest among submarginal cells; (3rm strongly narrowed anteriorly, and 2rm is the widest submarginal cell); 2M2 0.7 times as long as 3M1 (practically equal to 3M1); angle between 1r-m and 2M2 acute (obtuse or sometimes right); 3M1 almost 1.5 times as long as 4RS (c. 2.0 times as long as 4RS); 3M1 almost seven times as long as 3M2 (c. 2.5–3.0 times as long as 3M2); 1m-cu weakly curved anteriorly (roundly curved or distinctly angulate); 2m-cu uniformly curved outwards (S-shaped); and 1Cu2 twice as long as 2Cu2 (approximately equal in length).

*Oligobombus cuspidatus* Antropov, sp. nov.

Plate 17, figs 1–2; Text-fig. 63

**Etymology.** After *cuspidatus*, the Latin for pointed, alluding to the form of the wing tip.

**Holotype.** NHMUK In.17349 (part and counterpart), Smith Collection. Sex unknown. The part is a piece of rock 65.2 × 23.6 mm, with the impression of the well preserved middle

and apical parts of the right forewing. The counterpart is a piece of rock 67.4 × 24.8 mm, with the impression of the well preserved middle part of the right forewing.

**Description.** Forewing distinctly pointed apically with complete venation, including marginal cell (r), three submarginal cells (1rm, 2rm, 3rm), basal (m), sub-basal (1cua), subdiscal (2cua), and two discal cells (1mcu, 2mcu); marginal cell elongate, distinctly longer than distance between its apex and wing tip, with non-parallel veins 5RS and 1R, 5RS uniformly curved backwards, and roundly truncate apex bearing appendicular vein Rs-a; forewing distal membrane distinctly papillate; ptStg comparatively short, with straight margin within marginal cell, four times as long as prStg, slightly longer than 3RS, but shorter than 4RS; 2r-rs arising from distal part of ptStg after its midpoint, as long as 3RS and 0.77 times as long as 4RS; 3RS shorter than 4RS; 4RS half as long as 5RS; 2RS practically straight; 1r-m 0.45 times as long as 3M1; RS+M as long as 2M1; angle between 1r-m and 2M2 acute; angle between 1r-m and 3M1 obtuse; 2M1 slightly shorter than 2M2; 2M2 almost 1.5 times shorter than 3M1; 3M1 almost 1.5 times as long as 4RS, almost seven times as long as 3M2; 1rm with an oblique translucent vein rs and not wider than 2rm; 2rm shorter than 3rm, the widest submarginal cell; 1m-cu slightly curved anteriorly; 2m-cu uniformly curved outwards; distance between anterior ends of 1m-cu and 2m-cu distinctly exceeding their length; 1Cu2 twice as long as 2Cu2; 2Cu1 uniformly curved inwards; cu-a postfurcal, vertical and longer than 2M+Cu.

Vein measurements in mm: prStg 0.20; ptStg 0.83; 1R 3.70; 2r-rs 0.75; 3r-rs 0.35; 1RS 0.60; 2RS 0.80; 3RS 0.75; 4RS 0.95; 5RS 2.00; 2M+Cu 0.20; M 2.60; RS+M 1.20; 2M1 0.76; 2M2 0.97; 3M1 1.40; 3M2 0.21; 1Cu1 2.60; 1Cu2 0.84; 2Cu1 2.75; 2Cu2 0.40; 1r-m 0.63; 2r-m 1.26; 1m-cu 1.37; 2m-cu 1.27; cu-a 0.36; 2A 2.85; Rs-a 0.42; M-a 0.46; 2Cu-a 0.47. Forewing apex is distinctly pointed. Total length of forewing c. 13.25 mm.

### 3.10.5.4. Family Apidae Latreille, 1802

Subfamily Megachilinae Latreille, 1802

Megachilinae *incertae sedis*

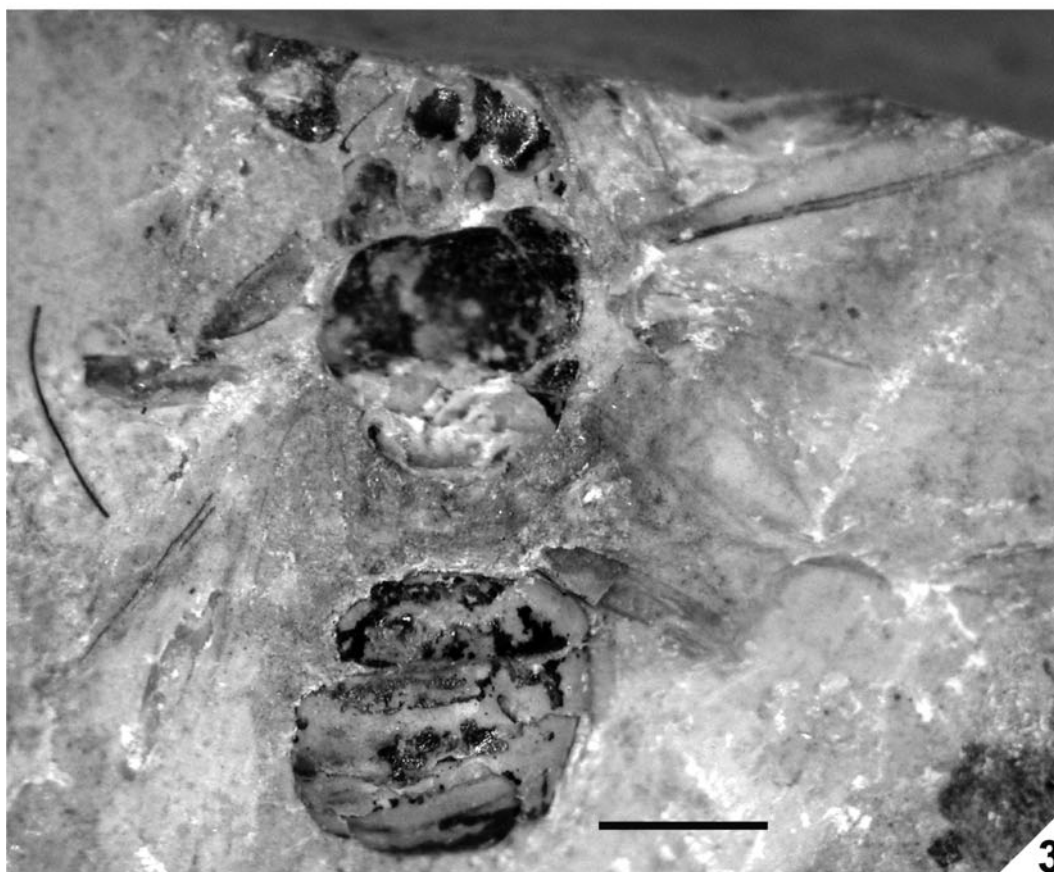
Plate 17, fig. 3; Text-fig. 64

**Material studied.** NHMUK I.10012, Bembridge Marls, NW Isle of Wight, UK, Brodie Collection. Sex unknown. A piece of rock 42.3 × 30.8 mm, with an impression of the partially preserved mesosoma, metasoma, fore coxae and fore and mid femora, a poorly preserved head, and a few longitudinal veins of forewings.

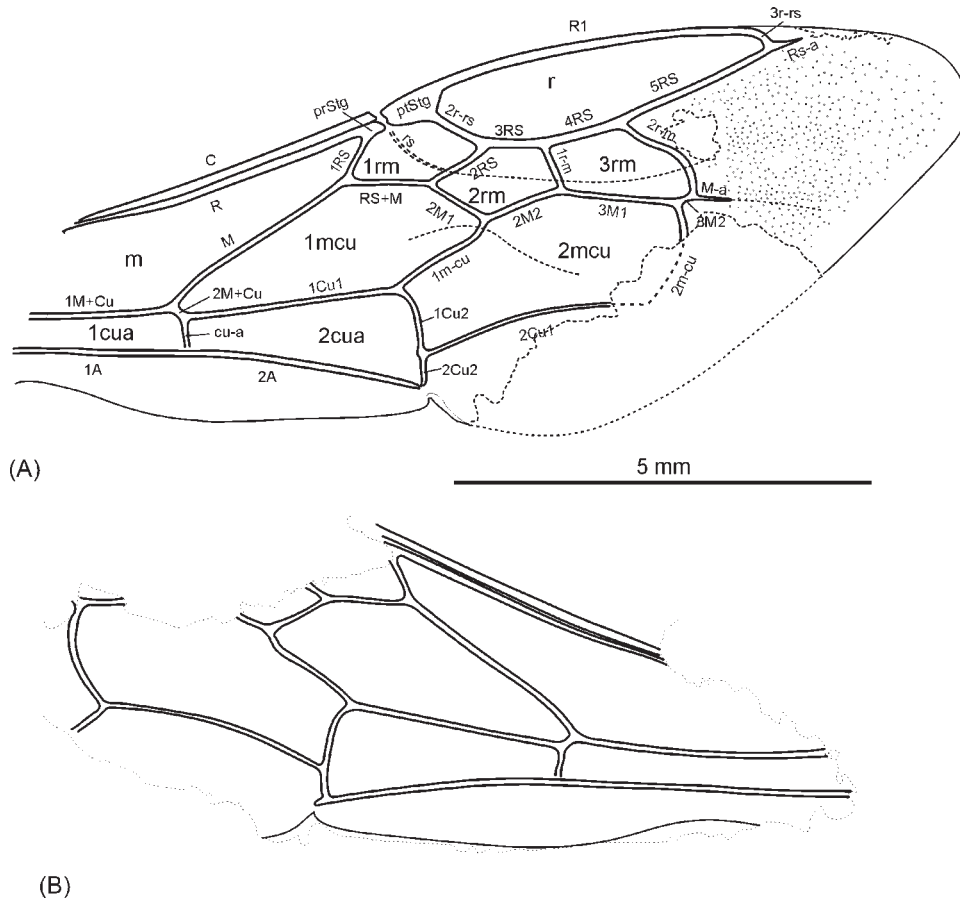
**Description.** Cuticle black, distinctly punctate; fore coxae, fore and mid femora, hind tibiae and tarsi reddish-brown; mesosoma rounded, with very short propodeum; metasoma robust, with middle tergites at least twice wide as long; constrictions between metasomal tergites distinct. Length of body parts in mm: scutum 1.17; mesosoma c. 1.66; metasoma c. 2.05. Scutal width 1.55; metasomal width 1.78. Total body length c. 4.7 mm.

**Remarks.** The flattened and moderately widened hind tibiae and basal tarsomeres and robust metasoma with convex tergites, separated by distinct constrictions, attribute the specimen to several genera of the subfamily Megachilinae Latreille, 1802, but absence of the most important features of wing venation and structure of the head and mesosoma prevents identification of its generic position.

**3.10.6. Discussion.** A comparison of the studied fauna of Aculeata, in the vicinity of the Eocene/Oligocene boundary, of Bembridge Marls, Isle of Wight, England, with two faunas



**Plate 17** (1–2) *Oligobombus cuspidatus* sp. nov., holotype forewing NHMUK In. 17349: (1) part; (2) counterpart. (3) Family Megachilidae Latreille, 1802, *incertae sedis*, head, thorax, middle part of metasoma, fore and mid femora, and hind tibia and tarsus, NHMUK In.10012. Scale bars = 1 mm.



**Text-figure 63** *Oligobombus cuspidatus* sp. nov., line drawings of holotype forewings, NHMUK In.17349: (A) part; (B) counterpart.



**Text-figure 64** Family Megachilidae Latreille, *incertae sedis*, line drawing of body, NHMUK In.10012. Body parts: cx1 = procoxae; f1–3 = fore, mid and hind femora; ti3 = hind tibia; 1t3 = hind tarsomere 1; 2t3 = hind tarsomere 2; t2–5 = metasomal tergites II–V (abdominal terga III–VI).

of similar age: the Upper Eocene Baltic amber and the uppermost Eocene shales of Florissant (USA, Colorado) are of obvious interest.

Various xylobionts are common in Baltic amber. Species of the genera *Vespa*, *Palaeovespa* and *Polistes* (Vespidae) probably built their nests on or inside trees and prey directly in their crowns. Several genera of Apidae (*Electrobombus*, *Electrapis*, *Protobombus*, *Thaumastobombus*) were also discovered in Baltic amber and, though nothing is known about their nesting habits, they were probably forest inhabitants which could build underground nests as do extant species of Bombini. It is necessary to take into consideration that Vespidae and Apidae are not particular xylobionts, because their provisioning territories may spread far from the nest, and their chances of being fossilised in resin depend directly on the density of populations. Representatives of the families Ampulicidae (*Protodolichurus* Nemkov, 1988, *Dolichurus* Latreille, 1809) and Crabronidae (*Eopinoecus*, *Eoxyloecus*, *Passaloecus* Shuckard, 1837, *Pison* Jurine, 1808, *Tracheliodes* A. Morawitz, 1866 and, judging from the absence of foretarsal digging rakes, *Palanga* and *Eomimesa*) also hunt in the same biotopes. Species of Ampulicidae used available cracks of the tree trunks for concealing their prey, and species of Crabronidae, as well as comparatively small bees of the families Megachilidae (*Glyptapis* Cockerell, 1909, *Ctenoplectrella* Cockerell, 1909, *Glaesosmia* Engel, 2001), like their extant relatives, built their nests either in wood or inside old borings of various xylophagous insects. Obviously, these groups are closely linked to trees and had more chances to be fossilised in resin. The only exception to the rule is *Tiphia* (?) sp., which was mentioned by Brischke (1886) from the lost collection of Menge.

The more diverse fauna of the Florissant beds is somewhat different. Though Vespidae (*Palaeovespa*, *Polistes*) and even real xylobionts (Crabronidae, Pemphredoninae – *Passaloecus*) are present here, the prevailing groups are connecting with prey inhabiting soil (Tiphidae, Myziniinae – *Lithotiphia* Cockerell, 1906, *Geotiphia* Cockerell, 1906; Tiphinae – *Paratiphia*; Scoliididae, Archaeoscoliinae – *Floriscolia* Rasnitsyn, 1993); with digging underground nests (Sphecidae, Ammophilinae – *Ammophila* Kirby, 1798; Crabronidae, Crabroninae – *Tracheliodes*, *Ectemnius* Dahlbom, 1845, Mellininae – *Mellinus* Fabricius, 1790, Bembicinae – *Gorytes* Latreille, 1805 (?*Psammaletes* Pate, 1936), *Psammaecius* Lepeletier, 1832, Philanthinae – *Philanthus* Fabricius, 1790, *Prophilantus* Cockerell, 1906); or so-called “mud-daubers”, who use moist clay from near water for building their nests (Sphecidae, Sceliphrinae – *Chalybion* Dahlbom, 1843, *Hoplisdia* Cockerell, 1906; Crabronidae, Crabroninae – *Pison*). Many other groups of Hymenoptera found in Florissant shales inhabited the shore territories of the fresh water lakes and also included ground-nesting or often weakly flying forms.

The composition of the aculeate wasp and bee fauna found in the Bembridge Marls, Isle of Wight, England, demonstrates wider generic diversity than is characteristic of the contemporary Palaeoarctic region. The studied fauna includes both xylobionts (*Palaeopolybia*, *Protopolistes*, *Sphaerancistrocerus*, *Tythopsen*, *Plisomena*, a megachilid bee and, probably, *Dryophia*) and ground-dwelling forms (*Philoponites*, *Palaeoscolia*, *Protosceliphron*, and *Oligobombus*). The most interesting fact is the high level of endemism; none of the discovered taxa were previously known from other formations. Another peculiarity is the presence of archaic forms, for instance the polistine wasp *Palaeopolybia*, which retains a vespine-like structure of its forewing submarginal area; *Protosceliphron* – the only known Sphecidae with a sessile metasoma lacking sternal petiole; and *Plisomena* – the only known Spilomenina with plesiomorphic (not enlarged) pterostigma. Presence of such

a generalised feature as a weak degree of costalisation of the anterior ends of recurrent veins in *Palaeopolybia*, *Tythopsen*, *Plisomena* and *Oligobombus* is also a very remarkable peculiarity of the studied fauna, because it is more archaic than those of the earlier resembling taxa. I think that it was a locally endemic fauna, probably isolated from the surrounding regions.

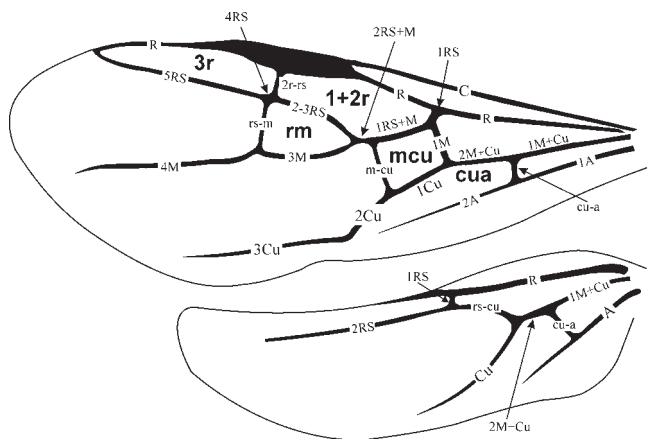
There are comparatively low numbers of Aculeata in Bembridge Marls (excluding winged ants, whose quantity is much more than of all other groups of Aculeata) and parasitic Hymenoptera. The first group perished in mass after swarming, while the second could easily be blown from the coast because of its comparatively weak flying abilities. However, this cannot explain the low quantity of Aculeates in the adjacent territories, because actively flying aculeate wasps and bees are not connected with a sea coast, where they can get neither prey nor building materials. That is why the probability of their fossilisation in such conditions was quite low. Nevertheless, the discovery of such taxa as Tiphidae, Scoliididae and Apidae indicates that large meadows with grasses and long-flowered herbs were present near the region of fossilisation. The occurrence of Spilomenina and small eumenine wasps occupying pre-existing borings in wood suggests the presence of broad-leaved trees. The comparatively weak degree of costalisation of the wing venation and significant sizes of the marginal cell in the discovered pemphredonines also testifies to their living in forest or forest-steppe regions. Furthermore, some taxa of apparently tropical affinities (*Palaeopolybia*) indicate a warm climate. I suggest that the region of fossilisation was a coastal plain with a warm climate, resembling the modern subtropical habitats of the Mediterranean region.

### 3.11. Superfamily Formicoidea Latreille, 1802

[By Gennady M. Dlussky and Ksenia S. Perfilieva.]

**3.11.1. Family Formicidae Latreille, 1802.** Ants are the most abundant group of insects in the Bembridge Marls. The first brief report on a finding of ants in the Bembridge Marls, with no description of species, appeared in the late 19th Century (Brodie 1875). A few years later Woodward (1879) referred to identifications by F. Smith and noted that ants from these deposits belong to genera *Myrmica* Latreille, 1804, *Formica* Linnaeus, 1758 and *Camponotus* Mayr, 1861. Donisthorpe (1915) reproduced this information. The same year, Cockerell (1915) finished the study of a small collection from the Bembridge Marls collected by Brodie (transferred to the United States National Museum as a part of the Lacoe collection) and published the first descriptions of species. His descriptions were based mainly on measurements of cells and veins of forewings of a few specimens. He described eight species and referred them to four subfamilies: *Ponera hypolitha* Cockerell, 1915 (Ponerinae); *Leptothorax gurnetensis* Cockerell, 1915 (Myrmicinae); *Dolichoderus britannicus* Cockerell, 1915, *D. anglicus* Cockerell, 1915, and *D. ovigerus* Cockerell, 1915 (Dolichoderinae); and *Oecophylla atavina* Cockerell, 1915, *O. perdita* Cockerell, 1915, and *O. megarche* Cockerell, 1915 (Formicinae).

Donisthorpe (1920) undertook revision of a large collection of fossils from the Bembridge Marls, collected by P. B. Brodie, E. J. A'Court Smith and R. W. Hooley and kept at the Natural History Museum, London. In this article, he demonstrated that the species described by Cockerell as *Leptothorax gurnetensis* actually belongs to the subfamily Formicinae, and placed it in a new genus *Leucotaphus* Donisthorpe, 1920. He also described seven new species: *Syntaphus wheeleri* Donisthorpe, 1920; *Euponera* (*Mesoponera*) *crawleyi* Donisthorpe, 1920; *Ponera minuta* Donisthorpe, 1920, *Emplastus emeryi* Donisthorpe, 1920 (Ponerinae); *Dolichoderus gurnetensis* Donisthorpe,



**Text-figure 65** Wing venation of *Emplastus britannicus*, to show venational symbols used. Capitalised are longitudinal veins; dashed are cross-veins; neither of above are cells.

1920 (Dolichoderinae); *Leucotaphus cockerelli* Donisthorpe, 1920; and *Camponotus (Colobopsis) brodiei* Donisthorpe, 1920 (Formicinae). Later Cockerell (1927) described one more species, *Leucotaphus permaneus* Cockerell, 1927. There are no later publications about the ant fauna of the Bembridge Marls.

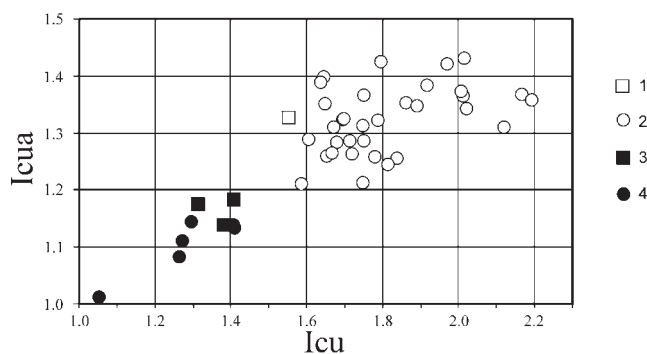
We studied 1136 specimens of fossil ants kept at the NHMUK including all of Donisthorpe's type specimens. We were able to study all of Cockerell's types, stored at the USNM (15 specimens) as well. It has only been possible to identify 105 of the specimens up to family level, due to insufficient preservation. The remaining 1031 specimens have been identified up to genus and 630 to species level. Digital photos have been prepared for 245 of the best preserved fossils, and some of them served as a base for line drawings.

Among the studied material, 234 specimens were earlier identified by Donisthorpe. Fifteen of them are not ants, and the references of 13 other fossils to ants seem doubtful. During the present revision, we found that Donisthorpe's article (1920) contains a number of mistakes, both in respect to definition of specimens and to taxonomic position of described species. There is an example, which can attest to the quality of this work: Donisthorpe described specimen NHMUK I.8675 as the holotype of *Euponera (Mesoponera) crawleyi* (Ponerinae) and identified its counterpart (I.8539) as *Leucotaphus gurnetensis*, hence referring it to another subfamily (Formicinae).

Some species described by Cockerell and Donisthorpe have been either synonymised or excluded from Formicidae, with only ten of 17 described species currently accepted, and the taxonomic status of six of them is changed. Additionally we describe ten new species below.

Preservation of the Bembridge fossils is unusual. In fact, they are not impressions but cavities. As a result, we see an insect from the inside, unlike other deposits where fossils are seen from the outside. This creates additional difficulties for interpretation. In particular, the fossils rarely split along the middle plane and as a result the insect contour looks deformed. It is rarely possible to combine part and counterpart. Also, measurements and their ratios are more variable than it is expected from intact ants.

In the line drawings, lines trace visible sclerite boundaries, dashed lines trace supposed sclerite boundaries, and dotted lines – visible margins of incomplete sclerites. Filled contours denote sclerites with the original matter preserved, even if chemically modified. Wing morphometrics were analysed using standard methods (cf. Bookstein 1991; Rohlf & Marcus 1993; Danforth 1989; Rust 1999). Measurements of larger structures



**Text-figure 66** Differences of position of cross-vein cu-a in Aneuretinae, Dolichoderinae and Ponerinae from Bembridge Marls. Symbols: 1 = *Britaneuretus anglicus* (Aneuretinae); 2 = Dolichoderinae; 3 = *Taphopone petrosa* (Ponerinae); 4 = other Ponerinae.

(total body length, length of alitrunk, forewing, etc.) were made using an ocular micrometer. Smaller details (petiole length, height and width, maximum eye diameter, etc.) were measured from digital pictures.

**Abbreviations of measurements taken:** AL = alitrunk (mesosoma) length from junction with head to that with petiole; AW = maximum alitrunk width; BL = total body length; ED = maximum eye diameter; F1 = fore femur length; F3 = hind femur length; FWL = forewing length; FWW = forewing width measured between the pterostigmal base and indentation of wing hind margin next to junction of sections 2Cu and 3Cu (Text-fig. 65); HL = head length without mandibles; HW = maximum head width; n = number of specimens measured; PtL = petiole length; PtH = maximum petiole height; PtW = maximum petiole width; SL = scape length; ScL = scutum length; ScW = scutum width; SctL = scutellum length; SctW = scutellum width.

Nomenclature of the wing venation (Text-fig. 65) is as in our previous publications (Dlussky 1983; Perfilieva 2002; Dlussky & Perfilieva 2003). Two indices are used to help discriminate ant subfamilies: these concern relative position of cross-vein cu-a and are calculated as follows:

$$Icu = [1Cu + (2M + Cu)] / 1Cu,$$

$$Icua = [(1M + Cu) + (2M + Cu)] / (1M + Cu).$$

They are particularly helpful in the distinguishing of Ponerinae and Dolichoderinae (Text-fig. 66). The second index is more precise, because Icu depends not only on the position of cu-a but also on the size of cell mCu. However, Icu is indispensable in the case of incomplete wings when Icua is not available for consideration.

Other terminology of ant morphology follows Bolton (1994). Statistics of the living and extinct genera and species are after Bolton (1995), corrected to consider overlooked and subsequent publications.

### 3.11.1.1. Subfamily Aneuretinae Emery, 1913

**Taxa included.** Subfamily Aneuretinae includes the unique extant species *Aneuretus semoni* Emery, 1893 that inhabits Sri Lanka (Wilson *et al.* 1956). Seven fossil species attributed to Aneuretinae were also described: *Cananeuretus occidentalis* Engel & Grimaldi, 2005 (Upper Cretaceous, Canadian amber); *Aneuretellus deformis* Dlussky, 1988 (Paleocene, Sakhalin amber); *Protaneuretus succineus* Wheeler, 1915, *Paraneuretus longicornis* Wheeler 1915 and *P. tornquisti* Wheeler, 1915 (Late Eocene, Baltic amber); *Mianeuretus mirabilis* Carpenter, 1930 (latest Eocene, Florissant); and *M. eocenicus* Dlussky & Rasnitsyn, 2002 (Middle Eocene, Green River). The incomplete

*Burmomyrma rossi* Dlussky, 1996 (mid-Cretaceous, Burmese amber) may also belong to this subfamily.

Genus *Britaneuretus* Dlussky & Perfilieva, gen. nov.

**Etymology.** After Britain and genus *Aneuretus*. Gender masculine.

**Type species.** *Dolichoderus anglicus* Cockerell, 1915.

**Diagnosis.** Female. Propodeum with a pair of short, obtuse teeth. Petiole elongate, without scale or distinct node. Forewing with cells 1+2r, rm, mcu and most likely 3r closed. 1RS and 2r-rs vertical to R. Cell mcu medium sized (visually about half area of cell 1+2r or little less), far distant of pterostigmal base level. Icu > 1.5, Icu<sub>a</sub> > 1.2. Gaster oval, without constriction between first and second segments.

**Remarks.** Differs from *Cananeuretus* Engel & Grimaldi, 2005 from the Upper Cretaceous Canadian amber, *Protaneuretus* Wheeler, 1915 and *Paraneuretus* Wheeler, 1915 from the Late Eocene Baltic amber, *Mianeuretus* Carpenter, 1930 from the Middle and latest Eocene of North America, and from extant *Aneuretus* Emery, 1893 by absence of petiolar node. *Protaneuretus* and *Cananeuretus* have no propodeal teeth. *Aneuretelus* Dlussky, 1988 from the Palaeocene Sakhalin amber has no petiolar node but, unlike the new genus, has no propodeal teeth. The only known species *A. deformis* is of very small size (body length about 3 mm, alitrunk length 0.88 mm).

Although no sting is visible in the fossil, a combination of petiole form and features of forewing venation indicate position of the new genus in Aneuretinae with sufficient confidence.

*Britaneuretus anglicus* (Cockerell, 1915), comb. nov.  
Plate 18, fig. 1; Text-fig. 67.

1915 *Dolichoderus anglicus* Cockerell, p. 484, plate 65. fig. 8.  
1978 *Dolichoderus anglicus* Cockerell; Burnham, p. 112.  
1995 *Dolichoderus anglicus* Cockerell; Bolton, p. 172.

**Holotype.** USNM 61415, Lacoe Coll. Bembridge Marls, Isle of Wight, UK.

**Description.** Female. Propodeum angular in side view, with blunt teeth on boundary of propodeal dorsum and declivity; propodeal dorsum longer than propodeal declivity. Petiole 1.7 times as long as high and about half as long as propodeum. Anterior part of petiolar dorsum almost flat, posterior one feebly rounded in side view. Petiole with maximal height behind its midlength. Forewing: distance from 1RS to pterostigma more than 1.5 times as long as RS+M. 1M inclined and parallel to m-cu. RS+M nearly 0.65 as long as 2-3RS. Cell mcu trapezoid (RS+M and 1Cu noticeably longer than m-cu and 1M). Icu = 1.6, Icu<sub>a</sub> = 1.3.

Measurements of holotype, mm: AL ~ 2.6; FWL ~ 8; PtL = 1.2; PtH = 0.7.

**Remarks.** Cockerell described *Dolichoderus anglicus* using only characters of forewing venation and ignored preserved body fragments. The wing venation is characteristic not only of *Dolichoderus*, but also of many other genera. At the same time, the form of the propodeum and petiole of the fossil is strictly unlike that of Dolichoderinae and particularly of *Dolichoderus*.

Cross-vein 2r-rs occurring distally in respect to rs-m appears to be an aberration in ant wings, being present often in only one wing of the pair. Therefore, we consider this feature as an aberration of the holotype and not as a diagnostic character of the taxon.

### 3.11.1.2. Subfamily Dolichoderinae Forel, 1878

Genus *Dolichoderus* Lund, 1831

**Type species.** *Formica attelaboides* Lund, 1831, by monotypy.

**Other species.** The genus *Dolichoderus* comprises 110 species in the modern fauna, the vast majority of which are distributed in the Neotropical (54) and Indo-Australian and Australian (41) regions. Only nine species are known from the Oriental region, four from the Nearctic region, two from the Palearctic region, and none were recorded from the Afrotropical region. The oldest species, *D. kohlsi* Dlussky & Rasnitsyn, 2002, is recorded in the Middle Eocene Formation Green River, USA. Sixteen species are described from the Late Eocene Baltic and Rovno ambers (Dlussky 2002; Dlussky & Perkovsky 2002) seven species from the near-boundary Eocene/Oligocene deposits: two from Florissant, USA (Carpenter 1930), five from the Isle of Wight, UK (Donisthorpe 1920), and six species from the Oligocene of continental Europe (Théobald 1937). Thirteen species are described from the Miocene of Central Europe (Radoboj, Croatia) and East Europe (Kerch, Ukraine), Eastern Asia (Shanwang, Shandong, China) and Central America (Dominican amber) (Mayr 1867; Dlussky 1981a; Wilson 1985; Zhang 1989; Zhang *et al.* 1994). However, after revision of the European fossil species, this number will undoubtedly change. As shown below, only one of five species described as *Dolichoderus* from the Bembridge Marls belongs there.

**Diagnosis** (for rock fossils of females and workers). Female. Head elliptical or oval, widest behind midlength of head sides; sometimes retort-shaped or subrectangular with rounded occipital corners, but never rectangular and longer than wide as in *Protazteca* Carpenter. Eyes of moderate size. Mandibles triangular, dentate. Propodeum bispinate, bidentate, or distinctly angular in side view, always with concave declivity. Petiole with scale proclined, or triangular in side view. Integument thick, often coarsely sculptured. Forewing with cells 1+2r, 3r, rm and mcu closed; rm usually triangular. 1RS vertical to R or slightly proclined. Cell 3r touching wing margin, cross-veins 2r-rs and rs-m coincide on RS, or rs-m slightly distal. Cell mcu medium-sized, rhomboid or trapezoid (1Cu appreciably longer than RS+M). Gaster oval, without constriction between first and second segments. Integument thick, often coarsely sculptured.

**Remarks.** Almost all modern species of the genus are dendrobionic, and there are no reasons to consider the present fossils as different.

*Dolichoderus vectensis* Donisthorpe, 1920  
Plate 18, figs 2-3; Text-fig. 68A-C.

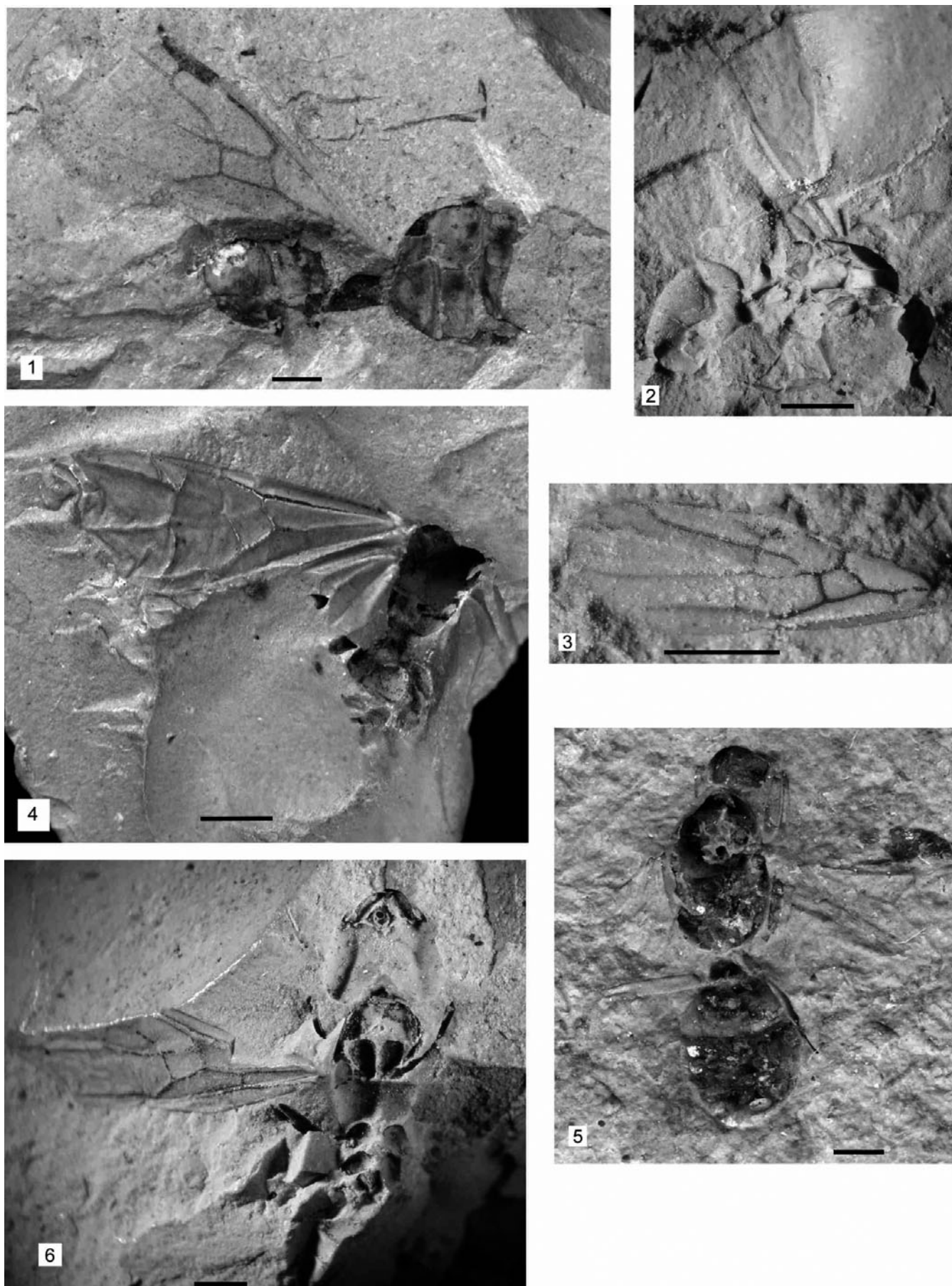
1920 *Dolichoderus vectensis* Donisthorpe, p. 88, plate V. fig. 6.  
1978 *Dolichoderus vectensis* Donisthorpe; Burnham, p. 112.  
1995 *Dolichoderus vectensis* Donisthorpe; Bolton, p. 177.

**Holotype.** NHMUK I.9198, ♀, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie (Plate 18, fig. 2; Text-fig. 68A).

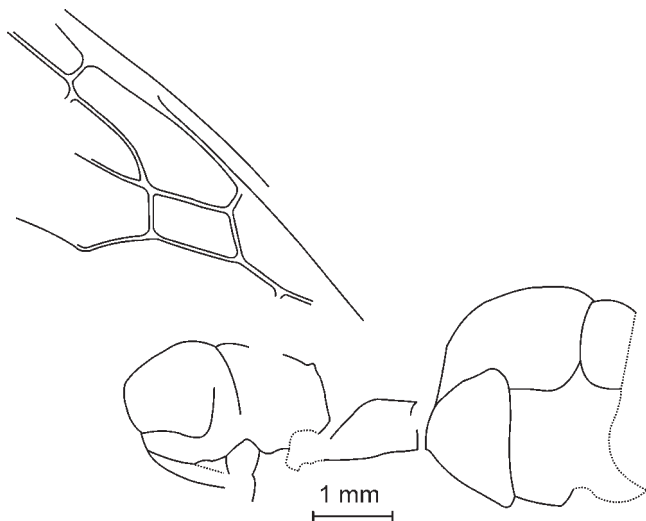
**Other material.** Two females and four forewings in NHMUK collection, including well preserved lateral imprint of the female NHMUK In.24934 (Text-fig. 68B) and forewing NHMUK I.9373 (Plate 18, fig. 3; Text-fig. 68C). Bembridge Marls, NW Isle of Wight, UK, .

**Diagnosis.** Similar to extant and extinct species of *D. quadri-punctatus* group, particularly to *D. tertiaris* (Mayr 1868) from the Baltic amber and *D. polessus* Dlussky, 2002 from the Rovno amber (Late Eocene), but differs from both in the rougher sculpture of the scutum and propodeum.

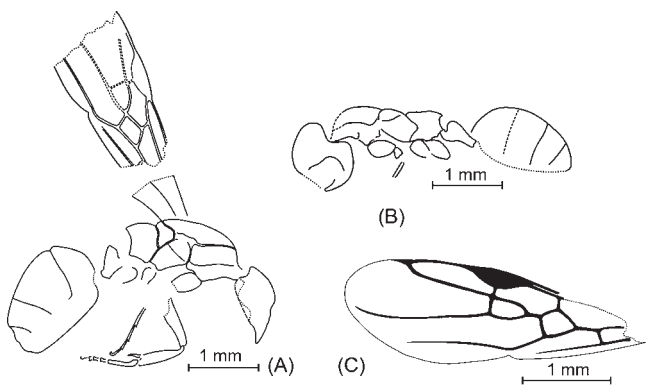
**Description.** Female. BL 4-4.5 mm. Head wide, with convex sides, rounded occipital corners and concave occipital



**Plate 18** (1) *Britaneuretus anglicus* (Cockerell) comb. nov., holotype of *Dolichoderus anglicus* Cockerell, USNM 61415. (2–3) *Dolichoderus vectensis* Donisthorpe: (2) holotype, NHMUK I.9198; (3) forewing, NHMUK I.9373. (4–6) *Emplastus britannicus* (Cockerell), comb. nov.: (4) holotype of *Dolichoderus britannicus* Cockerell, USNM 61413; (5) holotype of *Dolichoderus ovigerus* Cockerell USNM 61416; (6) holotype of *Emplastus emeryi* Donisthorpe, NHMUK In.24367. Scale bars = 1 mm.



**Text-figure 67** *Britaneuretus anglicus* (Cockerell), holotype of *Dolichoderus anglicus* Cockerell, USNM 6145.



**Text-figure 68** *Dolichoderus vectensis* Donisthorpe: (A) ♀, holotype, NHMUK I.9198; (B) ♀, NHMUK In.24934; (C) forewing, NHMUK I.9373.

margin. Mandibles well developed, triangular. Scutum feebly convex, not overhanging pronotum. Mesopleurae with distinct longitudinal furrow. Propodeum angular, with distinct carina separating propodeal dorsum and declivity; propodeal dorsum convex, propodeal declivity concave in side view. Legs rather short and thick. Petiole with rounded node, 1.5–1.8 times as long as high. At least scutum and propodeal sides with rather rough sculpture consisting of large isolated foveolae. Mesopleurae with no noticeable sculpture. Forewing: FWL 3.3–4 mm. 1RS and 2r–rs vertical to R. Cross-veins 2r–rs and rs–m coincide on RS, or rs–m shifted distally for about its width. Cell rm triangular, usually with distinct or (in holotype) short stem. Cell mcu of medium size. Icu = 1.55–2.0 mm; Icu = 1.4 mm.

**Measurements in mm.** NHMUK I.9198 (♀, holotype): BL = 4.4; AL = 1.6; HL ~ 0.8; FWL ~ 3.5. NHMUK In.24934 (♀): BL ~ 4; AL = 1.6; HL ~ 0.8; HW ~ 0.9; PtL = 0.52; PtH = 0.30. Other specimens (n = 3): FWL = 3.3–4.0; FWW = 0.8–1.1.

**Remarks.** Donisthorpe (1920) erroneously identified specimen I.9231 as *Leucotaphus gurnetensis*: all visible characters of this specimen including form of the propodeum are similar to the holotype of *D. vectensis*.

#### Morphogenus *Emplastus* Donisthorpe, 1920

1915 *Dolichoderus*; Cockerell, p. 483 (nec Lund, 1831).

1915 *Ponera*; Cockerell, p. 483 (nec Latreille, 1804).

1920 *Dolichoderus*; Donisthorpe, p. 87 (pars) (nec Lund, 1831).

1920 *Emplastus* Donisthorpe, p. 86.

**Type species.** *Emplastus emeryi* Donisthorpe, 1920 (= *Dolichoderus britannicus* Cockerell, 1915), by original designation.

**Other species.** *E. britannicus* (Cockerell, 1915), *E. hypolithus* (Cockerell, 1915), *E. gurnetensis* (Donisthorpe, 1920), and *E. kozlovi* sp. nov. Bembridge Marls, Isle of Wight, UK.

**Diagnosis.** Fossil ants not preserved well enough to fit orthotaxa, and with following combination of traits: size medium, head subrectangular with rounded occipital corners; mandibles triangular with denticulate masticatory margin; propodeum rounded in side view, petiole in side view with thick scale (females) or triangular (males). Forewing with cells 1+2r, 3r, rm and mcu closed; cell 3r with apex touching wing fore margin, 1RS vertical to R or slightly proclined. Icu > 1.45 mm, Icu > 1.2 mm. Gaster without constriction between first and second segments.

**Remarks.** Donisthorpe (1920) placed this genus in the subfamily Ponerinae and considered it similar to the modern *Myopias* Roger, 1861. In his opinion *Emplastus*, as well as *Myopias*, has mandibles without teeth. In fact, the holotype has the masticatory margin of mandibles hidden. Specimen NHMUK In.24461, similar to *E. emeryi* in the body proportions and other visible characters, has masticatory margin of mandibles with blunt teeth. The position of cross-vein cu-a in the type species (Icu = 1.7) differs clearly from that of Ponerinae. In contrast, all characters visible on the holotype of *Emplastus emeryi* are identical to those of *Dolichoderus britannicus* Cockerell, and so justify synonymy of the former under the latter species name.

Cockerell (1915) and Donisthorpe (1920) described several species now included in *Emplastus*, based on the *Dolichoderus*-like forewing venation (characteristic of several other dolichoderine genera). However, this character is combined with the propodeum rounded in side view and not angular or bispinate with concave hind contour, as in all living and fossil *Dolichoderus*.

The morphogenus *Emplastus* may embrace representatives of various dolichoderine genera, particularly those earlier included into *Iridomyrmex* (*Anonichomyrma* Donisthorpe, 1947, *Ctenobethylus* Brues, 1939, and *Iridomyrmex* Mayr, 1862). *Emplastus* differs from *Liometopum* Mayr, 1861 (Late Eocene–present) in the apex of cell 3r of the forewing touching the wing margin (distant from it in *Liometopum*). As mentioned above, *Emplastus* differs from females of *Dolichoderus* in the form of the propodeum.

*Emplastus britannicus* (Cockerell, 1915), comb. nov.

Plate 18, figs 4–6; Text-figs 65, 69A–G, 88C

1915 *Dolichoderus britannicus* Cockerell, pp 483–484, plate 65, figs. 6–7.

1915 *Dolichoderus ovigerus* Cockerell, p. 484, plate 65, fig. 9; syn. nov.

1920 *Dolichoderus britannicus* Cockerell; Donisthorpe, pp 87–88.

1920 *Dolichoderus ovigerus* Cockerell; Donisthorpe, p. 88.

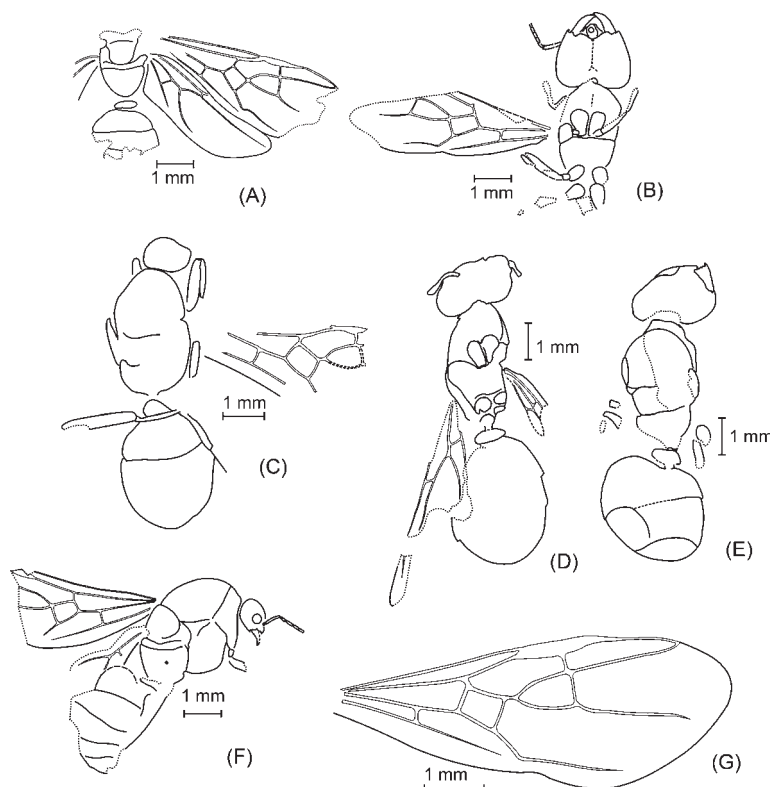
1920 *Emplastus emeryi* Donisthorpe, p. 86, plate V. fig. 5; syn. nov.

1978 *Dolichoderus britannicus* Cockerell; Burnham, p. 112.

1978 *Dolichoderus ovigerus* Cockerell; Burnham, p. 112.

1978 *Emplastus emeryi* Donisthorpe; Burnham, p. 109.

1992 *Emplastus emeryi* Donisthorpe; Carpenter, p. 492.



**Text-figure 69** *Emplastus britannicus* (Cockerell): (A) ♀, USNM 61413 (holotype of *Dolichoderus britannicus* Cockerell); (B) ♀, NHMUK In.24367 (holotype of *Emplastus emeryi* Donisthorpe); (C) ♀, USNM 61416 (holotype of *Dolichoderus ovigerus* Cockerell); (D) ♀, NHMUK I.8922; (E) ♀, NHMUK In.2461; (F) ♂, NHMUK I.10142; (G) forewing, NHMUK In.25083.

1995 *Dolichoderus britannicus* Cockerell; Bolton, p. 173.

1995 *Dolichoderus ovigerus* Cockerell; Bolton, p. 175.

1995 *Emplastus emeryi* Donisthorpe; Bolton, p. 188.

**Holotype.** USNM 61413, part and counterpart, Bembridge Marls, NW Isle of Wight, UK; ♀ (Plate 18, fig. 4; Text-fig. 69A).

**Paratype.** USNM 61414, Bembridge Marls, NW Isle of Wight, UK, Lacoe coll., ♀.

**Other material.** Holotype of *Dolichoderus ovigerus*, USNM 61416, Bembridge Marls, NW Isle of Wight, UK, coll. Lacoe, ♀ (Plate 18, fig. 5; Text-fig. 69C). Holotype of *Emplastus emeryi*, NHMUK In.24367, Bembridge Marls, NW Isle of Wight, UK, coll. R. W. Hooley, ♀ (Plate 18, fig. 6; Text-fig. 69B). Fifteen females, two males and 27 forewings in NHMUK collection and one forewing in CAMSM collection, including well preserved dorsal (NHMUK In.24461, Text-fig. 69E) and ventral (NHMUK I.8922, Text-fig. 69D) imprints of females, lateral imprint of male (NHMUK I.10142, Text-fig. 69F) and completely preserved imprint of forewing (NHMUK In.25083, Text-fig. 69G). Bembridge Marls, Isle of Wight, UK.

**Description.** Female. BL 6–8.5 mm, AL 2.5–3.5 mm, FWL 6–7 mm. Head wide (HW/HL = 1.2), as wide as alitrunk, broader behind than in front, with feebly convex sides, rounded occipital corners and concave occipital margin. Scape short, not reaching occipital margin. Mandibles massive, triangular, with few blunt teeth on masticatory margin. Alitrunk c. 1.8 times as long as wide. Scutum flat dorsally, rounded anteriorly, nearly as long as wide; not overhanging pronotum. Scutellum feebly convex, 1.7–1.9 times as wide as long. Propodeum evenly rounded in side view. Legs rather short and thick. Petiole with scale wide and thick, scale 2.5–3 times as

wide as thick. Gaster oval. Body integument matter preserved with chagrined surface.

Male. BL c.6 mm, AL c. 3 mm, FWL 5.5–6 mm. Head small, clearly narrower than alitrunk. Eyes oval, situated slightly before midlength of head sides. Scape short, not reaching occipital margin, about as long as two or three flagellomeres. Mandibles well developed, triangular. Pronotum narrow, along midline less than 0.25 as long as scutum. Scutum and scutellum feebly convex, scutum half as long as alitrunk. Propodeum rounded in side view. Legs rather long and thin. Petiole triangular in side view, with rounded top, higher than long. As preserved, surface sculpture similar to, but finer than, in female.

Forewing 1RS slightly proclined or, sometimes, vertical to R. Cross-veins 2r-rs and rs-m coincide on RS or, rarely, rs-m distal for cross-vein width. Cell rm triangular, not pedunculate or with very short stem, comparatively wide (1.6–2.1 times as long as wide). Cell mcu medium-sized, trapezoid or, rarely, rhomboid. Icu = 1.55–2.2, Icu<sub>a</sub> = 1.2–1.4. Hindwing with two longitudinal veins RS and Cu and well developed 1RS. Cross-vein rs-cu concave, longer than r-rs and 2Cu. Cross-vein cu—a more close to rs-cu than to wing base.

**Measurements in mm.** Types: USNM 61413 (♀, holotype of *Dolichoderus britannicus*): AW = 1.3; PtW = 0.66; FWL ~ 6. USNM 61414 (♀, paratype of *Dolichoderus britannicus*): BL = 5.7; AL = 2.7, FWL = 6.2. USNM 61416 (♀, holotype of *Dolichoderus ovigerus*): BL = 6.9; AL = 2.9, AW = 1.6. NHMUK In.24367 *Emplastus emeryi*): AL = 3.3; HL = 1.6; HW = 2.0. Other specimens: females: AL = 2.7–3.55 (n = 4); HW = 1.9–2.1 (n = 3); ScL = 1.4–1.5 (n = 3); SctW = 1.6–1.7 (n = 2); SctL = 0.65–0.7 (n = 3); PtW = 0.6–0.85; FWL = 6.0–7.05 (n = 3). Males: AL = 2.5–3.2 (n = 2); HL = 1.0 (n = 1); SL = 0.4 (n = 1); ED = 0.25 (n = 1);

FWL = 6.8 (n = 1); FWW = 1.9–2.1 (n = 2). Isolated forewings: FWL = 5.4–7.3 (n = 13); FWW = 1.6–2.1 (n = 11).

**Remarks.** Three specimens (NHMUK I.8922, NHMUK In.17067/NHMUK In.17074, NHMUK In.24461) were found in the NHMUK collection with the general appearance and available characters identical to those of the holotype of *Emplastus emeryi*. There are a significant number of incomplete fossils as well that form a complete transition between *Emplastus emeryi* Donisthorpe, *Dolichoderus britannicus* Cockerell and *D. ovigerus* Cockerell. Cockerell (1915) wrote that *D. ovigerus* had “Structure, including petiole, essentially as in *D. britannicus*, from which it may not be distinct” (Cockerell 1915, p. 484). The only indicated differences between these species are details in the forewing venation, particularly the pedunculate cell *rm* in *D. ovigerus*. However, this character is variable: for example, the specimen NHMUK In.25085 has cell *rm* long pedunculate in the right wing and with very short stem in the left one. This justifies synonymy of the three species resulting in the valid name *Emplastus britannicus* (Cockerell 1915).

There are ovoid bodies seen inside the abdomen of the holotype female of *D. ovigerus*. Cockerell described them as eggs. However, we observed similar structures inside the head and alitrunk of some other specimens, so we consider them to have a mineral origin.

Of the specimens listed by Donisthorpe (1920), we failed to find specimens NHMUK I.10257, NHMUK I.9347, NHMUK H.114, NHMUK H.262, NHMUK H.440, and NHMUK H.497, and found NHMUK I.9198 designated as the holotype of *Dolichoderus vectensis* Donisthorpe.

Donisthorpe (1920) erroneously identified specimen NHMUK I.8922 as *Oecophylla* sp. In fact, the specimen is very similar to the holotype of *Emplastus emeryi* and not to *Oecophylla*. He also identified a poorly preserved specimen NHMUK In.24410 (coll. R. W. Hooley) as *Oecophylla*, which probably belongs also to *E. britannicus*. Donisthorpe (1920, p. 84) cited W. M. Wheeler as having identified specimens NHMUK In.17074 and NHMUK In.24461 = NHMUK H.127 doubtfully as *Liometopum* queens.

Females of *Emplastus britannicus* have a general appearance similar to those of *Ctenobethylus* Brues, 1939 from Late Eocene ambers, and *Liometopum* Mayr, 1861 (Late Eocene–present). However, the key diagnostic characters of these genera (form of anterior margin of clypeus, presence or absence of ocelli, position of propodeal spiracles, etc.) are not visible on the studied specimens. Females of *E. britannicus* differ from *C. goepperti* in the wide petiole scale and from *Liometopum* in the wing venation (position of top of cell *rm*) and in the short antennal scape. Construction of the cranio-mandibular system of all of these females indicates that they built their nests most likely in rotten wood, similar to modern *Liometopum* and *Lasius* (subgenus *Dendrolasius* Ruzsky, 1912).

*Emplastus hypolithus* (Cockerell, 1915), comb. nov.  
Plate 19, figs 1–2; Text-fig. 70A–C.

- 1915 *Ponera hypolitha* Cockerell, p. 483, plate 64, figs. 3–4.  
1978 *Ponera hypolitha* Cockerell; Burnham, p. 109.  
1964 *Poneropsis hypolitha* (Cockerell); Taylor, 139.  
1995 *Poneropsis hypolitha* (Cockerell); Bolton, p. 363.

**Holotype.** USNM 61411, Bembridge Marls, NW Isle of Wight, UK, coll. Lacoe; forewing (Plate 19, fig. 1; Text-fig. 70A).

**Other material.** NHMUK In.17311, Bembridge Marls, NW Isle of Wight, UK, coll. E. J. A’Court Smith; male (Plate 19,

fig. 2; Text-fig. 70C) and 13 forewings (including NHMUK In.24368, Text-fig. 70B) in NHMUK collection. Bembridge Marls, Isle of Wight, UK.

**Diagnosis.** Differs from *E. britannicus*, the most common species of *Emplastus* in the Bembridge Marls, in form of the forewing cell *rm* and in position of 2r–rs.

**Description.** Male. BL 6.5 mm, AL 3 mm, FWL 5–6 mm. Head small, distinctively narrower than alitrunk. Eyes oval, situated slightly before midlength of head sides. Scape short, not reaching occipital margin, as long as three flagellomeres. Mandibles well developed, triangular. Pronotum comparatively wide, along midlength 0.33 times as long as scutum. Scutum feebly convex, half as long as alitrunk. Propodeum evenly rounded in side view. Legs rather long and thin. Petiole higher than long, in side view its anterior surface feebly concave, posterior surface straight, top rounded. Preserved remnants of body integuments chagrined.

**Forewing.** Cell *rm* triangular, not pedunculate, rarely quadrangular, comparatively narrow (2.5–2.8 times as long as wide). Cross-vein rs–m usually distal of 2r–rs, rarely coinciding. Cell *mcu* of medium size, trapezoid. *Icu* = 1.6–2.1, *Icu*a = 1.3–1.4.

**Measurements in mm.** USNM 61412 (holotype of *Ponera hypolitha*): FWL ~ 6. NHMUK In.17311 (♂): BL = 6.5; AL = 3.0; HL ~ 1.1; F3L = 1.7; FWL ~ 6. Other specimens: FWL = 5.0–5.8 (n = 7); FWW = 1.2–1.5 (n = 4).

**Remarks.** Cockerell (1915) described this species as *Ponera hypolitha* based on a fragment of the forewing (holotype, USNM 61412) and a poorly preserved hind wing (paratype, USNM 61412). Position of cross-vein cu–a in the holotype (*Icu* = 1.6) clearly indicates the fossil belongs to Dolichoderinae. A complete male (NHMUK In.17311) apparently conspecific with the holotype permits the assignment of the species to the morphogenus *Emplastus*. Paratype hind wing has no visible venation: structures considered by Cockerell to be veins are in fact wing folds. There are no features justifying attribution of the paratype to either *Ponera* or *Emplastus*; it is therefore considered to be Formicidae *incertae sedis*.

Donisthorpe erroneously identified specimens NHMUK In.17311, NHMUK In.24368 and NHMUK In.24373 as *Dolichoderus britannicus*, and NHMUK In.9347 as *D. ovigerus*, in spite of their forewing venation and particularly position of 2r–rs and form of *rm* which show no similarity to *E. britannicus*.

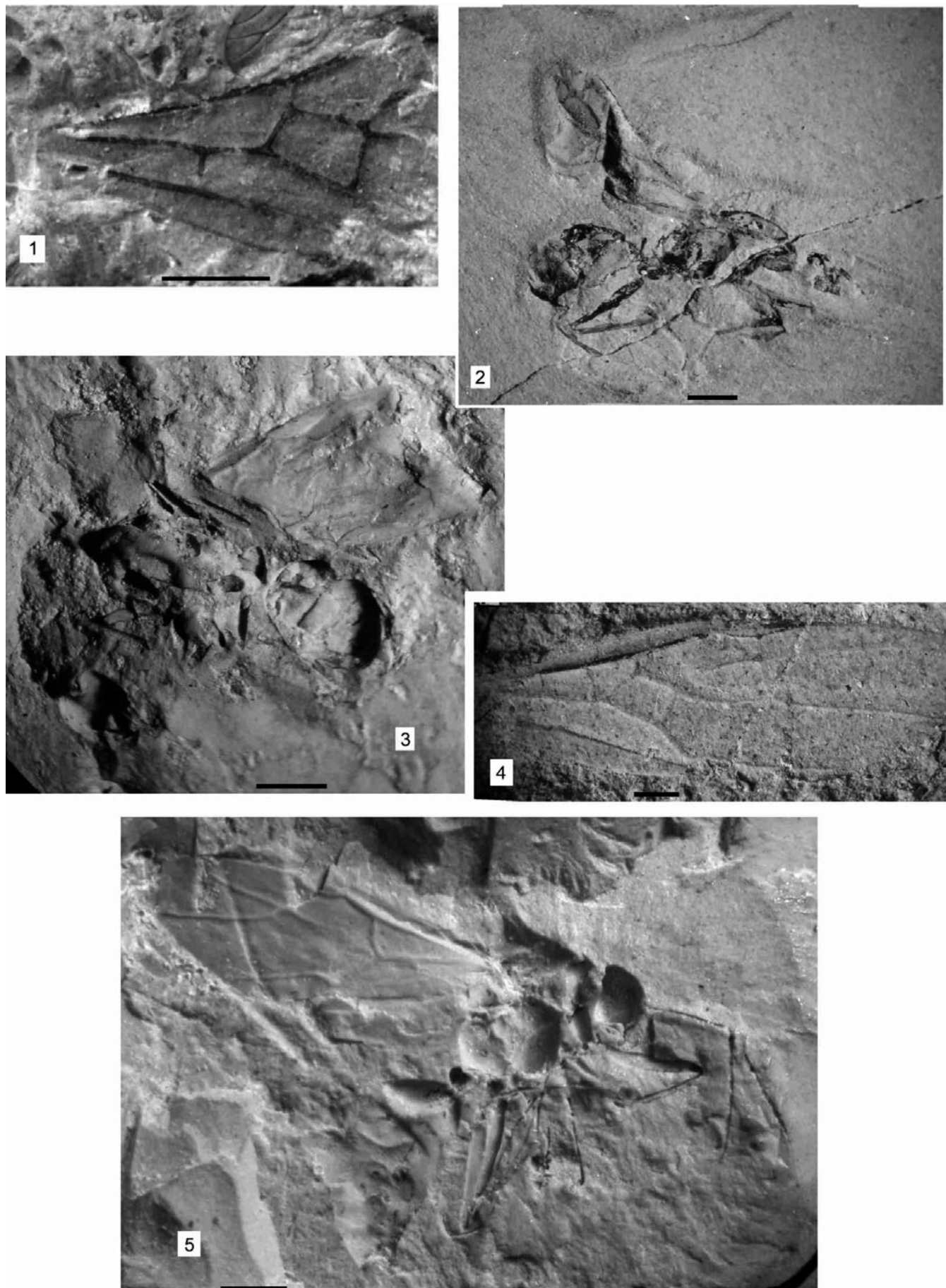
*Emplastus gurnetensis* (Donisthorpe, 1920), comb. nov.  
Plate 19, fig. 3; Text-fig. 71

- 1920 *Dolichoderus gurnetensis* Donisthorpe, p. 88, plate V, fig. 7.  
1978 *Dolichoderus gurnetensis* Donisthorpe; Burnham, p. 112.  
1995 *Dolichoderus gurnetensis* Donisthorpe; Bolton, p. 174.

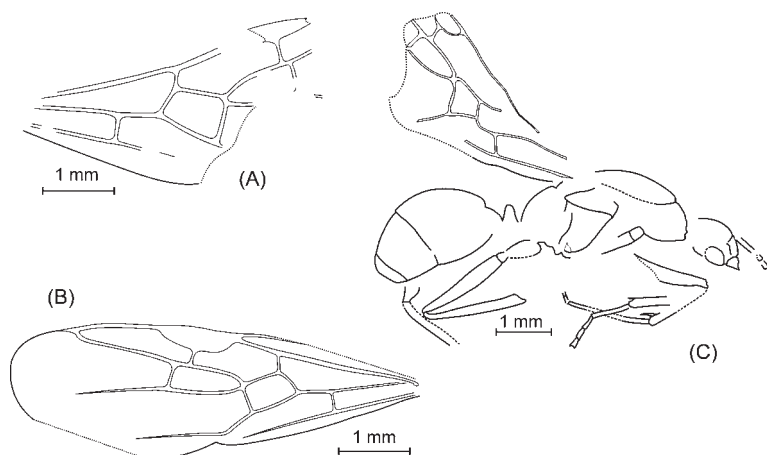
**Holotype.** NHMUK: I.9755, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie; ♀ (Plate 19, fig. 3; Text-fig. 71A).

**Other material:** Female, two males (including I.9510; Text-fig. 71B) and four forewings in NHMUK collection. Bembridge Marls, Isle of Wight, UK.

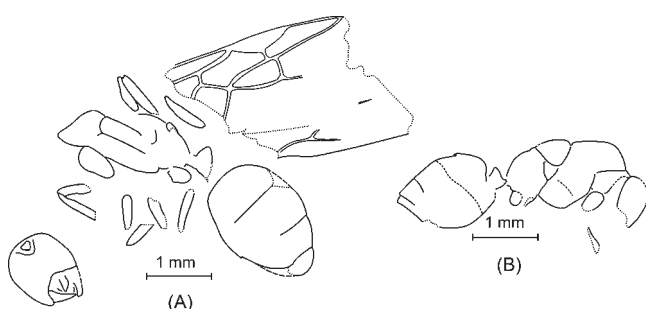
**Diagnosis.** The species occupies an intermediate position between *E. britannicus* and *E. hypolithus*. Forewing venation is similar to *E. hypolithus*, and males are similar in general appearance to *E. britannicus*, but differing in smaller size. The female holotype differs from females of *E. britannicus* in head proportions and in form of petiole. Body fossils of female *E. hypolithus* are unknown, but male *E. hypolithus* is 1.5 times as long as female *E. gurnetensis* and therefore these two cannot be conspecific.



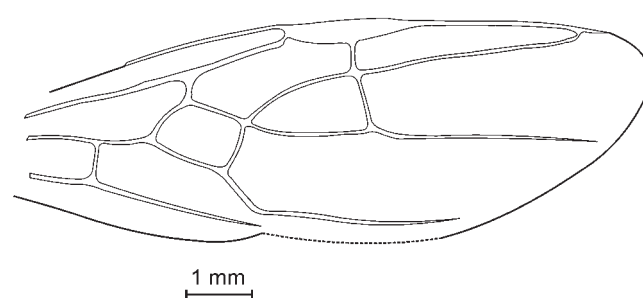
**Plate 19** (1–2) *Emplastus hypolithus* (Cockerell) comb. nov.: (1) holotype of *Ponera hypolitha* Cockerell, USNM 61411; (2) ♂, NHMUK In.17311. (3) *Emplastus gurnetensis* (Donisthorpe), comb. nov., holotype of *Dolichoderus gurnetensis* Donisthorpe, NHMUK I.9755. (4) *Emplastus kozlovi* Perfilieva, sp. nov., holotype, NHMUK PI II.2784. (5) *Camponotus cockerelli* (Donisthorpe) comb. nov., holotype of *Leucotaphus cockerelli* Donisthorpe, NHMUK I.8517. Scale bars = 1 mm.



**Text-figure 70** *Emplastus hypolithus* (Cockerell): (A) forewing, holotype of *Ponera hypolitha* Cockerell, USNM 61411; (B) forewing, NHMUK In.24368; (C) ♂, NHMUK In.17311.



**Text-figure 71** *Emplastus gurnetensis* (Donisthorpe): (A) ♀, holotype of *Dolichoderus gurnetensis* Donisthorpe, NHMUK I.9755; (B) ♂, NHMUK I.9510.



**Text-figure 72** *Emplastus kozlovi* Perfilieva, sp. nov., holotype, NHMUK PI II.2784.

**Description.** Female. BL *c.* 5.5 mm, AL *c.* 2 mm, FWL 5.5–6 mm. Head longer than wide (HL:HW = 1.07), about half as long as alitrunk, rectangular with slightly convex sides, rounded occipital corners and straight or feebly convex occipital margin. Mandibles massive, triangular. Propodeum evenly rounded in side view. Legs rather short and thick. Petiole with scale low, triangular in side view, about as long as high. Gaster oval. Body integument matter preserved with chagrined surface.

Male. BL *c.* 4 mm, AL 1.8–1.9 mm, FWL 3–4 mm. Head small. Alitrunk 2.5 times as long as head. Ocelli large (visible in NHMUK I.9082). Pronotum narrow, along midline less than 0.25 times as long as scutum. Scutum and scutellum convex in side view, scutum about half as long as alitrunk. Propodeum rounded in side view. Legs rather long and thin. Petiole in side view triangular, slightly higher than long. Body integument matter preserved with chagrined surface.

Forewing cell *rm* triangular, not pedunculate or with short stem 2.2–2.4 times as long as high. Cross-vein *rs–m* distal of 2*r–rs*. Cell *mcu* of medium size, rhomboid or trapezoid. *Icu* = 1.7–2.0, *Icu*<sub>a</sub> = 1.24–1.38. Hind wing with three longitudinal veins (*RS*, *M*, *Cu*). 1*M* and *rs–m* of roughly same length. Cross-vein *cu–a* closer to *rs–cu* than to wing base.

**Measurements in mm.** NHMUK I.9755 (♀ holotype): BL ~ 5.5; AL = 2.1; HL = 1.01; HW = 0.94; FWL ~ 5.7. NHMUK I.8723: FWL = 6.1; FWW = 1.7. Males: AL = 1.8–1.9 (*n* = 2); HL = 0.75–0.8 (*n* = 2); ScL = 0.9 (*n* = 1); ScTL = 0.54 (*n* = 1). Forewings: FWL = 4.0–6.3 (*n* = 2); FWW = 2.1 (*n* = 1).

**Remarks.** Donisthorpe identified specimen NHMUK I.9082 as *Leucotaphus gurnetensis* (Cockerell), but it is larger and differs in body proportions. He also identified forewing NHMUK In.24373 as *Dolichoderus britannicus* in spite of its venational difference (in position of *r–rs* and in form of *rm*).

*Emplastus kozlovi* Dlussky & Perfilieva, sp. nov.

Plate 19 fig. 4; Text-fig. 72

**Etymology.** In memory of Mikhail Kozlov, who collected the holotype.

**Holotype.** NHMUK PI II.2784 a, b, Bembridge Marls, Burnt Wood, Isle of Wight, UK, collected by Belokobylsky, Khalaim, and Kozlov 24.05.2005; forewing, (Plate 19, fig. 4; Text-fig. 72).

**Paratypes.** NHMUK In. 24372, NHMUK In.24936, NHMUK In.25112, NHMUK In.25141, forewings, Bembridge Marls, NW Isle of Wight, UK, coll. R. W. Hooley.

**Diagnosis.** Venationally similar to *E. britannicus*, but differs in larger size (7–9 mm in males and 11–12 mm in females; in contrast to, respectively, 5.5–6 mm and 6–7 mm in *E. britannicus*) and in relatively long cell *rm*.

**Description.** FWL 11–12 mm in females, 7.8–8.8 mm in males. 1*RS* and 2*r–rs* vertical to *R* and *RS*, respectively. Cell *rm* triangular, with no stem, rarely with very short stem, 2.2–2.5 times as long as wide. Cross-veins *rs–m* and 2*r–rs* coinciding. Cell *mcu* of medium size, trapezoid. *Icu* = 1.6–1.8, *Icu*<sub>a</sub> = 1.3–1.5.

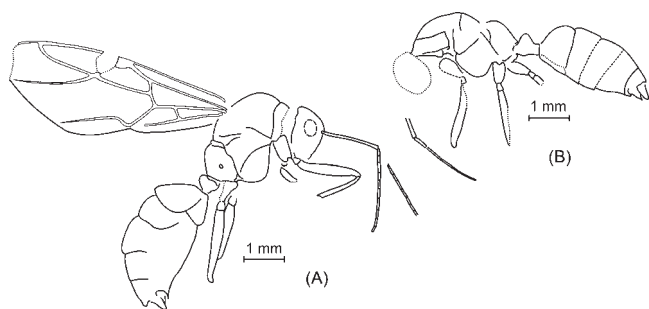
**Measurements in mm.** NHMUK: PI II.2784 (holotype): FWL = 11.3, FWW = 3.4; paratypes: FWL = 7.8–11.0, FWW = 1.8–3.0 (*n* = 4).

**Remarks.** Donisthorpe erroneously identified forewing NHMUK In.24372 as *E. britannicus*.

### 3.11.1.3. Subfamily Formicinae Latreille, 1809

Genus *Camponotus* Mayr, 1861

**Type species.** *Formica ligniperda* Latreille, 1802, by subsequent designation by Bingham (1903).



**Text-figure 73** *Camponotus cockerelli* (Donisthorpe): (A) ♂, holotype of *Leucotaphus cockerelli* Donisthorpe, NHMUK I.8517; (B) ♂, NHMUK In.17280.

**Other species.** In the modern fauna *Camponotus* totals c. 950 species distributed on all continents from forest–tundra in the north of Eurasia and Boreal America to Tierra del Fuego in the south. The majority of species frequent humid arboreal ecosystems, but occur also in deserts and savannas, where they build underground nests.

Bolton (1995) listed 23 fossil species of *Camponotus*. He missed nine species described by Zhang (1989) from the Miocene deposits of China (Shanwang). Zhang further reported (apparently erroneously) three extant species there: *C. festinus* F. Smith, 1857; *C. luteus* F. Smith, 1858; and *C. mitis* F. Smith, 1858. There are also indications of findings of unidentified *Camponotus* in the Oligocene of Camoin-les Bains in France (Timon-David 1944), in the Miocene of the Dominican amber (Wilson 1985) and in the rock fossils of Bakhioti in Georgia and Chon-Tuz in Kirghizia (Dlussky 1981a). One more species is described below.

As a result, c. 40 species of fossil *Camponotus* are currently known. However, only a subset of them can be attributed to the genus with reasonable confidence. *C. menzei* Mayr, 1868, described from the Baltic amber, as well as those from the Miocene of Georgia and Kirghizia are undoubtedly *Camponotus*. *C. fuscipennis* Carpenter, 1930 (latest Eocene, Florissant, USA), ? *shanwangensis* Hong, 1984, *C. longus* Zhang, 1989, *C. plenius* Zhang, 1989 and *C. ambon* Zhang, 1989 (Miocene, Shanwang, China) also belong to this genus, based on their figures and descriptions. Preservation and/or quality of descriptions of other species leave some doubts about their identification. Until re-examination of the types, these species should be either attributed to morphogenus *Camponotites* Dlussky, 1981 (wings and winged fossils), or considered Formicidae incertae sedis.

**Diagnosis** (for rock fossils). Size medium to large, head sub-rectangular with rounded occipital corners; male and female antennae geniculate, scape attached at some distance from posterior clypeal margin; mandibles triangular with denticulate masticatory margin; petiole with scale thick (females) or triangular in side view (males). Forewing with cells 1+2r, 3r; closed rm and m-cu lost; RS usually convex downwards within cell 3r, 5RS and 4M join basally (rs–m lost). 1M and 1RS form smooth, almost straight, distinctly reclined vein. Cross-vein 2r–rs distinctly reclined in respect to RS. Icu > 1.3. Hindwing with two longitudinal veins RS and Cu. Gaster without constriction between first and second segments.

*Camponotus cockerelli* (Donisthorpe, 1920), comb. nov.  
Plate 19, fig. 5; Text-fig. 73A, B

1920 *Leucotaphus cockerelli* Donisthorpe, pp 90–91, Plate V, fig. 10 (♂).

1978 *Leucotaphus cockerelli* Donisthorpe; Burnham, p. 111.

1995 *Leucotaphus cockerelli* Donisthorpe; Bolton, p. 246.

**Holotype.** NHMUK I.8517, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodi; ♂ (Plate 19, fig. 5; Text-fig. 73 A).

**Other material.** Four males (including well preserved lateral imprint) NHMUK In.17280; (Text-fig. 73B) and eight forewings in NHMUK collection, and two males in CAMSM collection. Bembridge Marls, Isle of Wight, UK.

**Description.** Male. BL 6–7.5 mm. Head about half as long as alitrunk. Eyes oval, situated above the midlength of head sides. Antennae geniculate, scape attached at some distance from clypeus; extending more than half its length beyond occipital margin. Scutum feebly convex in side view. Propodeum with dorsal surface short, convex, forming rounded blunt angle at junction with longer and almost straight declivous surface. Legs long and thin. Petiole triangular, a little longer than high in side view. Gaster cylindrical. Genital stipes short, narrow, rounded apically.

Forewing length of female 6.5–8 mm, male 5–6.5 mm. 2+3RS and 5RS both convex downward, not S-like. Distance between 1M and cu–a not longer than joint length 1M and 1RS. Icu = 1.3–1.6. Hindwing with two longitudinal veins RS and Cu. Section rs–cu about as long as 2M+Cu. Cross-vein cu–a near wing base.

**Measurements in mm.** Holotype ♂ NHMUK I.8517: BL = 7.5; AL = 2.6; HL ~ 1.3; SL = 1.4; ScL+ScTL = 1.9; FWL ~ 6; F3 = 1.8; NHMUK I.9225 (♂): AL = 1.9; NHMUK In.17280 (♂): BL = 7.5; AL = 2.7; ScL = 1.3; PtL = 0.6.

**Remarks.** Donisthorpe described *Leucotaphus cockerelli* based on two specimens: holotype ♂ NHMUK I.8517 and paratype ♀ NHMUK I.9028. The holotype is described as having the forewing cell m-cu closed, but figured with cross-vein m–cu closing this cell and represented by a dotted line. The holotype examined shows no trace of m–cu and so with no closed cell m-cu, which is not a wing anomaly. In case of aberrative loss of m–cu in a species with cu–a normally present, the wing retains a trace of the formerly closed cell m-cu in form of a distinct angulation at the junction of 1RS and 1M: the holotype has this composite vein smooth. Additionally, Bembridge *Leucotaphus* exhibits a more proximal position of cu–a in comparison with the present specimen. However, the strongest evidence of the species belonging to *Camponotus* is the scape attached to the head distinctly above the clypeus.

The paratype NHMUK I.9028 is actually Diptera indet. Instead we have found the specimen NHMUK I.9731 labelled erroneously as the holotype of *Leucotaphus cockerelli* which fits the description of the paratype, including measurements (body length 5.5 mm), and apparently represents the female paratype. We identify it as *Leucotaphus gurnetensis* (Cockerell).

Thirteen more specimens fit the diagnosis of *Camponotus cockerelli* and are found in the NHMUK collection. Of them, specimens NHMUK I.8692, NHMUK I.9225 and NHMUK In.17280 are misidentified by Donisthorpe (1920) as *Oecophylla* sp., and NHMUK In.17066 as *Leucotaphus gurnetensis*. The petiole form of NHMUK I.8692 and NHMUK I.9225 is not characteristic for *Oecophylla*; the posterior margin of cell rm, formed by RS of wing impression NHMUK In.17066 is convex (concave in *Oecophylla*); cell m-cu is closed in NHMUK In.17066. Specimen NHMUK In.17280, a male body with poorly preserved wings, is almost identical to the holotype. Other specimens are forewings of Camponotini of a size similar to that of the holotype.

Forewing length of the studied fossils varies from 5 mm to 8 mm. The range is wide enough to suppose it covers more than a single species and sex. In living *Camponotus*, male wings are narrower than those of the female (Perfilieva 2005). The present material shows Iw (ratio of forewing length to width) range as 2.9–4.3, indicating the presence of both sexes.

Wing morphometrics were analysed using standard methods (see above). Two comparatively uniform groups have been identified, apparently representing males and females. Moreover, the results indicate the presence of two species, one including males NHMUK I.8517 (holotype, FWL = 5.6 mm; Iw = 2.9) and NHMUK In.24794 (FWL = 6.5 mm), and female NHMUK In.24931 (FWL = 8.0 mm; Iw = 4.3); and another with males NHMUK I.8692 (FWL = 5.0 mm; Iw = 3.0) and NHMUK I.9359 (FWL = 6.0 mm) and females NHMUK In.24842 (FWL = 6.5; Iw = 3.7), NHMUK In.24881 (FWL = 7.5 mm; Iw = 3.6) and NHMUK In.25175 (FWL = 7.7 mm). Unfortunately, the material is too limited to justify formal description of a new species.

#### Morphogenus *Leucotaphus* Donisthorpe, 1920

**Type species.** *Leptothorax gurnetensis* Cockerell, 1915, by original designation.

**Other species.** *Leucotaphus gurnetensis* (Cockerell, 1915), *L. peraneus* Cockerell, 1927, *L. donisthorpei* sp. nov., Bembridge Limestone, Isle of Wight, UK.

**Diagnosis.** Fossil ants not well enough preserved to fit orthotaxa, and with following combination of traits: mandibles triangular; male antennae geniculate; scape articulate to the posterior margin of clypeus; female and male scape protruding beyond the occipital margin of the head; propodeum rounded or angulate in side view; petiole with high scale (♀) or triangular (♂) in side view; gaster without constriction between the first and second gastral (III and IV abdominal) segments. Forewing with cells 1+2r, 3r and mcu and not rm closed. 2-3RS convex, rarely feebly arching, 5RS convex. Cell 3r with apex touching wing margin. Sections 5RS and 4M with joint start (rs-m lost). Cell mcu trapezoid (RS+M appreciably shorter than 1Cu), small (in terms of area, less than half cell 1+2r). Icu > 1.5, Icu a > 1.2. Hindwing with two longitudinal veins RS and Cu, 1RS lost (2RS and rs-cu join at R). Cross-vein cu-a near wing base.

**Remarks.** Many genera of formicine ants from the tribes Formicini and Lasiini (in particular, *Formica* Linnaeus, 1758 and *Lasius* Fabricius, 1804) fit the diagnosis of *Leucotaphus*. Undoubtedly many fossil species described as *Formica* and *Lasius* from poorly preserved impressions deserve transfer into this morphogenus. *Leucotaphus* would range at least from the Paleocene (undescribed wings from Tadushi Formation in the Russian Far East) up to the Miocene (*Lasius vetulus* Dlussky, 1981).

*Leucotaphus gurnetensis* (Cockerell, 1915)  
Plate 20, figs 1-3; Text-figs 74A-E, 88B

1915 *Leptothorax gurnetensis* Cockerell, p. 485, plate 65, figs. 4, 5 (wing).

1920 *Leucotaphus gurnetensis* (Cockerell): Donisthorpe, p. 89, Plate V, fig. 8,9 (male).

1978 *Leptothorax gurnetensis* Cockerell: Burnham, p. 111.

1992 *Leptothorax gurnetensis* Cockerell: Carpenter, p. 492.

1995 *Leucotaphus gurnetensis* (Cockerell): Bolton, p. 246.

**Holotype.** USNM 61417, Bembridge Marls, NW Isle of Wight, UK, coll. Lacoe (holotype of *Leptothorax gurnetensis* Cockerell); ♂ (Plate 20, fig. 1; Text-fig. 74A).

**Other material** USNM 61418 (♂, Lacoe Coll., designated by Cockerell as *Leptothorax gurnetensis* var. a), 100 males, ten females, and 36 isolated forewings (32 ♂ and four ♀) in the NHMUK collection, and four ♂ forewings in the CAMSM collection, including: NHMUK I.9731 (♀, which is the para-

type of *Leucotaphus cockerelli* Donisthorpe, 1920); NHMUK I.9744 (♂, designated by Donisthorpe (1920) as ergatotype of *Leucotaphus gurnetensis*, Text-fig. 74E); NHMUK I.9756 (♂, designated by Donisthorpe (1920) as plesiotype of *Leucotaphus gurnetensis*); a well preserved dorsal imprint of female NHMUK I.8733 (Plate 20, fig. 2; Text-fig. 74B); and lateral imprints of males NHMUK I.10040 (Text-fig. 74C) and NHMUK I.9796 (Plate 20, fig. 3; Text-fig. 74D). Four males in CAMSM collection.

**Description.** Male. BL 2.2-3.8 mm, AL 1.0-1.7 mm, FWL 2.2-3.7 mm. Head subtrapezoid, narrower in front than behind, longer than wide, occipital corners rounded, occipital margin feebly convex. Eyes oval, situated slightly before mid-length of head sides; maximum eye diameter 2.6-3.1 times less than head length. Scape slightly protrudes beyond the occipital margin of the head; scape length a little less than head length. Mandibles well developed, triangular; mandibular teeth not visible in any specimen. Alitrunk 1.2-1.4 times as wide as head, twice as long as wide and 1.3-1.5 times as long as high. In side view scutum feebly convex, propodeum rounded or with the poorly expressed angle, in which case propodeal dorsum much shorter than declivity. Legs rather thin and long. Petiole in side view triangular, with rounded top, higher than long. Gaster oviform, narrowed behind. Genitals slightly exposed, with stipes short, narrow, rounded at the top.

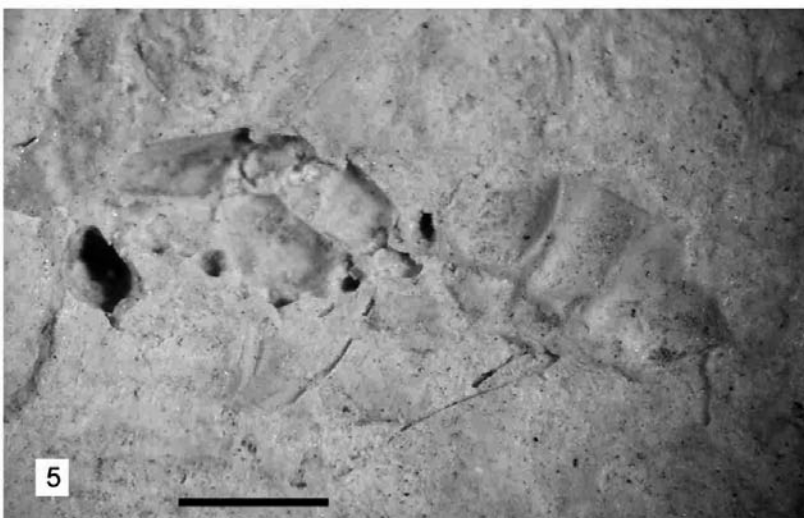
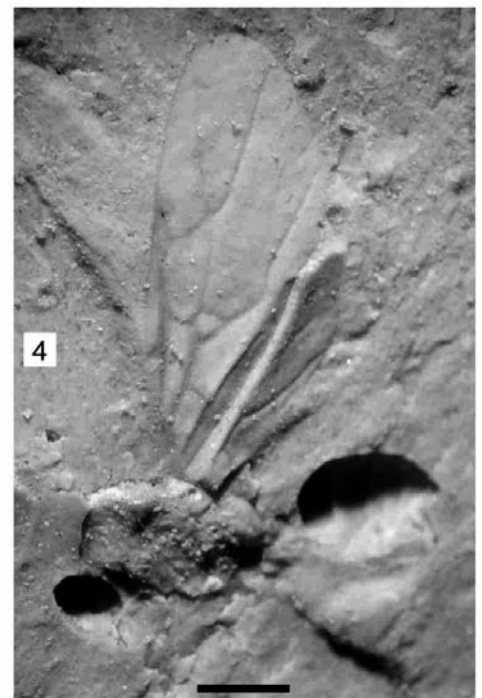
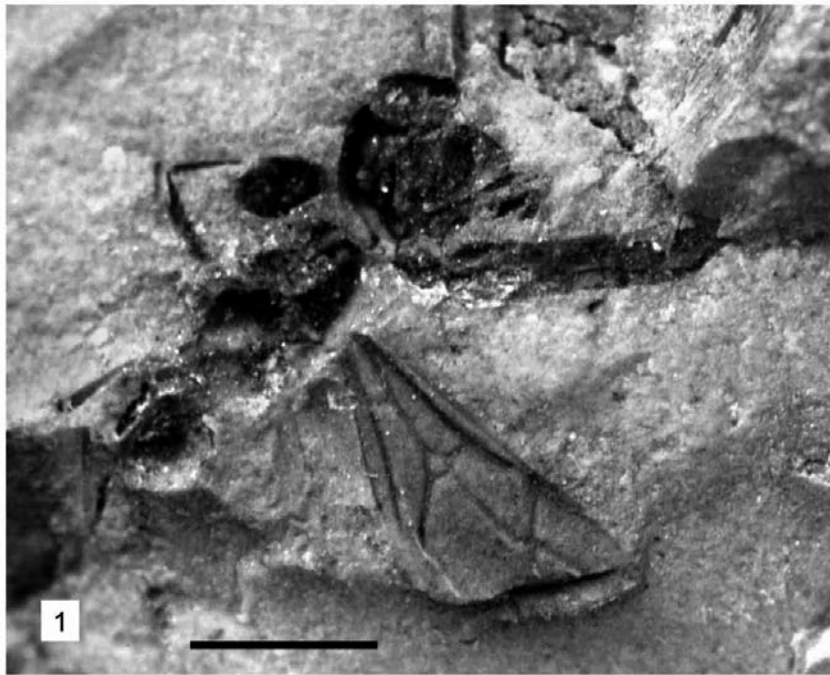
Female. BL 5.0-6.5 mm, AL 2.2-3.2 mm, FWL 5-5.5 mm. Head narrower than alitrunk. Alitrunk 1.8-1.9 times as long as wide, 1.3-1.5 as long as high. In side view scutum rounded in front, flat behind. Propodeal dorsum and declivity forming rounded blunt angle at junction. Petiole 2.2-2.4 times as high as long, with scale high and wide, 2.5-3 times as wide as thick. Gaster oval.

Forewing length c. 5 mm (♀), 2.5-3.7 mm (♂). Icu 2.0-3.6 (mean  $2.51 \pm 0.09$ ,  $n = 17$ ), varying widely because of variable size of cell mcu which is always small (in terms of area far less than half cell 1+2r). Icu a = 1.2-2.2. Hind wing with cross-vein rs-cu almost straight, comparatively short (clearly shorter than 2M+Cu).

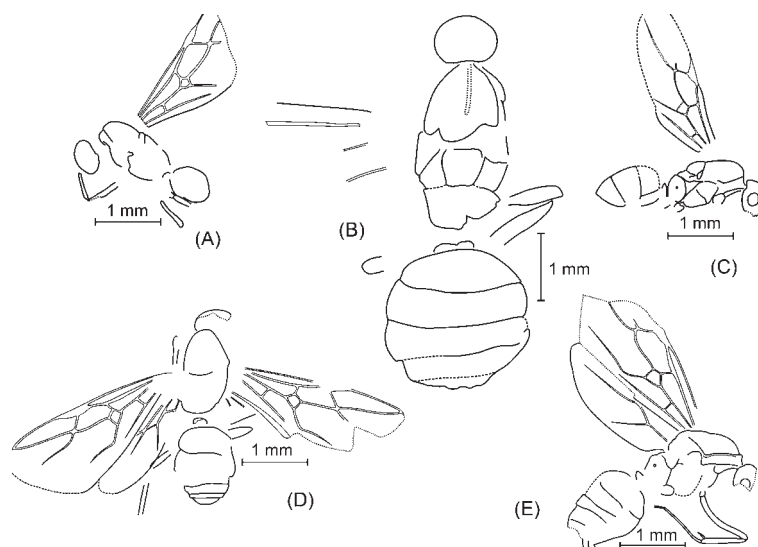
**Measurements in mm.** USNM 61417 (♂, holotype): BL = 3.05; AL = 1.2; FWL ~ 2.6. Other specimens: Males: BL = 2.2-3.8 (average = 2.92, std = 0.330,  $n = 40$ ); AL = 1.0-1.7 (average = 1.35, std = 0.166,  $n = 40$ ); HL = 0.44-0.69 (average = 0.54, std 0.09,  $n = 8$ ); HW = 0.54 ( $n = 1$ ); SL = 0.54 ( $n = 1$ ); ED = 0.18-0.24 ( $n = 4$ ); ScL = 0.57-0.87 (average = 0.67, std = 0.10,  $n = 7$ ); SctL = 0.27-0.45 ( $n = 4$ ); FWL = 2.2-3.7 (average = 3.05, std = 0.356,  $n = 55$ ). Females: AL = 2.4-3.2 (average = 2.68, std = 0.34,  $n = 7$ ); AW = 1.3-1.6 ( $n = 3$ ); ScL = 1.1 ( $n = 1$ ); ScW = 1.3 ( $n = 1$ ); PtW = 0.58-0.65 ( $n = 5$ ).

**Remarks.** When describing the holotype forewing (USNM 61417), Cockerell (1915) referred to the very small size of cell mcu as a key diagnostic character. The second studied specimen of this species (USNM 61418), with large mcu, he considered to be probably an individual variation. We found the holotype Icu = 3.1, Icu a = 1.7, and the specimen USNM 61418 Icu = 2.1, Icu a = 1.6. Cell mcu is really variable in size in *L. gurnetensis*: sometimes it is larger than even in var.  $\alpha$ , sometimes it is less than in the holotype, and the majority of studied specimens it is of intermediate size.

Male *L. gurnetensis* is quite variable in size, with the forewing length distribution being bimodal: based on 55 measured wings, we found maximums at 2.8-2.9 mm and 3.2-3.3 mm. It can be excluded that the material represents a mixture of two different species. No discrete differences were revealed to discriminate the composing species. The current taxonomic concept of *L. gurnetensis* is best left until more knowledge is obtained.



**Plate 20** (1–3) *Leucotaphus gurnetensis* (Cockerell): (1) holotype of *Leptothorax gurnetensis* Cockerell, USNM 61417; (2) ♀, NHMUK I.8733; (3) ♂, NHMUK I.9796. (4–5) *Leucotaphus donisthorpei* Dlussky, sp. nov.: (4) holotype, NHMUK I.8765; (5) paratype, NHMUK I.10097. Scale bars = 1 mm.



**Text-figure 74** *Leucotaphus gurnetensis* (Cockerell): (A) ♂, holotype of *Leptothorax gurnetensis* Cockerell, USNM 61417; (B) ♀, NHMUK I.8733; (C) ♂, NHMUK I.10040; (D) ♂, NHMUK I.9796; (E) ♂, NHMUK I.9744.

Donisthorpe (1920) listed 116 specimens identified as *Leucotaphus gurnetensis*. We have found two more specimens labelled and not listed (NHMUK I.9343, NHMUK I.9827), but have failed to locate 24 listed specimens: NHMUK I.7286, NHMUK I.8764, NHMUK I.8889, NHMUK I.8927, NHMUK I.9142, NHMUK I.9184, NHMUK I.9246, NHMUK I.9266, NHMUK I.9351, NHMUK I.9356, NHMUK I.9386, NHMUK I.9526, NHMUK I.9593, NHMUK I.9718, NHMUK I.9688, NHMUK I.10005, NHMUK I.10018, NHMUK I.10120, NHMUK I.1026, NHMUK In.17202, NHMUK In.17203, NHMUK In.17298, NHMUK In.19602 (= H.189). As a result, we were able to study 94 specimens identified by Donisthorpe. Most of them were really males of *Leucotaphus gurnetensis*, but some were found either to be of poor preservation, permitting only identification as Formicidae incertae sedis (NHMUK I.8994, NHMUK I.9281, NHMUK I.9677, NHMUK I.9958, NHMUK I.10265, NHMUK In.24376), or to be misidentified and belonging in fact to *Dolichoderus vectensis* (NHMUK I.9231), *Emplastus gurnetensis* sp. (NHMUK I.9082), *Camponotus cockerelli* (NHMUK In.17066), *Leucotaphus donisthorpei* (NHMUK I.8678, NHMUK I.8737, NHMUK I.8765, NHMUK I.10097), *Oecophylla atavina* (NHMUK In.17213), *Ponerites crawleyi* (NHMUK I.8539, NHMUK I.8684, NHMUK I.8759, NHMUK I.9406, NHMUK I.9774), *Ponerites hooleyi* (NHMUK I.9602, NHMUK I.9869), *Ponerites antropovi* (NHMUK I.9996), *Ponerites* sp. (NHMUK I.10123) and *Taphopone microptera* (NHMUK I.9763, NHMUK In.24378).

Wheeler identified specimen NHMUK In.24380 (=H.334) as a worker of *Camponotus* sp. (Donisthorpe 1920, p. 84). Actually, the constitution of its alitrunk, well developed and with scutum and scutellum separated, clearly specifies the fossil as a wingless female and not a worker. Specimen NHMUK I.8731 is designated by Donisthorpe as the paratype of *Leucotaphus cockerelli*. As shown above, the holotype of this species is actually a male of *Camponotus*. The specimen NHMUK I.8731 has the petiolar scale high and narrow, attesting it as a female, with size and body proportions fitting *Leucotaphus gurnetensis*.

Re-describing the male of *Leucotaphus gurnetensis*, Donisthorpe (1920, p. 90) noted that "there are several larger specimens, which may be the females of this species, this sex being, on that view, larger than the male, as in some of the subgenera

of *Acanthomyops*; some are dealated, others winged. One specimen (NHMUK I.10097) is very perfect, showing well the segments of the thorax and gaster." Actually, this specimen has clearly visible male genitals. It is described below as a paratype of *Leucotaphus donisthorpei*.

Donisthorpe also described a cocoon and workers of *Leucotaphus gurnetensis*. Concerning workers he wrote (p. 89): "I have seen some eight workers, nearly all of them being on the same piece of rock as winged specimens. The head is small, the scale distinct, and some parts of the legs are present, but the general outline is not very clear. The length is 2–2.7 mm". We have studied four of eight samples listed by him and found no workers. Sample NHMUK I.10248 includes poorly preserved impression of an insect other than an ant, samples NHMUK I.8722 and NHMUK I.9744 include only impressions of male *Leucotaphus gurnetensis*, and sample NHMUK In.17250 is poorly preserved fragments of *Oecophylla* sp. In general, ant workers and the more so their cocoons have very low chance of being preserved as rock fossils.

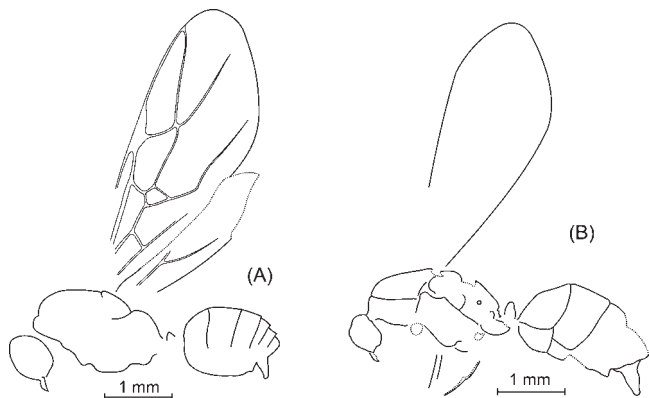
*Leucotaphus gurnetensis* is the third most abundant species (after two *Oecophylla* species) in the Bembridge Marls ant assemblage. It comprises 25% of all ant fossils identified to the species level. This ant is similar to the Recent *Lasius* Fabricius, 1804 in general appearance, and perhaps really belongs to this genus. However the level of preservation does not allow any sight of key characters that differ *Lasius* from other genera of Formicinae. It is also very similar to fossil ants described as *Lasius* from other deposits: *L. schiefferdeckeri* Mayr, 1868 (Late Eocene Baltic amber), *L. peritulus* (Cockerell, 1927) (latest Eocene, Florissant, USA), *L. epicentrus* Théobald, 1937 (Oligocene Aix-en-Provence, France), *L. redtenbacheri* (Heer, 1849) and others (Miocene, Radoboj, Croatia), *L. vetulus* Dlussky, 1981 (Vishnevaja Balka, Russia) and others. Like *L. gurnetensis* these ants are numerous in their taphocenoses (11% in Baltic amber, 25% in Florissant, 33% in Radoboj, 34% in Vishnevaja Balka).

*Leucotaphus donisthorpei* Dlussky & Perfilieva, sp. nov.

Plate 20, figs 4–5; Text-fig. 75A, B

**Etymology.** In honour of Horace St. J. K. Donisthorpe.

**Holotype.** NHMUK I.8765, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie, ♂ (Plate 20, fig. 4; Text-fig. 75 A).



**Text-figure 75** *Leucotaphus donisthorpei* sp. nov.: (A) ♂, holotype, NHMUK I.8765; (B) ♂, paratype, NHMUK I.10097.

**Paratypes.** NHMUK: I.8678, I.8750, I.8948, I.10097, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie; ♂♂ (Plate 20, fig. 5; Text-fig. 75 B); NHMUK In 25124, Bembridge Marls, NW Isle of Wight, UK, coll. R. W. Hooley; ♀ forewing.

**Other material.** Three males, three male forewings, and two female forewings in NHMUK collection. Bembridge Marls, Isle of Wight, UK.

**Diagnosis.** Differs from the type species in larger wing size.

**Description.** Male. BL 4.0–4.5 mm, AL 1.4–2.0 mm, FWL 3.8–4.5 mm. Head small, narrower than alitrunk. Scape longer than head, distinctly surpassing occipital margin. Mandibles well developed, triangular. Alitrunk 1.4–1.6 times as long as high. Scutum in side view rounded in front and flat behind. Propodeum in side view rounded. Petiole higher than long, with thick scale, narrowed upward, with rounded top. Gaster oviform. Genitals strongly exposed, with stipes long, triangular, rounded at apex.

Forewing length ♀ c. 7.5 mm, ♂ 3.8–4.5 mm, Icu = 1.9–2.3, Icu<sub>a</sub> = 1.35–1.55. Fore and hind wing venation as in *L. gurnetensis*.

**Measurements in mm.** NHMUK I.8765 (♂, holotype): BL = 4.0; AL = 1.9; HL = 0.75; FWL = 4.5. Other specimens (♂♂); AL = 1.5–2.8 (n = 6); HL = 1.1 (n = 1); FWL = 3.8–4.3 (n = 4).

**Remarks.** There are two forewings (NHMUK In.24920 and NHMUK In.25124) with forewing venation characteristic for *Leucotaphus*, but larger than female forewings of *L. gurnetensis*. We assume that they belong to females of *L. donisthorpei*. Forewing fragment NHMUK I.9549 may also be a male of that species.

Donisthorpe erroneously identified specimens NHMUK I.8737, NHMUK I.8765, NHMUK I.8678, and NHMUK I.10097 as *Leucotaphus gurnetensis*, and NHMUK I.8673 as *Oecophylla* sp.

#### Genus *Oecophylla* F. Smith, 1860

**Type species.** *Formica virescens* Fabricius, 1775, a junior synonym of *Oecophylla smaragdina* (Fabricius, 1775), by subsequent designation of Bingham, 1903.

**Other species.** There are two extant species of this genus: *Oecophylla smaragdina* (Fabricius 1775) in the Oriental and Indo-Australian Regions, and *O. longinoda* (Latreille 1802) in the Afrotropical Region. Fifteen fossil species are described, the most ancient being the Middle Eocene: *O. longiceps* Dlussky in Dlussky *et al.*, 2008 from Grube Messel, Germany, and *O. eckfeldiana* Dlussky, in Dlussky *et al.* 2008 from Eckfeld,

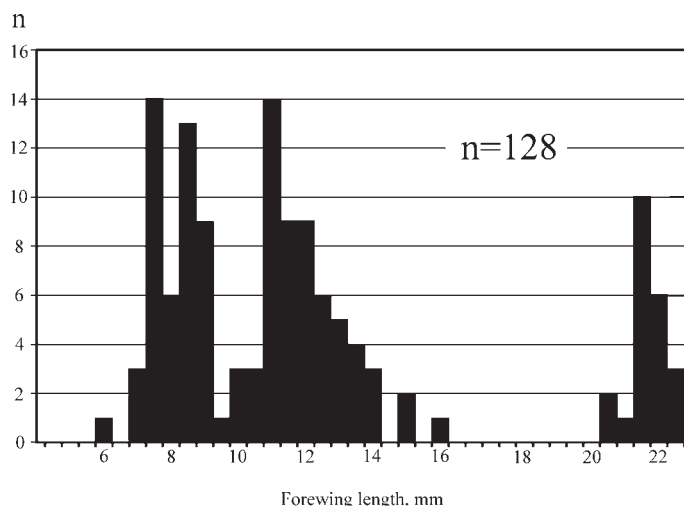
Germany (Dlussky *et al.*, 2008); and the Late Eocene: *O. brischkei* Mayr, 1868 and *O. crassinoda* Wheeler, 1922 (= *O. brevinodis* Wheeler, 1915) from the Baltic amber, *O. praeclara* Foerster (Brunstadt, Germany), *O. atavina* Cockerell, 1915, *O. perdita* Cockerell, 1915 and *O. megarche* Cockerell, 1915, from Bembridge, Isle of Wight, UK and *O. bartoniana* Cockerell, 1920b from the Eocene Bagshot Beds, Bournemouth, UK. Two species are described from European Oligocene deposits: *O. superba* Théobald, 1937 (Haut-Rhin, France) and *O. sicula* Emery, 1905 (Sicilian amber). Also, four species are known from Miocene deposits: *O. obesa* (Heer 1849), *O. radobojana* (Heer 1849) (Radoboj, Croatia), *O. leakeyi* Wilson & Taylor, 1964 (Victoria, Kenya) and *O. xiejiaheensis* (Hong, 1984) (Shanwang, China). However, the real number of fossil species is less. *O. xiejiaheensis* is transferred to the morphogenus *Campopotites* Dlussky, 1981 (Dlussky *et al.* 2008), and *O. perdita* is synonymised herein under *O. atavina*. *O. bartoniana* and *O. praeclara* were described from detached forewings, so they may be conspecific with other species known from complete fossils.

**Diagnosis** (for rock fossils). Size large, head subrectangular with rounded occipital corners; antennae geniculate in both sexes; scape attached at some distance from clypeus; mandibles large, triangular, with sharp teeth on the masticatory margin; petiole elongate, without scale, or nodiform (only at females); gaster without constriction between first and second gastral (III and IV abdominal) segments; forewing with closed cells 1+2r, 3r, with no closed rm and mcu; posterior margin of cell 3r (formed by RS) always concave.

**Remarks.** Both modern species of the genus (“weaver ants”) live in the crown of trees of tropical forests and construct nests from leaves, fastened by silk secreted by the larvae. Morphology of *Oecophylla* agrees with their mode of life. Long legs and antennae allow them to step or jump in the crown from one leaf to another. The original petiole construction is connected with their ability to lift the gaster upwards, providing large maneuverability during leaps (Dlussky 1981b). Large flat mandibles with sharp teeth are adapted to cutting leaves for construction of the nest. The morphological resemblance of extinct and extant species allows confidence to assert that their modes of life also were similar. It proves to be true also from the fossilised nest of *O. leakeyi* in the Miocene deposits of Kenya (Wilson & Taylor 1964). However, their tropical preference should not be considered as strict, because of their discovery in the European Miocene which had a subtropical to warm temperate rather than a tropical climate.

*Oecophylla* are the most numerous ants in Bembridge Marls assemblage comprising 69.8% of all ant fossils identified to the generic level. However, this proportion is obviously overestimated. Firstly, large fossils such as *Oecophylla*, the largest ant in the assemblage, are better preserved and easier to collect. Secondly, the original shape and structure of these ants, jointly with their large size, helps identification, even from small fragments that are usually not noticed in the case of other ants. When comparing only more complete fossils identifiable to the species level, the proportion of *Oecophylla* falls to 51.8%, and yet remains higher than that of any other ant (*Leucotaphus gurnetensis* 24.9%; *Emplastus britannicus* 10%).

Cockerell (1915) described three *Oecophylla* species from the Bembridge Marls: *O. atavina*, *O. perdita* and *O. megarche*. He distinguished them by using small details of forewing venation, which lie well within the range of intraspecific variability. Donisthorpe (1920) distinguished only *O. megarche* based on the very large size, and wrote about other species: “They are all smaller than specimens of *O. megarche*. A chart of the measurements allows that they vary in every possible



**Text-figure 76** Frequency distributions of forewing length classes among *Oecophylla*.

way – in size, size of wings, and all other measurements. Nor does it appear to me that any good purpose would be served by creating a number of new species, which would probably embrace or overlap Cockerell's *O. atavina* and *O. perdita*" (p. 92).

Initially we supposed that all *Oecophylla* fossils belong to one species. However, when we made a diagram of frequency distribution of forewing size, we found it three-modal (Text-fig. 76). All specimens can be divided into three distinct clusters: the small ones, with FWL 6.0–9.4 mm, mode 7.5 mm; the medium ones (FWL 9.5–16 mm, mode 11 mm); and the large ones (FW 21–23 mm). The ranges of variation are, respectively, 3.5 mm (47% of modal wing length), 6.5 mm (59%), and 2.0 mm (9%). The studied samples of living Formicinae (Perfilieva 2005) show variation ranges of forewing length as some 17% and not above 40% (in case of polymorphic females). This indicates that the present middle and, less certainly, smaller clusters are probably of a mixed nature.

Small specimens with bodies preserved are mostly males, including ♂ holotype of *O. atavina*, FWL = 7.6 mm and ♂ paratype of *O. perdita*, FWL = 6.8 mm; except the only female NHMUK In.24400 with FWL = 8.5 mm. Large fossils, including the holotype of *O. megarche* with FWL = 23 mm, are mainly represented by isolated wings; except the paratype of *O. megarche* USNM 61425, which shows additionally a head and a fragment of alitrunk. However there is no doubt that all of them belong to females. In contrast, specimens of the medium group with bodies preserved represent both males and females. Holotype (♀, FWL = 12.7 mm) and one paratype (♀ FWL = 11.8 mm) of *O. perdita* belong to this group. Standard morphometric analysis (see above) of wings 9–17 mm long for characters discriminating male and female ant wings (Perfilieva 2005) reveals two distinct clusters. This makes it possible to appreciate two species of *Oecophylla* present in the Bembridge Marls ant assemblage, with the small species (AL 2.0–2.8 mm; FWL 6.0–9.5 mm; ♀ AL 3.5–4.5 mm; FWL 8.5–13.7 mm) being *Oecophylla atavina* Cockerell, 1915 (= *O. perdita* Cockerell, 1915), and the large species (♂ AL 4.5–5.5 mm; FWL 10–15.9 mm; ♀ FWL 20.7–23.0 mm) *Oecophylla megarche* Cockerell, 1915.

We have identified 195 specimens as *O. atavina*, and 122 specimens as *O. megarche*. 366 incomplete fossils (fragments of bodies and wings) in the NHMUK collection and 37 in the CAMSM collection are left identified as *Oecophylla* sp.

*Oecophylla atavina* Cockerell, 1915

Plate 21, figs 1–5; Text-figs 77, 88A

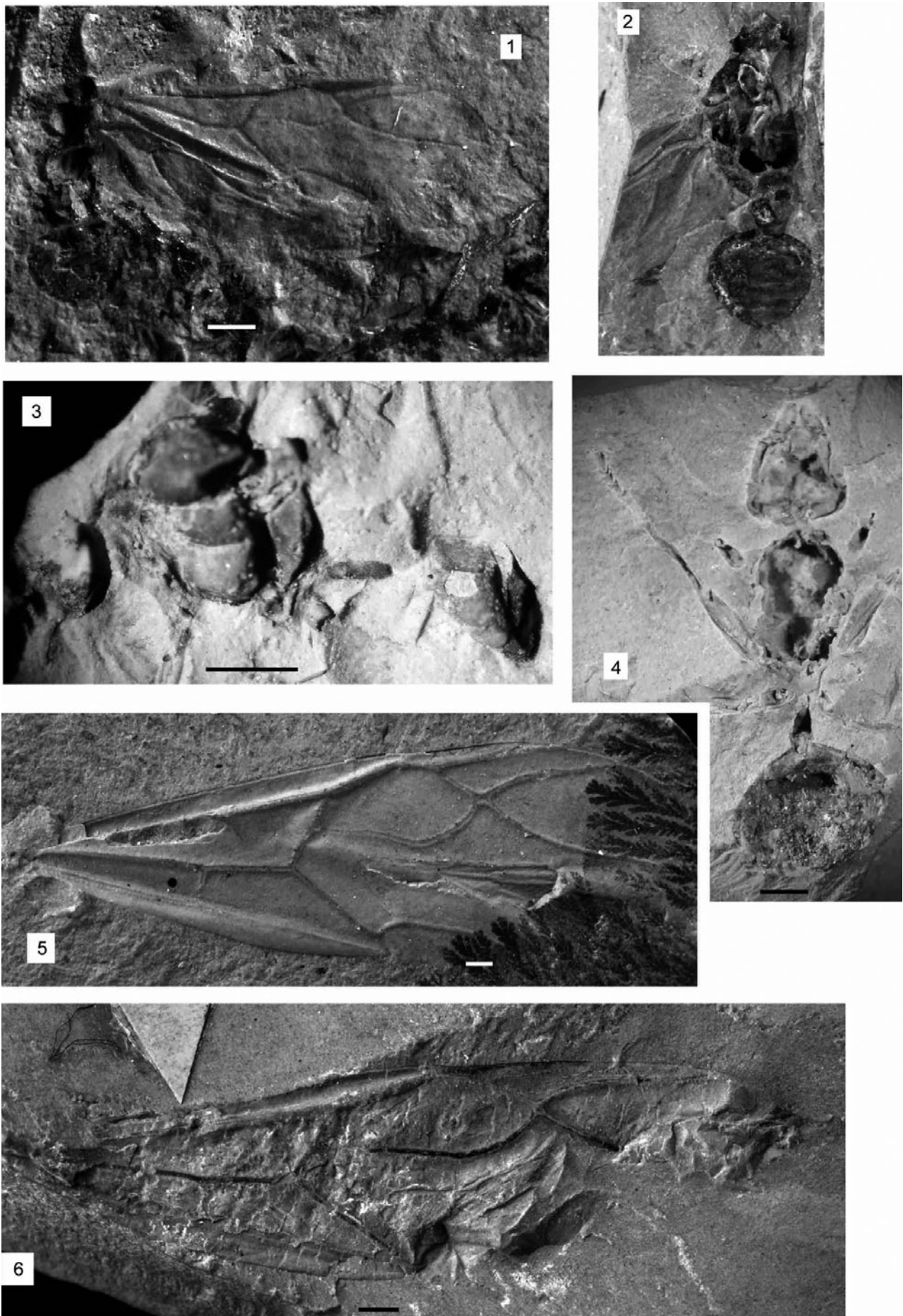
- 1915 *Oecophylla atavina* Cockerell, p. 485, plate 64, fig. 7 (♂).  
 1915 *Oecophylla perdita* Cockerell, pp 485–486, plate 64, figs. 5–6 (♀♀) syn. nov.  
 1920 *Oecophylla megarche* Cockerell; Donisthorpe, pp 91–92, (partim, *quoad* Plate V, fig. 12).  
 1978 *Oecophylla atavina* Cockerell; Burnham, p. 114.  
 1978 *Oecophylla perdita* Cockerell; Burnham, p. 114.  
 1995 *Oecophylla atavina* Cockerell; Bolton, p. 298.  
 1995 *Oecophylla perdita* Cockerell; Bolton, p. 298.  
 2008 *Oecophylla atavina* Cockerell; Dlussky *et al.*, p. 616

**Holotype.** USNM 61419, Bembridge Marls, NW Isle of Wight, UK; ♂ (Plate 21, figs 1, 2; Text-fig. 77A).

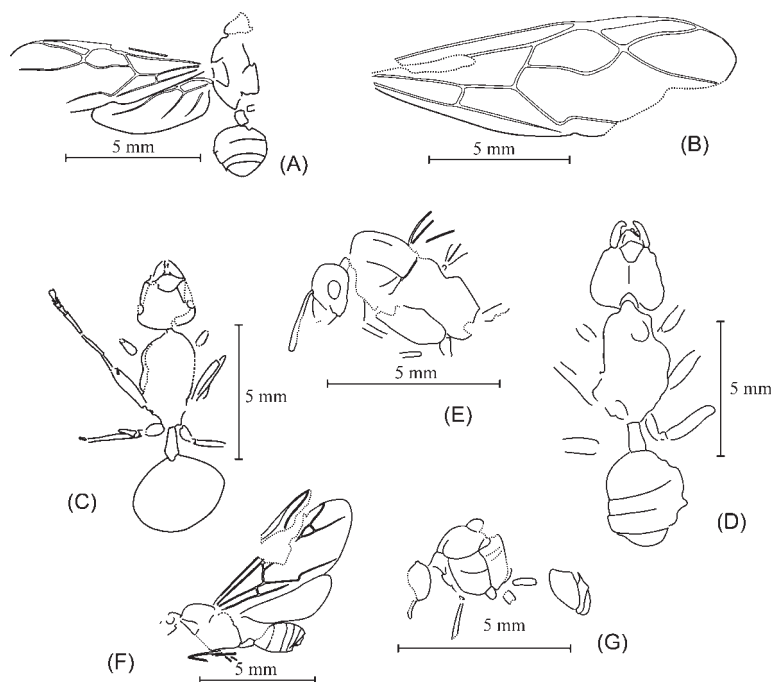
**Other material.** USNM 61420, ♀, forewing, holotype of *Oecophylla perdita* Cockerell (Plate 21, fig. 5; Text-fig. 77B); USNM 61421, ♀, paratype of *Oecophylla perdita* Cockerell; USNM 61422 ♂, forewing, paratype of *Oecophylla perdita* Cockerell, coll. Lacoe (Plate 21, fig. 5); 156 males and 29 females in the NHMUK collection and six males in the CAMSM collection, including NHMUK I.8702, ♀, designated by Donisthorpe (1920) as the plesiotype of *O. megarche* Cockerell (Text-fig. 77D), NHMUK I.8711 (♀, identified by Donisthorpe as *O. megarche* Cockerell, Plate 21, fig. 4; Text-fig. 77C), NHMUK I.8768 (Text-fig. 77E), NHMUK I.9143 (Plate 21, fig. 3; Text-fig. 77G) and NHMUK I.9345 (♂) (Text-fig. 77F). All from Bembridge Marls, Isle of Wight, UK.

**Diagnosis.** Smaller species: ♂AL 2.0–2.8 mm; FWL 6.0–9.5 mm; ♀ AL 3.5–4.5 mm; FWL 8.5–13.7 mm.

**Description.** Female. BL 8–12 mm, AL 3.5–4.5 mm, FWL 8.5–13.7 mm. Head subtrapezoidal, narrower in front than behind, about as long as wide. Occipital corners rounded, occipital margin feebly convex. Anterior margin of clypeus rounded. Eyes comparative small, oval, situated slightly behind midlength of head sides; maximum eye diameter about 0.3 head length. Mandibles large, triangular with large sharp teeth. Scape protrudes far beyond the occipital margin of the head, about twice as long as head. Alitrunk massive, a little wider than head. Scutum wider than long (in NHMUK I.8527, ScW/ScL = 1.2), flat above, rounded anteriorly, overhanging pronotum in dorsal view. Propodeum feebly convex in



**Plate 21** (1–5) *Oecophylla atavina* Cockerell: (1–2) holotype, USNM 61419 (part and counterpart); (3) ♂, NHMUK I.9143; (4) ♀, NHMUK I.8711; (5) holotype of *Oecophylla perdita* Cockerell, USNM 61420. (6) *Oecophylla megarche* Cockerell, holotype, USNM 61423. Scale bars = 1 mm.



**Text-figure 77** *Oecophylla atavina* Cockerell: (A) ♂, holotype, USNM 61419; (B) forewing, holotype of *Oecophylla perdita* Cockerell, USNM 61420; (C) ♀, NHMUK I.8711; (D) ♀, NHMUK I.8702; (E) ♀, NHMUK I.8968; (F) ♂, NHMUK I.9345; (G) ♂, NHMUK I.9143.

side view. Legs rather short and thick; hind femur shorter than alitrunk. Petiole without scale or node, narrowed anteriorly, twice as long as wide and high. Gaster oval.

Male. BL 5–6.3 mm, AL 2.0–2.8 mm, FWL 6–9.5 mm. Head comparatively small, considerably narrower than alitrunk. Eyes convex. Alitrunk short and high (in NHMUK I.9143, 1.2 times as long as high). Scutum convex, a little longer than wide. Propodeum feebly convex or straight in side view. Legs rather long and thin. Petiole without scale or node, narrowed anteriorly, 2.5–3 times as long as wide and high. Gaster oviform, narrowed behind. Posterior margin of subgenital plate uniformly concave. Genitals small, poorly exposed from gaster. Genital stipes with rounded apex, about 1.5 times as long as wide.

Forewing IRS proclined, forming acute angle with R, 2r–rs proclined. Cell 3r comparatively narrow, 5RS bent into 3r. Section 2–3RS (delimiting cell 1+2r from below) bent. 3Cu convex. Hind wing with two longitudinal veins RS and Cu. 1RS distinct, short. Section rs–cu concave, much longer than 2M+Cu. Cross-vein cu–a about as long as rs–cu, placed near it.

**Measurements in mm.** USNM 61419 (♂, holotype): BL = 5.9; AL = 2.7; FWL = 7.6. USNM 61420 (♀, holotype of *O. perdita*): FWL = 12.7. USNM 61421 (♀, paratype of *O. perdita*): FWL = 11.8. USNM 61422 (♂, paratype of *O. perdita*): HW = 0.90. SL = 0.94. FWL = 6.8. Other specimens: Males: BL = 5.0–6.3 (average = 5.61, std = 0.46, n = 16). AL = 2.0–2.8 (average = 2.44, std = 0.35, n = 11). PtL = 0.6–0.75 (n = 3). FWL = 6.0–9.4 (average = 7.66, std = 0.723, n = 81). Females: BL = 8.2–12.1 (n = 20). AL = 2.6–4.6 (average = 4.0, std = 0.69, n = 8). HL = 1.5–2.8 (n = 5). HW = 2.8 (n = 1). SL = 1.9 (n = 1). ED = 0.4–0.5 (n = 2). PtL = 0.9–1.1 (n = 4). FWL = 10.0–15.9 (average = 11.72, std = 1.408, n = 22).

**Remarks.** Donisthorpe (1920) identified the wingless females NHMUK I.8702 and NHMUK I.8711 as *O. megarche*, and even designated the former a plesiotype and represented it on Pl. V. fig. 12. However, their size (AL 4.6 and 3.7 mm, respectively) indicates their FWL is not above 13 mm, well below the range of *O. megarche*.

*Oecophylla megarche* Cockerell, 1915  
Plate 21, fig. 6; Text-fig. 78A–E

1915 *Oecophylla megarche* Cockerell, p. 486, plate 65, figs. 1–3.

1920 *Oecophylla megarche* Cockerell; Donisthorpe, pp 91–92 (part., *quoad* fig. 11).

1978 *Oecophylla megarche* Cockerell; Burnham, p. 114.

1995 *Oecophylla megarche* Cockerell; Bolton, p. 298.

**Holotype.** USNM 61423, Bembridge Marls, NW Isle of Wight, UK, coll. Lacoe; ♀ forewing (Plate 21, fig. 6; Text-fig. 78A).

**Paratypes.** USNM 61424, Bembridge Marls, NW Isle of Wight, UK; hind wing of ♀; USNM 61425, Bembridge Marls, NW Isle of Wight, UK, coll. Lacoe; ♀ (Text-fig. 78B).

**Other material.** 46 males and 68 females in the NHMUK collection and on male and four females in the CAMSM collection, including: NHMUK I.8729 (Text-fig. 78D); NHMUK In.24391 (Text-fig. 78C); NHMUK In.24909 (Text-fig. 78E). All from Bembridge Marls, Isle of Wight, UK.

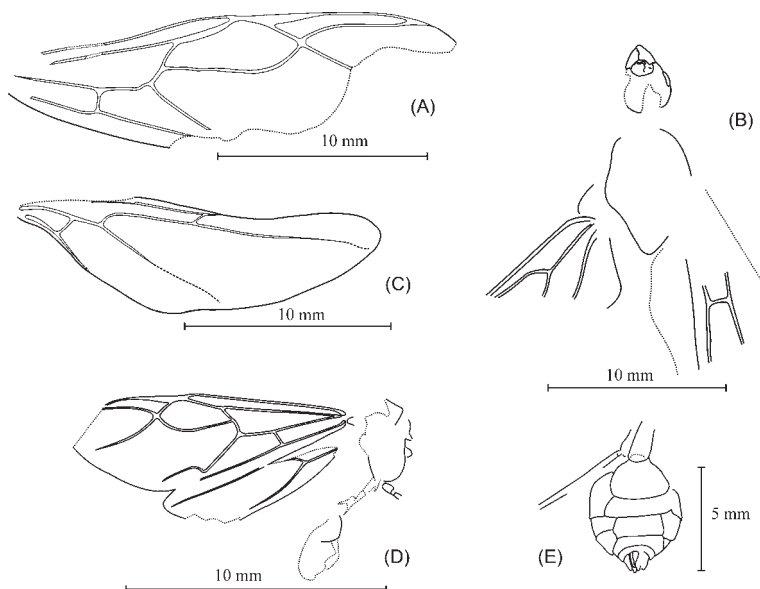
**Diagnosis.** Bigger species: ♂ AL 4–5.5 mm, FWL 10–15.9 mm, ♀ FWL 20.7–23.0 mm.

**Description.** Male. AL 4–5.5 mm, FWL 10–16 mm. Similar to *O. atavina*, except for larger size, shorter petiole and longer genital stipes. Scutum a little longer than wide (1.1 times as long as wide in NHMUK I.8670). Petiole 1.8–2.1 times longer than wide. Genital stipes twice as long as wide.

Female. FWL 20–24 mm. Based on proportions of *O. atavina* and living *O. smaragdina*, apparently AL c. 6 mm, BL > 15 mm. The only known head about as long as wide and c. 3.5 times as long as maximum eye diameter (paratype USNM 61425, Text-fig. 37).

Wing venation of fore and hind wings as in *O. atavina*.

**Measurements in mm.** USNM 61423 (♀, holotype): FWL = 20.7. USNM 61424 (♀, paratype): hind wing 16.8. USNM 61425 (♀, paratype): HW = 3.2. NHMUK I.8703 (♂): AL = 4.0; FWL = 11.4. NHMUK I.10086 (♂): PtL = 1.3. Other specimens: ♂♂: FWL = 10.0–15.9 (average = 11.74, std = 1.408, n = 40). ♀♀: FWL = 20.7–23.0 (average = 21.94, std = 0.767, n = 25).



**Text-figure 78** *Oecophylla megarche* Cockerell: (A) ♀ forewing, holotype, USNM 61423; (B) ♀, paratype, USNM 61425; (C) hind wing of ♀, NHMUK In.24391; (D) ♂, NHMUK I.8729; (E) gaster of ♂ from above, NHMUK In.24909.

#### 3.11.1.4. Subfamily Ponerinae Lepeletier, 1836

Morphogenus *Ponerites* Dlussky & Rasnitsyn, 2002

**Type species.** *Ponerites eocenicus* Dlussky & Rasnitsyn, 2002, by original designation.

**Other species.** *P. coloradensis* Dlussky & Rasnitsyn, 2002 and *P. hypoponeroides* Dlussky & Rasnitsyn, 2002 from the Green River Formation, Middle Eocene of USA (Dlussky & Rasnitsyn 2002), *P. umbrus* (Popov 1932) from the Middle Miocene of North Caucasus, and *P. crawleyi* (Donisthorpe, 1920) and two new species from the Bembridge Marls described below. Some European Miocene and Oligocene ants described as *Ponera* also apparently deserve placement here.

**Diagnosis.** Fossil ants not well enough preserved to fit orthotaxa, and with following combination of traits: size small to medium, head subrectangular with rounded occipital corners; eyes comparatively small, situated before midlength of head sides; mandibles triangular with denticulate masticatory margin; promesonotal and mesopropodeal sutures present in workers; sculpture conspicuous, head, alitrunk and waist with no denticles or spines; petiole with thick scale; gaster with constriction between first and second segments of gaster (III and IV abdominal segments); second gastral tergite and sternite subequal, apex of the gaster not directed downwards; sting well developed. Forewing with closed cells 1+2r, 3r, rm and mcu; cross-vein cu—a situated near mcu, so usually  $Icu < 1.45$  and  $Icu < 1.2$ .

**Remarks.** Key traits used to distinguish extant genera of the tribe Ponerini (shape of ventral petiolar lobe, dentition of mandibular masticatory margin, number of spurs of middle and hind tibiae) are not visible in most rock fossils of small, nonspecialised Ponerinae. Taylor (1964) designated these fossils as “*Ponera* (?)”, and the informal name “*Ponerites*” was later coined for them (Dlussky 1981a). Later Dlussky & Rasnitsyn (2002) proposed to use the latter name as valid for a morphogenus with the above diagnosis.

The extant genera *Ponera* Latreille, 1804, *Hypoponera* Santachi, 1938 and, partially, *Pachycondyla* F. Smith, 1858 and perhaps some other ponerine genera agree with the diagnosis of *Ponerites*.

*Ponerites crawleyi* (Donisthorpe, 1920), comb. nov.  
Plate 22, fig. 1; Text-fig. 79A–E

1920 *Euponera* (*Mesoponera*) *crawleyi* Donisthorpe, p. 85, plate V, fig. 3.

1978 *Euponera* (*Mesoponera*) *crawleyi* Donisthorpe: Burnham, p. 109.

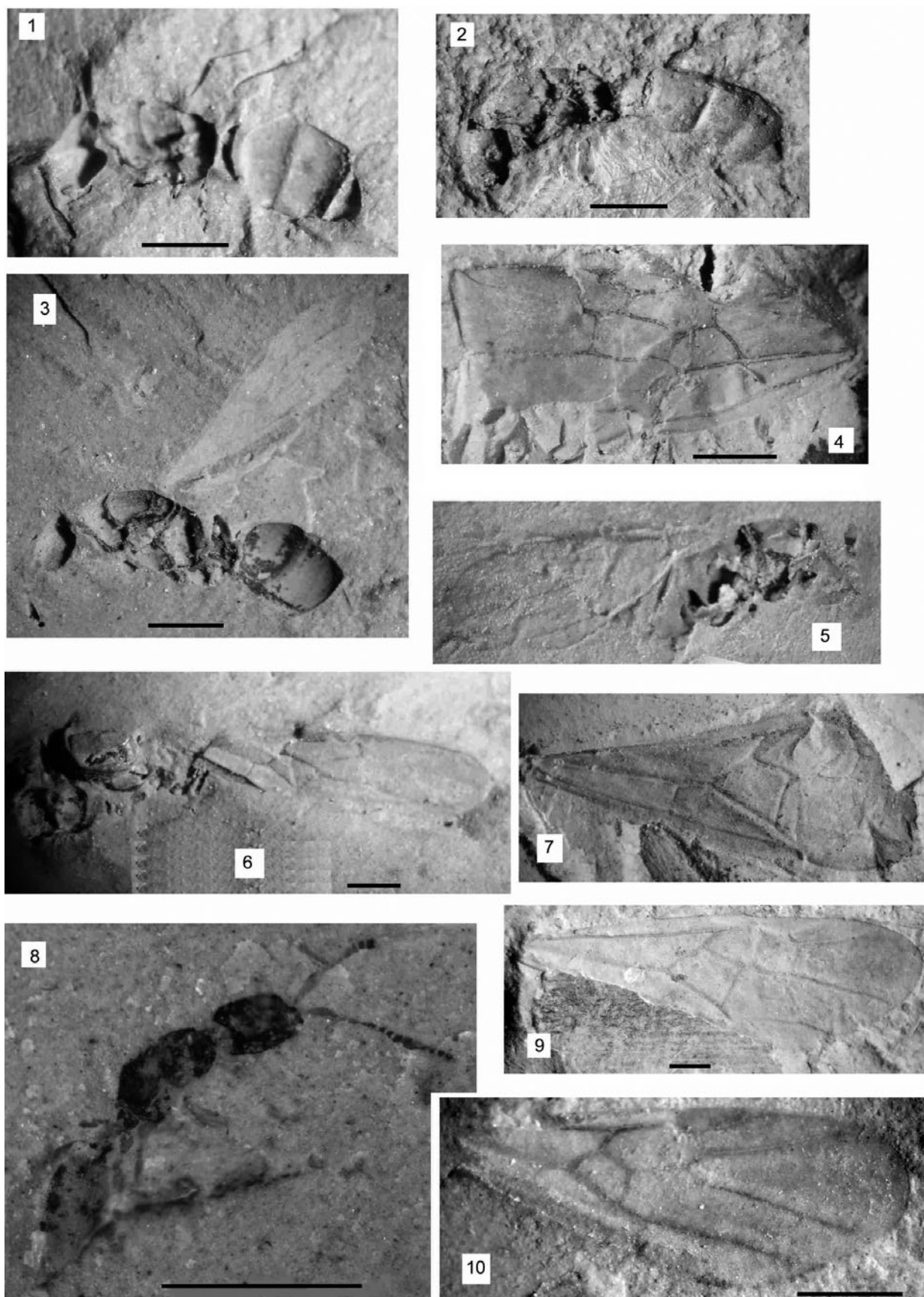
1995 *Pachycondyla crawleyi* (Donisthorpe): Bolton, p. 304.

**Holotype.** NHMUK I.8675/I.8539 print and counterprint, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie; ♀ (Plate 22, fig. 1, Text-fig. 79A, B).

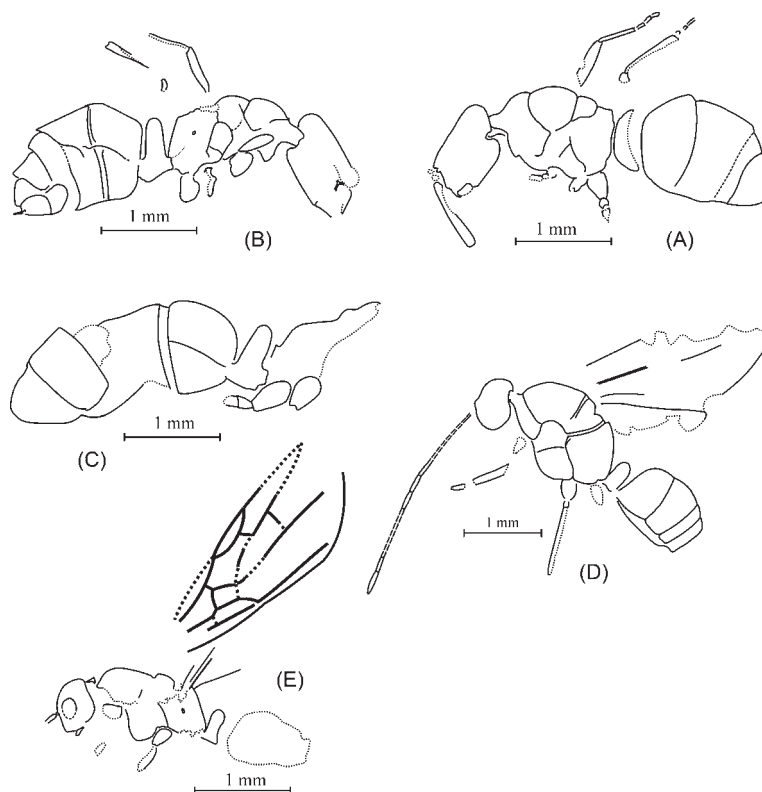
**Other material.** Three females and three males in the NHMUK collection, including NHMUK I.8759 (♀) (Text-fig. 79C), NHMUK I.9406 (♂) (Text-fig. 79E) and NHMUK I.9774, coll. P. B. Brodie; ♂ (Text-fig. 79D); all from Bembridge Marls, Isle of Wight, UK.

**Description.** Female. BL c. 4 mm, AL c. 1.5 mm. Head elongate, longer than wide, with parallel sides, rounded occipital corners, and feebly concave occipital margin. Eyes oval, strongly displaced forward, length of cheek less than scape diameter. Scape reaching occipital margin or perhaps slightly protruding beyond it. Mandibles well developed, triangular, with numerous small teeth on masticatory margin. Scutum feebly convex, not overhanging pronotum anteriorly. Scutum and scutellum separated with distinct impression in side view. Mesopleural suture well developed. Propodeum angulated in side view with dorsum somewhat shorter than declivity. Petiole 1.6–1.7 times as high as long, with high scale, in side view, with almost parallel anterior and posterior surfaces and smoothly rounded top. Rear part of petiole with ventral rounded lobe. No spurs preserved with fragments of middle and hind tibiae as preserved in holotype. Tubular articulation between first and second gastral segments well developed (preserved in specimens NHMUK I.8539 and NHMUK I.8759). First gastral segment higher than long. Sting short and sharp. Preserved organic integument of alitrunk and gaster with very fine shagreened sculpture.

Male. BL 3–3.5 mm. Eyes of moderate size, oval, displaced a little forward. Antenna filiform, long, deflexed back reaching



**Plate 22** (1) *Ponerites crawleyi* (Donisthorpe), comb. nov., holotype (*Euponera crawleyi* Donisthorpe), NHMUK I.8675. (2) *Ponerites antropovi* Dlussky, sp. nov., holotype, NHMUK I.9996. (3) *Ponerites hooleyi* Dlussky & Perfilieva sp. nov., holotype, NHMUK In.24971. (4–5) *Taphopone macroptera* Perfilieva, 1 sp. nov.: (4) holotype, NHMUK I.9494; (5) paratype, NHMUK In.24378. (6) *Taphopone petrosa* Perfilieva, sp. nov., holotype, NHMUK In.24369. (7) *Taphopone aberrans* Perfilieva, sp. nov., paratype, NHMUK In.24910. (8) *Solenopsites rossi* Dlussky, sp. nov., holotype, NHMUK I.9968. (9) *Taphopone aberrans* Perfilieva, sp. nov., holotype, NHMUK In.24917. (10) *Paraphaenogaster hooleyana* Perfilieva, sp. nov., holotype, NHMUK In.24963. Scale bars = 1 mm.



**Text-figure 79** *Ponerites crawleyi* (Donisthorpe): (A) ♀, holotype of *Euponera crawleyi* Donisthorpe, NHMUK I.8675; (B) counterprint of the same specimen, identified by Donisthorpe as *Leucotaphus gurnetensis* (Cockerell); (C) ♀, NHMUK I.8759; (D) ♂, NHMUK I.9774; (E) ♂ NHMUK I.9406.

middle of gaster. Scape very short, flagellomeres elongate, four times as long as thick in middle of flagellum. In side view, scutum uniformly convex, propodeum gradually rounded. Propodeal spiracles small, oval. Legs rather long and thin. Petiole with high scale, in side view, with almost parallel anterior and posterior surfaces and smoothly round top. Constriction between first and second gastral segments poorly expressed.

Forewing (known only for males) with closed cells 1+2r, rm, mcu (specimen NHMUK I.9406), and supposedly 3r. 1RS perpendicular to R. 1M more than twice as long as 1RS. Cross-vein 2r-rs slightly proclined, rs-m more distal than 2r-rs, a little distal comparing level of pterostigmal apex. Cell rm triangular, not pedunculate. Cell mcu of medium size, rhomboid, reaching level of pterostigmal base. Cross-vein cu-a joining 1M+Cu near mcu. Icu = 1.25–1.3, Icu-a = 1.1.

**Measurements in mm.** NHMUK I.8675 (♀, holotype): BL = 3.6; AL = 1.5; HL = 0.9; SL = 0.75; ScL = 0.5; SctL = 0.23; PtL = 0.31; PtH = 0.60. NHMUK I.8759 (♀): PtL = 0.39; PtH = 0.69. NHMUK I.9774 (♂): AL = 1.6; ScL = 0.7; AntL = 2.7; PtH = 0.4. NHMUK I.9406 (♂): AL = 1.3; HL = 0.55; FWL = 2.9; PtH = 0.34.

**Remarks.** Donisthorpe (1920) misidentified specimens NHMUK I.8539, NHMUK I.8684, NHMUK I.8759, NHMUK I.9406 and NHMUK I.9774 as *Leucotaphus gurnetensis*. Constriction between first and second gastral segments, the key character of Ponerinae, is clearly visible on specimens NHMUK I.8539, NHMUK I.8759 and NHMUK I.9774. Furthermore, NHMUK I.8539 represents a counterpart of NHMUK I.8675, the holotype of *Euponera crawleyi*. Forewings of NHMUK I.9406 show cross-vein cu-a near cell mcu (Icu = 1.3; Icu-a = 1.1), which is equally a key character of Ponerinae. Male NHMUK I.9774 has filiform antennae unlike males of *Leucotaphus*, which have geniculate antennae.

Unfortunately there is no diagnostic character discriminating *Ponera*, *Hypoponera* or *Pachycondyla* (= *Euponera*) (tibial

spur formula, ventral lobe of petiole, dentation of mandibles etc.) preserved in the material available, thus forcing us to refer this species to the morphogenus *Ponerites*.

*Ponerites antropovi* Dlussky & Perfilieva, sp. nov.

Plate 22, fig. 2; Text-fig. 80

**Etymology.** After Alexander V. Antropov who was the first to identify this ant as Ponerinae.

**Holotype.** NHMUK I.9996, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie; ♀ (Plate 19, fig. 22; Text-fig. 80).

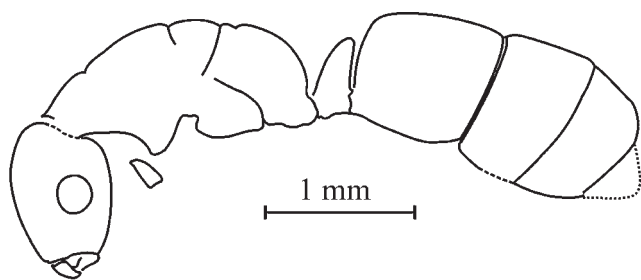
**Diagnosis.** Differs from *P. crawleyi* and *P. hooleyi* in position of eyes near the middle of head sides, propodeum rounded in side view, scale with fore-surface feebly convex, and in elongated first gastral segment.

**Description.** Female. BL 4.5 mm. Eyes round, situated near the middle of head sides. Scutum feebly convex, not overhanging pronotum anteriorly. Scutellum feebly convex. Propodeum rounded in side view. Petiole with high scale, fore-surface of scale feebly convex, back surface feebly concave, top rounded. Gaster with distinct constriction between first and second segments showing well preserved tubular rings, first gastral segment a little longer than high.

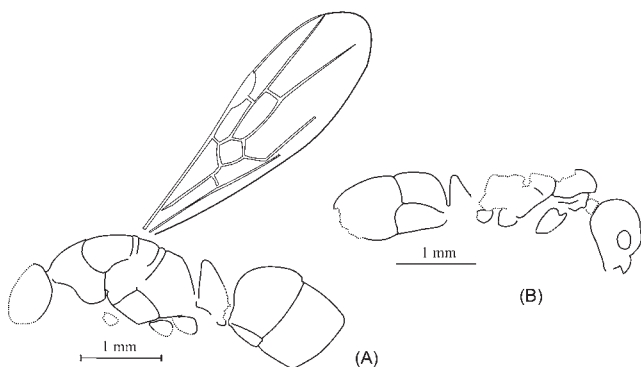
Male unknown.

**Measurements of holotype in mm.** AL = 1.65; HL = 1.0; ED = 0.25.

**Remarks.** Donisthorpe misidentified the holotype as *Leucotaphus gurnetensis*. During preliminary study of the material, Dr Alexander Antropov correctly identified it as Ponerinae. The new species is attributed to the morphogenus *Ponerites* because the diagnostic characters of the genera *Ponera*, *Hypoponera* or *Pachycondyla* (= *Euponera*) (tibial spur formula, ventral lobe of petiole, dentation of mandibles etc.) cannot be seen in the current material.



**Text-figure 80** *Ponerites antropovi* Dlussky, sp. nov., ♀, holotype, NHMUK I.9996.



**Text-figure 81** *Ponerites hooleyi* Dlussky et Perfilieva, sp. nov.: (A) ♀, holotype, NHMUK In.24971; (B) ♀, paratype, NHMUK I.9869.

*Ponerites hooleyi* Dlussky & Perfilieva sp. nov.  
Plate 22, fig. 3; Text-fig. 81A, B.

**Etymology.** In honour of R. W. Hooley, collector of the type specimen.

**Holotype.** NHMUK In.24971, Bembridge Marls, NW Isle of Wight, UK, coll. R. W. Hooley; ♀ (Plate 22, fig. 3; Text-fig. 81A).

**Paratypes.** NHMUK I.9602 (♂), NHMUK I.9869 (♀) (Text-fig. 81B); both from Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie.

**Diagnosis.** Differs from *P. crawleyi* by high scale, in side view, clearly narrowed upwards with a narrowly round top; in propodeum angulate, in side view, eyes displaced forward; differs from *P. antropovi* by the above position of eyes and short first segment of the gaster.

**Description.** Female. BL about 4–5 mm. Eyes oval, displaced forward; length of cheek similar to maximal diameter of eye. Scutum rather small, feebly convex, not over-hanging pronotum anteriorly. Scutellum feebly convex. Mesopleural suture well developed. Propodeum in side view angulated; propodeal dorsum much shorter than propodeal declivity. Petiole more high than long, with high scale, in side view clearly narrowed upwards with the narrowly round top. Gaster with distinct constriction between first and second segments. First segment of the gaster higher than long.

Male. BL about 3 mm. Head absent in both paratypes. Scutum and scutellum are feebly convex. Propodeum rounded in side view. Petiole in side view triangulate, with rounded top. Gaster with weak constriction between first and second segments.

Forewing. 1RS and 2r–rs perpendicular to R. Cell 3r comparatively long, more than four times as long as wide, its apex touching wing margin. Cell rm quadrangular. Cross-vein rs–m far distal in respect of 2r–rs, but not distal of level of pterostigmal apex. Cell mcu pentagonal, in terms of area more than

half as large as 1+2r. 2M and RS+M of subequal length. Cell mcu not reaching level of pterostigmal base. 1M more than twice as long as 1RS. Cross-vein cu–a meeting 1M+Cu near mcu. Icu = 1.4, Icu<sub>a</sub> = 1.13.

**Measurements in mm.** NHMUK In.24971 (♀, holotype): AL = 1.9; FWL = 3.75. NHMUK I.9859 (♀, paratype): AL = 1.5; HL = 0.9; ED = 0.23. NHMUK I.9602 (♂, paratype): BL = 3.1, AL = 1.25. NHMUK In.24378 (♂, paratype): AL = 1.5, FWL = 3.0.

**Remarks.** Donisthorpe (1920) identified specimens NHMUK I.9602, NHMUK I.9869, and NHMUK In.24378 as *Leucotaphus gurnetensis*. However, the presence of a constriction between the first and second segments of NHMUK I.9602 and NHMUK I.9869 and the presence of closed cell rm and position of cross-vein cu–a on forewing of NHMUK In.24378 indicate these ants belong to subfamily Ponerinae.

As no diagnostic characters separating the genera *Ponera*, *Hypoponera* or *Pachycondyla* (tibial spur formula, ventral lobe of petiole, dentation of mandibles etc.) can be seen on impressions, we include this species in the morphogenus *Ponerites*.

Morphogenus *Taphopone* Dlussky & Perfilieva, gen. nov.

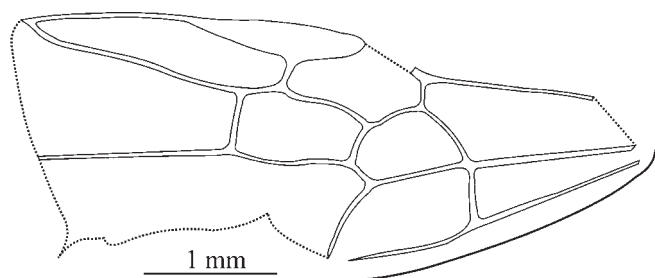
**Etymology.** From Greek *taphos* for grave and genus *Ponera*. Gender feminine.

**Types species.** “*Ponerites*” *karaganensis* Dlussky, 1981, Miocene, Vishnevaya Balka, Stauropol reg., Russia.

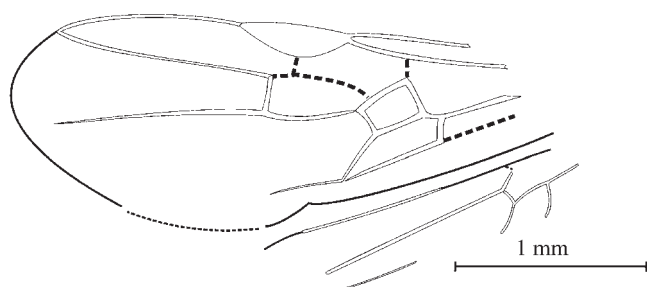
**Other species.** Attributed with confidence to this morphogenus are *Taphopone karaganensis* (Dlussky, 1981), comb. nov. (= “*Ponerites*” *karaganensis*) and *Taphopone stauropolitana* (Dlussky, 1981), comb. nov. (= “*Ponerites*” *stauropolitanus*) from the Miocene of Stavropol region (Russia) and four species described below. Most likely, some other species from other Miocene deposits of Europe, described in genera *Ponera* Latreille, 1804 and *Poneropsis* Heer, 1867 and included in the morphogenus *Poneropsis* Heer, 1867 by Taylor (1964), may also deserve transfer to this genus.

**Diagnosis.** Isolated ant forewings with closed cells 1+2r, 3r, rm and mcu, and with distal position of cross-vein cu–a (usually with Icu < 1.4). Cross-vein 1r–rs (subdividing cell 1+2r on two separated cells), or its rudiment, lost (unlike some Armaniidae and Sphecomyrminae). Cells 3r, rm and mcu are not grouped in the central part of the wing (unlike *Formicium* Westwood, 1854, subfamily Formiciinae).

**Remarks.** While revising the genus *Ponera* Latreille, 1804, Taylor (1964) placed 19 species described previously from forewing fossils from the Miocene of Europe as *Ponera* Latreille and *Poneropsis* Heer, as well as *Ponera hypolitha* Cockerell from the Bembridge Marls, in the morphogenus *Poneropsis*. Taylor’s re-definition of *Poneropsis* resulted in its much wider application for all detached ant wings with complete venation. This is because Taylor overlooked some important traits, particularly the position of cross-vein cu–a, which can be used to distinguish forewings of Ponerinae and several other primitive ant subfamilies (Armaniinae, Sphecomyrminae, Myrmeciinae, Amblyoponinae, Ectatomminae, Cerapachyinae, Ecitoninae) from the venationally advanced subfamilies Myrmicinae, Aneuretinae, Dolichoderinae and Formicinae. As a result, *Poneropsis* sensu Taylor can accommodate the majority of the ant wing fossils. The name *Poneropsis* further causes confusion. Mayr (1867) re-examined three specimens identified as *Ponera fuliginosa* (type species of genus *Poneropsis*) by Heer and found that they had a two-segmented waist and might be females of *Aphaenogaster* Mayr, 1853, a genus of Myrmicinae. Therefore we propose a new morphogenus for impressions of ant forewings with closed cells 1+2r, 3r, rm and mcu, and



**Text-figure 82** *Taphopone macroptera* Perfilieva, sp. nov., forewing, holotype, NHMUK I.9494.



**Text-figure 83** *Taphopone microptera* Perfilieva, sp. nov., forewing, holotype, NHMUK In.9763.

with a distal position to cross-vein cu-a ( $Icu < 1.4$ ), which certainly belong to Ponerinae or other venationally-primitive ant subfamilies. We also include some additional characters in the diagnosis which exclude the most archaic (Armaniidae, Sphecomyrminae) and aberrant ants (Formiciinae).

The new species are diagnosed in the key below (section 3.11.3).

*Taphopone macroptera* Dlussky & Perfilieva, sp. nov.  
Plate 22, fig. 4; Text-fig. 82

**Etymology.** *Macroptera* is Latin for “large wings”.

**Holotype.** NHMUK I.9494, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie; forewing (Plate 22, fig. 4; Text-fig. 82).

**Paratype.** NHMUK I.10172, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie; forewing (Plate 22, fig. 5).

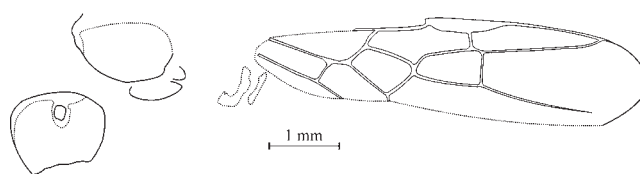
**Description.** FWL  $c. 5.5$ – $6$  mm. IRS slightly reclined, not strictly perpendicular to R. Cell 3r comparatively short (less than four times as long as wide), its apex touching wing margin. Cell rm quadrangular, rs-m distal in respect to 2r-rs, but not in respect of pterostigmal apex. 2r-rs somewhat proclined. Cell mcu large (more than half cell 1+2r in terms of area), pentagonal, 2M shorter than RS+M. Cross-vein m-cu surpassing level of pterostigmal base. 1M more than twice as long as 1RS. Cell cua well developed. Cross-vein cu-a almost reaching mcu (2M+Cu practically lost).  $Icu = 1.05$ ,  $Icua = 1.01$  (holotype; paratype  $Icu = 1.09$ ).

**Remarks.** The holotype is labelled as *Pachycondyla* sp., however Donisthorpe did not mention this specimen in his paper. This might well be correct, but cannot be reasonably confirmed because of the absence of diagnostic characters (cf. above). That is why the new species is included in the morpho-genus *Taphopone*.

*T. macroptera* is venationally similar to *Ponerites hooleyi*, but differs in having a larger wing (3–4 mm in the latter).

*Taphopone microptera* Dlussky & Perfilieva, sp. nov.  
Text-fig. 83

**Etymology.** *Microptera* is Latin for “small wings”.



**Text-figure 84** *Taphopone petrosa* Perfilieva, sp. nov., holotype, NHMUK In.24369.

**Holotype.** NHMUK I.9763, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie; forewing (Text-fig. 83).

**Paratype.** NHMUK In.24378 Bembridge Marls, NW Isle of Wight, UK, coll. R. W. Hooley; forewing.

**Description.** FWL  $c. 3$  mm. Cell 3r comparatively long (four times as long as wide), touching wing margin. Cell rm quadrangular. Cross-vein rs-m far distal in relation to 2r-rs but not distal of pterostigmal apex. Cell mcu pentagonal, reaching level of pterostigmal base. Cross-vein m-cu and 1M shorter than 1Cu. Cell cua well developed. Cross-vein cu-a almost reaching mcu (2M+Cu practically lost).  $Icu = 1.1$ – $1.3$ ,  $Icua = 1.1$  (holotype).

Hindwing (holotype) with three longitudinal veins RS, M, and Cu. 1RS and rs-m of near equal length, m-cu longitudinal, longer than rs-m.

**Measurements in mm** (holotype): AL = 1.55; FWL = 3.1; FWW = 0.9.

**Remarks.** Donisthorpe misidentified both forewings of this species as *Leucotaphus gurnetensis*, even though they have cell rm closed and forewing cross-vein cu-a takes distal position.

*Taphopone petrosa* Dlussky & Perfilieva, sp. nov.  
Plate 22, fig. 6; Text-fig. 84

**Etymology.** *petrosa* is Latin for stony.

**Holotype** NHMUK In.24369, Bembridge Marls, NW Isle of Wight, UK, coll. R. W. Hooley; ♀ (Plate 22, fig. 6, Text-fig. 84).

**Paratypes.** NHMUK In.24927; NHMUK In.24949 (forewings); NHMUK In.25006 (two forewings), coll. R. W. Hooley; CAMSM X.50140.21 (TN 71) (forewing). All from Bembridge Marls, Isle of Wight, UK,

**Description.** Female. FWL 6–7.5 mm. Head subrectangular, wider than long, with convex sides and rounded occipital corners.

Forewing. 1RS reclined. Apex of cell 3r touching wing margin. Cell rm triangular with short stem or not pedunculate, 2.2–2.4 times as long as wide. Cross-vein rs-m level with 2r-rs, or more distal than that but not more than for vein width. Cell mcu trapezoid, of medium size, 1M shorter than m-cu. M+Cu distinctly bent at junction with cu-a. Cross-vein cu-a near cell mcu.  $Icu 1.3$ – $1.4$ ,  $Icua 1.14$ – $1.18$ .

**Measurements in mm.** Holotype: HW = 1.45; FWL = 6.1. Paratype NHMUK In.24927: FWL = 6.1; FWW = 2.1. Paratype NHMUK In.24949: FWL = 7.2; FWW = 1.9.

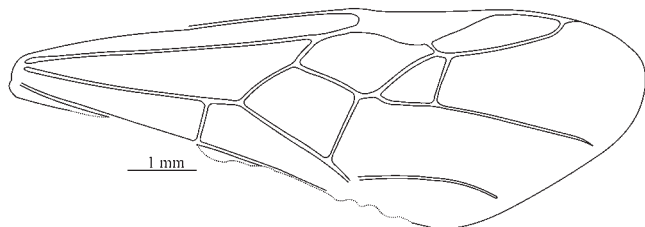
**Remarks.** Donisthorpe (1920) misidentified the holotype as *Dolichoderus britannicus*, even though the distal position of cross-vein cu-a indicates clearly that it belongs to Ponerinae.

*Taphopone aberrans* Dlussky & Perfilieva, sp. nov.  
Plate 22, figs 7, 9; Text-fig. 85

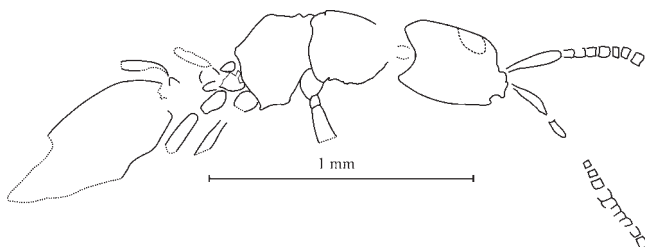
**Etymology.** *Aberrans* is Latin for aberrant.

**Holotype.** NHMUK In.24917, Bembridge Marls, NW Isle of Wight, UK, coll. R. W. Hooley; forewing (Plate 22, fig. 9; Text-fig. 85).

**Paratypes.** Forewings. NHMUK I.8574, coll. P. B. Brodie; NHMUK In.24847; NHMUK In.24910 (Plate 22, fig. 7);



**Text-figure 85** *Taphopone aberrans* Perfilieva, sp. nov., forewing, holotype, NHMUK In.24917.



**Text-figure 86** *Solenopsites rossi* Dlussky, sp. nov., ♀, holotype, NHMUK I.9968.

NHMUK In.24917; NHMUK In.24960, coll. R. W. Hooley; CAMSM: X.50140.117 (TN.173). All from Bembridge Marls, Isle of Wight, UK,

**Description.** Forewing. FWL *c.* 9 mm. 1RS distinctly reclined. Cell 3r short, less than three times as long as wide, its apex touching wing margin. Cell rm rather small, triangular, usually with long stem. Cross-vein 2r-rs reclined, cross-vein rs-m somewhat distal in respect to 2r-rs, but not beyond pterostigmal apex, often meeting M more proximal than RS. Cell mcu large, subequal to 1+2r in terms of area, trapezoid (RS+M and 1M shorter than m-cu and 1Cu, respectively). 1M more than twice as long as 1RS. Cross-vein m-cu placed distal of level of pterostigmal base. Cross-vein cu-a meeting 1M+Cu near cell mcu. Icu = 1.3–1.45, Icu<sub>a</sub> = 1.13–1.14.

**Measurements in mm.** FWL = 9.1 (holotype).

**Remarks.** The distal position of cu-a indicates the wing belongs to a representative of Ponerinae but differs from other Bembridge Ponerinae in the form of cell rm.

**3.11.1.5. Subfamily Myrmicinae Lepeletier de Saint-Fargeau, 1836**

Morphogenus *Solenopsites* Dlussky & Rasnitsyn, 2002

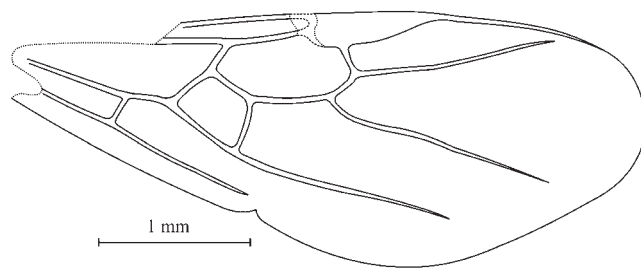
**Type species.** *Solenopsites minutus* Dlussky & Rasnitsyn, 2002, by original designation.

**Other species.** *S. minutus* (Middle Eocene, Green River Formation, USA) and *S. rossi* sp. nov. (Bembridge Marls, Isle of Wight, UK).

**Diagnosis.** Ant rock fossils with waist two-segmented and long (more than half as long as gaster and nearly half as long as the alitrunk), with propodeum unarmed, petiole pedunculate and postpetiole narrowly attached to gaster, which cannot be identified more precisely.

**Remarks.** This morphogenus can include all poorly preserved impressions of small Myrmicinae without propodeal spines or teeth and with a pedunculate petiole, but really belong to many different orthotaxa.

*Solenopsites rossi* Dlussky & Perfilieva, sp. nov.  
Plate 22, fig. 8; Text-fig. 86



**Text-figure 87** *Paraphaenogaster hooleyana* Perfilieva, sp. nov., forewing, holotype, NHMUK In.24963.

**Etymology.** In honour of Dr Andrew Ross, appreciating his help in preparing this work.

**Holotype** NHMUK I.9968, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie, ♀ (Plate 22, fig. 8; Text-fig. 86).

**Description.** Wingless female. BL about 2 mm. Head sub-rectangular, longer than wide, with feebly convex sides, rounded occipital corners and concave occipital margin. Eyes oval, strongly displaced forward. Antennae geniculate, scape thick and short, not reaching the occipital margin, middle flagellomeres thicker than long. Propodeum angulate in side view, without spines or teeth. Legs short and thick.

**Measurements in mm.** AL = 0.65; HL = 0.39, HW = 0.29; SL = 0.23.

**Remarks.** Despite the waist being only partially preserved, position of the alitrunk, gaster and preserved fragments of the waist indicate two-segmented waist. The new species differs from all other ants in the Bembridge assemblage in its small size, and from *S. minutus* in the form of head.

Morphogenus *Paraphaenogaster* Dlussky, 1981

**Type species.** *Paraphaenogaster microphthalmus* Dlussky, 1981, by original designation.

**Other species.** *P. hooleyana* sp. nov. *Aphaenogaster shanwangensis* (Hong, 1984), *A. lapidescens* Zhang, and *A. paludosa* Zhang from the Middle Miocene of Shanwang (Shandong, China) are similar venationally (Zhang, 1989) and might belong there as well.

**Diagnosis.** Isolated ant wings with cells 1+2r and mcu, cell 3r open at apex, and r-m present, apical sections of RS and M leaving cell 1+2r+rm separately.

**Remarks.** Similar venation is characteristic of some Myrmicinae: Pheidolini (a part of *Aphaenogaster* Mayr) and Solenopsidini (*Solenopsis* Westwood, *Oligomyrmex* Mayr).

*Paraphaenogaster hooleyana* Dlussky & Perfilieva, sp. nov.  
Plate 22, fig. 10; Text-fig. 87

**Etymology.** After Mr. R. W. Hooley.

**Holotype.** NHMUK In.24963, Bembridge Marls, NW Isle of Wight, UK, coll. R. W. Hooley; forewing (Plate 22, fig. 10).

**Paratype** NHMUK In.25082, Bembridge Marls, NW Isle of Wight, UK, coll. R. W. Hooley; forewing.

**Description.** Forewing. Cell mcu trapezoid (1Cu appreciably longer than 1M and RS+M), of medium size (not more than half of cell 1+2r in terms of area), not reaching level of pterostigmal base. 4M leaving RS+M basal of 2r-rs and at level of pterostigmal base in holotype (slightly more distal in paratype). 1RS reclined. Cross-vein cu-a well distant from mcu. Icu = 1.5–1.7, Icu<sub>a</sub> ~ 1.2.

**Remarks.** Holotype and paratype wings differ in size significantly, restored values being 4.6 mm and 3.8 mm, respectively. Position of base of apical section of M on cell rm, relative length of 1M and 1RS, and inclination of 2r-rs differ as

well. Nevertheless they may be the same species, supposing the holotype is a female and paratype a male.

There is also a poorly preserved wing NHMUK In.25081 of apparently similar venation, but differing in still smaller size (FWL ~ 3 mm).

### 3.11.2. Species doubtfully and erroneously described as Formicidae

*Camponotus brodiei* Donisthorpe, 1920

1920 *Camponotus brodiei* Donisthorpe, p. 93, plate V. fig. 13.

1995 *Camponotus brodiei* Donisthorpe, Bolton, p. 89.

**Holotype** NHMUK I.9551, Bembridge Marls, NW Isle of Wight, UK, coll. P. B. Brodie.

**Remarks.** The holotype is possibly an ant but so poorly preserved that it does not permit attribution to a subfamily. We refer it to as Formicidae *incertae sedis*.

*Leucotaphus peraneus* Cockerell, 1927

1927 *Leucotaphus peraneus* Cockerell, p. 165 (footnote).

The original description is short and of little help, and the holotype is lost from the USNM collection. We propose to consider this species as *nomen dubium*.

*Ponera minuta* Donisthorpe, 1920

**Remarks.** Belongs to Agaonidae (see Donisthorpe 1920, p. 17).

*Syntaphus wheeleri* Donisthorpe, 1920

**Remarks.** Belongs to Braconidae (see Donisthorpe 1920, p. 69).

**3.11.3. Keys for the identification of ant fossils from the Bembridge Marls.** Preservation of ant fossils in Bembridge Marls is variable: forewings are unknown for some species (*Solenopsis rossi*, *Ponerites antropovi*) and are the only known parts of some others (*Emplastus kozlovi*, *Paraphaenogaster hooleyana* and all *Taphopone* species). This meant that separate keys had to be constructed for bodies and for forewings.

#### 3.11.3.1. Key to bodies with or without wings

- 1 – Waist with one segment (petiole); forewing with closed cell 3r. . . . .go to 2.
- Waist with two segments (petiole and postpetiole); forewing often with opened cell 3r (Myrmicinae). The only myrmecine species with a preserved body is *Solenopsis rossi* sp. nov. BL ~ 2 mm. Head longer than width, subrectangular, with slightly convex sides and concave occipital margin. Wing venation unknown.
- 2 – Gaster with constriction between first and second segments; male antennae filiform, with very short scape; forewing with closed cells 1+2r, 3r, rm and mcu; cross-vein cu—a close to cell mcu, so usually Icu < 1.45 and Icu < 1.2 (Ponerinae). . . . .go to 3.
- Gaster without constriction between first and second segments; cross-vein cu—a displaced proximally, so usually Icu > 1.5 and Icu > 1.2. . . . .go to 5.
- 3 – Petiole with high scale, in side view with almost parallel anterior and posterior surfaces and smoothly round top; forewing with triangular cell rm; BL 3–3.5 mm. ♀: eyes oval; strongly displaced forward, so length of cheek less than scape diameter; propodeum in side view angular, with dorsum somewhat shorter than declivity. ♂: eyes of moderate size, oval, less displaced forward; propodeum evenly rounded in side view. = *Ponerites crawleyi* (Donisthorpe).

- Petiole with high scale, in side view clearly narrowed upwards with narrowly rounded top; cheek about as long as eye diameter or longer. BL 4–4.5 mm. . . . .go to 4.
- 4 – First gastral tergite longer than high; eyes oval, slightly displaced forward; ♀ cheek about as long as maximum eye diameter; wing venation unknown = *Ponerites antropovi* sp. nov.
- First gastral tergite shorter than high; eyes rounded, situated near the middle of head sides; ♀ cheek longer than eye diameter; forewing with cell rm quadrangular = *Ponerites hooleyana* sp. nov.
- 5 – Propodeum angular, with distinct carina separating propodeal dorsum and declivity, with dorsum convex and declivity concave in side view.; petiole with rounded node, 1.5–1.8 times as long as high; forewing with closed cells 1+2r, 3r, rm and mcu; BL 3–3.5 mm; FWL 3.5–4 mm. (Dolichoderinae) = ♀ *Dolichoderus vectensis* Donisthorpe.
- Propodeum gradually rounded or angular in side view, with straight or feebly convex propodeal declivity; petiole of other form. . . . .go to 6.
- 6 – Petiole elongate, longer than wide, without node or scale. . . . .go to 7.
- Petiole with scale (♀) or triangular in side view (♂). . . . .go to 11.
- 7 – Propodeum angular in side view, with blunt teeth on the border of propodeal dorsum and declivity, with dorsum longer than declivity; petiole 1.7 times as long as high and about half as long as propodeum; anterior part of petiolar dorsum almost straight, and posterior feebly rounded in side view; maximum petiole height is behind its midlength; forewing with closed cells 1+2r, rm, mcu and apparently 3r; FWL about 8 mm. (Aneuretinae) = *Britaneuretus anglicus* (Cockerell).
- Propodeum feebly convex or straight in side view; petiole narrowed anteriorly, 1.8–3 times as long as wide and high. scape extending far beyond occipital margin; ♀ mandibles large, triangular, with large sharp teeth; forewings with closed cells 1+2r and 3r and not rm and mcu; 5RS and 4M leaving the same point (no cross-vein rs–m). (Formicinae, *Oecophylla*). . . . .go to 9.
- 8 – BL 5–6.3 mm; AL 2–3 mm; FWL 6–9.5 mm; petiole 2.5–3 times as long as wide and high; scutum a little longer than wide = ♂ *Oecophylla atavina* Cockerell.
- AL > 3.5 mm; FWL usually > 9.5 mm; petiole about twice as long as wide. . . . .go to 9.
- 9 – FWL 19–25 mm, restored BL more than 16 mm = ♀ *Oecophylla megarche* Cockerell.
- FWL 10–16 mm; BL < 10 mm. . . . .go to 10.
- 10 – Scutum a little longer than wide; alitrunk short and high, a little longer than high; gaster with six segments and exposed genitals; AL 3.5–4.5 mm; FWL 8.3–16 mm = ♂ *Oecophylla megarche* Cockerell.
- Scutum a little wider than long; alitrunk longer, nearly twice as long as high; gaster with five segments; AL 4–5.55 mm; FWL 8.5–13.7 mm = ♀ *Oecophylla atavina* Cockerell.
- 11 – Forewing with closed cells 1+2r, 3r, rm and mcu; male antennae filiform with very short scape. (Dolichoderinae). . . . .go to 12.
- Forewing without closed cell rm; 5RS and 4M starting from the same point (cross-vein rs–m lost). Antennae geniculate in both sexes, scape always extending beyond occipital margin. (Formicinae). . . . .go to 16.
- 12 – Head about as wide as alitrunk or a little narrower; antennae geniculate; gaster with five segments (♀♀) . . . . .go to 13.

- Head distinctly narrower than alitrunk; antennae filiform, with very short scape; petiole triangular in side view; gaster with six segments (♂♂). . . . . go to 14.
  - 13 – Head wider than long; scape short, not reaching occipital margin; petiole with scale 2.5–3 times as wide as thick; cell rm comparatively wide (1.6–2.1 times as long as wide); cross-veins 2r–rs and rs–m aligned at RS (4RS absent), or rs–m slightly distal, leaving 4RS as long as about vein thickness; BL 6–8.5 mm; AL 2.5–3.5 mm; FWL 6–7 mm = ♀ *Emplastus britannicus* (Cockerell).
  - Head longer than wide; petiole with low scale (about as long as high), triangular in side view; cell rm 2.2–2.4 times as long as wide; cross-veins rs–m noticeable distal on RS than 2r–rs, 4RS much longer than vein thickness; BL 5–6.5 mm; AL 2.2–3.2 mm; FWL 5–5.5 mm = ♀ *Emplastus gurnetensis* (Donisthorpe).
  - 14 – Cell rm 1.6–2.1 times as long as wide; cross-veins 2r–rs and rs–m aligned at RS, 4RS lost, or rs–m a little more distal, leaving 4RS about as long as vein thickness; BL ~ 6 mm; AL ~ 3 mm; FWL 5–5.6 mm = ♂ *Emplastus britannicus* (Cockerell).
  - Cell rm 2.2–2.8 times as long as wide; cross-vein rs–m noticeably distal on RS in respect of 2r–rs, 4RS distinctly longer than vein thickness. . . . . go to 15.
  - 15 – Cell rm 2.5–2.8 times as long as wide; BL 6.5 mm; AL 3 mm; FWL 5–6 mm = ♂ *Emplastus hypolithus* (Cockerell).
  - Cell rm 2.2–2.4 times as long as wide; BL ~ 4 mm; AL sim; 2 mm; FWL 4 mm. = ♂ *Emplastus gurnetensis* (Donisthorpe).
  - 16 – Scape attached distant from posterior margin of clypeus; forewing lacking closed cell mcu = *Camponotus cockerelli* (Donisthorpe).
  - Scape attached at the posterior clypeal margin. Cell mcu present. . . . . go to 17.
  - 17 – ♂: BL 2.2–3.8 mm; AL 1.0–1.7 mm; FWL 2.2–3.7 mm; petiole triangular in side view; genitals poorly exposed, with stipes short, narrow, rounded at top. ♀: BL 5.0–5.7 mm; AL 2.2–3.2 mm; FWL 5–5.5 mm; petiole with scale high and wide, 2.2–2.4 times as high as long and 2.5–3 times as wide as thick = *Leucotaphus gurnetensis* (Cockerell).
  - ♂: BL 4.0–4.5 mm; AL 1.4–2.0 mm; FWL 3.8–4.5 mm; petiole triangular in side view; genitals well exposed, with stipes long, triangular, rounded at top. ♀: FWL 7.4–7.5 mm = *Leucotaphus donisthorpei* sp. nov.
- 3.11.3.2. Key to wings**
- 1 – Forewings with cells 1+2r, 3r, rm and mcu closed. . . . . go to 2.
  - Closed cell rm absent. . . . . go to 13.
  - 2 – Cross-vein cu–a near cell mcu, so usually  $Icu < 1.45$  and  $Icua < 1.2$  (Ponerinae). . . go to 3.
  - Cross-vein cu–a displaced proximal, so usually  $Icu > 1.5$  and  $Icua > 1.2$ . . . . . go to 8.
  - 3 – Cell rm triangular. . . . . go to 4.
  - Cell rm quadriangular. . . . . go to 6.
  - 4 – Cell rm small, pedunculate, with long stem; FWL c. 9 mm;  $Icu = 1.3–1.45$ ;  $Icua = 1.13–1.14$  = *Taphopone aberrans* sp. nov.
  - Cell rm with no or very short stem. . . . . go to 5.
  - 5 – Cross-vein 2r–rs clearly proximal of rs–m; FWL c. 3 mm;  $Icu = 1.25–1.3$ ;  $Icua = 1.1$  = *Ponerites crawleyi* (Donisthorpe).
  - Cross-veins 2r–rs and rs–m coinciding on RS (4RS is absent) or rs–m a little distal leaving 4RS about as long as vein thickness; FWL c. 7 mm;  $Icu = 1.3–1.4$ ;  $Icua = 1.14–1.18$  = *Taphopone petrosa* sp. nov.
  - 6 – Cell cua lost; cross-vein cu–a distant from cell mcu, so as  $2M+Cu$  well developed and  $Icu \geq 1.4$ ;  $Icua \geq 1.13$ ; FWL 3–4 mm = *Ponerites hooleyi* sp. nov.
  - 2Cu leaving cell mcu at clear angle forming distinct cell cua; cross-vein cu–a close to mcu, resulting in  $Icu \leq 1.3$ ;  $Icua \leq 1.1$ . . . . . go to 7.
  - 7 – Cell 3r comparatively short, four times as long as wide; FWL 5.5–6 mm;  $Icu = 1.05$ ;  $Icua = 1.01$  (holotype) = *Taphopone macroptera* sp. nov.
  - Cell 3r more than four times as long as wide; FWL c. 3 mm;  $Icu = 1.1–1.3$ ;  $Icua = 1.08–1.10$ ; Hind wing with three longitudinal veins (RS, M and Cu) = *Taphopone microptera* sp. nov.
  - 8 – Cross-vein 2r–rs distal of rs–m (possibly an aberration, for normally it is rs–m which can be more distal); FWL c. 8 mm;  $Icu = 1.6$ ;  $Icua = 1.3$  = *Britaneuretus anglicus* (Cockerell) (Aneuretinae).
  - Cross-vein 2r–rs either proximal of, or coinciding with, rs–m. (Dolichoderinae). . . . . go to 9.
  - 9 – FWL 3–3.5 mm. Cell rm triangular, pedunculate (rarely with stem very short). Cross-vein 2r–rs coinciding with rs–m or a little distal of it = *Dolichoderus vectensis* Donisthorpe.
  - FWL > 4.5 mm. . . . . go to 10.
  - 10 – Cell rm 1.5–2.1 times as long as wide; cell rm triangular with very short or no stem; cross-veins 2r–rs and rs–m coinciding on RS (4RS absent), or rs–m distal a little (4RS about as long as vein thickness); FWL 5.5–7 mm;  $Icu = 1.55–2.2$ ;  $Icua = 1.2–1.4$ ; hindwing with two longitudinal vein (RS and Cu) = *Emplastus britannicus* (Cockerell).
  - Cell rm 2.2–2.8 times as long as wide. . . . . go to 11.
  - 11 – Cell rm 2.5–2.8 times as long as wide; cell rm quadrangular or triangular, not pedunculate; cross-vein rs–m noticeably distal on RS compared to 2r–rs; 4RS distinctly longer than vein thickness, rarely rs–m and 2r–rs coinciding on RS; FWL 5–6 mm;  $Icu = 1.6–2.1$ ;  $Icua = 1.3–1.4$  = *Emplastus hypolithus* (Cockerell).
  - Cell rm 2.2–2.5 times as long as wide. Cell rm triangular without stem. . . . . go to 12.
  - 12 – Cross-vein rs–m distinctly distal on RS compared to 2r–rs, 4RS distinctly longer than vein thickness; FWL 4–6 mm;  $Icu = 1.8$ ;  $Icua = 1.25$ ; hindwing with three longitudinal veins (RS, M and Cu) = *Emplastus gurnetensis* (Donisthorpe).
  - Cross-veins 2r–rs and rs–m coinciding on RS; FWL 7.8–12 mm;  $Icu = 1.6–1.8$ ;  $Icua = 1.3–1.5$  = *Emplastus kozlovi* sp. nov.
  - 13 – Cell mcu closed. . . . . go to 14.
  - Cell mcu absent (Formicinae). . . . . go to 18.
  - 14 – 4M leaving cell 1+2r long before 2r–rs and 5RS; cell 3r open apically, even if sometimes very narrowly and inconspicuously; FWL 3.5–4.5 mm;  $Icu = 1.5–1.7$ ;  $Icua = 1.2$  = *Paraphaenogaster hooleyana* sp. nov. (Myrmicinae).
  - 5RS and 4M with joint origin (no rs–m); cell 3r always closed;  $Icu = 2.0–3.6$ ;  $Icua = 1.2–2.2$  (*Leucotaphus*, Formicinae). . . . . go to 15.
  - 15 – FWL 2.2–3.7 mm = ♂ *L. gurnetensis* (Cockerell).
  - FWL > 3.7 mm. . . . . go to 16.
  - 16 – FWL 3.8–4.5 mm = ♂ *L. donisthorpei* sp. nov.
  - FWL > 4.5 mm. . . . . go to 17.
  - 17 – FWL 5–5.5 mm = ♀ *L. gurnetensis* (Cockerell).
  - FWL c. 7.5 mm = ♂ *L. donisthorpei* sp. nov.
  - 18 – 5RS always convex (cell 3r bent outward); FWL 5–8 mm; hindwing with cross-vein rs–cu subequal to  $2M+Cu$ , cross-vein cu–a near to wing base compared to rs–cu = *Camponotus cockerelli* (Donisthorpe).

**Table 2** Taxonomic composition (numbers and % of total) of ants in the Bembridge Marls.

Taxa	All fossils		Identified up to species level	
	n	%	n	%
<b>Aneuretinae</b>				
<i>Britaneuretus anglicus</i> (Cockerell)	1	0.1	1	0.2
<b>Dolichoderinae</b>				
<i>Dolichoderus vectensis</i> Donisthorpe	7	0.6	6	1.1
<i>Emplastus britannicus</i> (Cockerell)	52	4.6	52	8.3
<i>Emplastus hypolithus</i> (Cockerell)	15	1.3	15	2.4
<i>Emplastus gurnetensis</i> (Donisthorpe)	8	0.7	8	1.3
<i>Emplastus kozlovi</i> Perfilieva, sp. nov.	5	0.4	5	0.8
<i>Emplastus</i> sp.	1	0.1		
<b>Formicinae</b>				
<i>Camponotus cockerelli</i> (Donisthorpe)	15	1.3	15	2.4
<i>Camponotus</i> sp.	1	0.1		
<i>Leucotaphus gurnetensis</i> (Cockerell)	158	13.9	158	25.1
<i>Leucotaphus donisthorpei</i> Dlussky, sp. nov.	16	1.4	16	2.5
<i>Leucotaphus</i> sp.	1	0.1		
<i>Oecophylla atavina</i> Cockerell	199	17.5	199	31.6
<i>Oecophylla megarche</i> Cockerell	124	10.9	124	19.7
<i>Oecophylla</i> sp.	397	34.9		
<b>Ponerinae</b>				
<i>Ponerites crawleyi</i> (Donisthorpe)	7	0.6	7	1.1
<i>Ponerites hooleyi</i> Dlussky et Perfilieva, sp. nov.	3	0.3	4	0.5
<i>Ponerites antropovi</i> Dlussky, sp. nov.	1	0.1	1	0.2
<i>Ponerites</i> sp.	1	0.1		
<i>Taphopone macroptera</i> Perfilieva, sp. nov.	2	0.2	2	0.3
<i>Taphopone microptera</i> Perfilieva, sp. nov.	2	0.2	2	0.3
<i>Taphopone petrosa</i> Perfilieva, sp. nov.	6	0.5	6	1.0
<i>Taphopone aberrans</i> Perfilieva, sp. nov.	6	0.5	6	1.0
<i>Taphopone</i> sp.	1	0.1	1	0.2
<b>Myrmicinae</b>				
<i>Solenopsis rossi</i> Dlussky, sp. nov.	1	0.1	1	0.2
<i>Paraphaenogaster hooleyana</i> Perfilieva, sp. nov.	2	0.2	2	0.3
<i>Paraphaenogaster</i> sp.	1	0.1		
<b>Formicidae</b> indet.	105	9.2		
<b>TOTAL</b>	1136	100.1	630	100.5

- 5RS always concave (bent into cell 3r); hindwing with rs-cu much longer than 2M+Cu and with cu-a more close to rs-cu than to wing base. . . . .go to 19.
- 19 – FWL 6–9.5 mm = ♂ *Oecophylla atavina* Cockerell.
- FWL > 9.5 mm. . . . .go to 20.
- 20 – FWL 20–23 mm = ♀ *Oecophylla megarche* Cockerell.
- FWL 10–16 mm. . . . .go to 21.
- 21 – Apical corner of cell 1+2r obtuse, rarely *c.* 90°; FWL 10–16 mm = ♂ *Oecophylla megarche* Cockerell
- Apical corner of cell 1+2 more acute; FWL 8.5–13.7 mm. = ♀ *Oecophylla atavina* Cockerell.

**3.11.4. Discussion.** The Bembridge Marls Insect Bed has yielded 20 ant species belonging to ten genera and five sub-families (Table 2). The most striking feature of the assemblage is the hyperabundance of *Oecophylla*, not recorded in any other circumstance in the past. *Oecophylla* fossils represent 63.4% of all the ants collected and 69.8% of those identified up to genus level. There is no question that the figures are exaggerated, because these ants are generally of larger size and rather striking general appearance, which make them easier to be collected and identified, even from small fragments. The collecting bias is not easy to account for; unlike the identification bias, which can be partially taken into account due to considering only the better preserved fossils which permit species level identification. This approach excludes 403 fossils identified as *Oeco-*

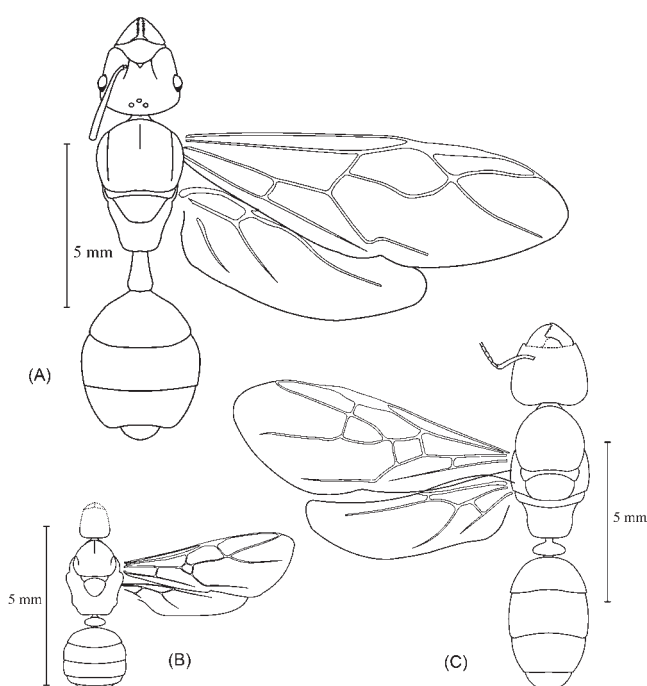
*phylla* sp. and 111 as Formicidae indet. The resulting figures are a bit lower, with *Oecophylla* making up 51.3% of ants (31.6% – *O. atavina* and 19.7% – *O. megarche*) while the second most abundant species, *Leucotaphus gurnetensis*, comprises 25.1%, and the fourth, *Emplastus britannicus* is 8.3%. The general appearance of females of the most abundant ant species are reconstructed in Text-figure 88. All remaining species identified in the assemblage are much rarer, each with its share being under 3% of the ant fauna.

The above data make it possible to draw some inference on the environments near the target water body of the Bembridge Marls insect burial. No doubt the landscape was rich in forest, as the abundance of dendrobiotic ants suggests. These dendrobionts form the most common three species (both species of *Oecophylla* and *Emplastus britannicus*) whose joint share is 59.5% of all ants. Their morphology discussed above, as well as the biology of their living relatives or analogues, indicates their dendrobiotic habits with considerable confidence. Judging from habits of extant relatives, *Dolichoderus vectensis* and *Camponotus cockerelli* were dendrobiotic as well: they are responsible for 3.5% of the ant assemblage.

Bembridge Ponerinae, which make up 4.3% of the assemblage, were most probably forest floor dwellers, judging from both their morphology (forward position of female eyes) and analogy with Ponerinae in the Baltic amber and other late

**Table 3** Composition (% of total) of ant subfamilies in better known fossil assemblages

Subfamily	Late Cretaceous (all data)	Eocene				Oligocene	Miocene	
		Green River	Baltic amber	Bembridge Marls	Florissant	Sicilian amber	Radoboj	Visnevaja balka
Aneuretinae	1.9	3.1	0.3	0.2	0.02	0	0	0
Dolichoderinae	1.9	75.0	64.1	13.8	62.6	7.7	13.1	3.9
Formicinae	1.9	3.1	32.8	81.3	32.5	35.9	64.2	52.6
Ponerinae	1.9	10.4	0.7	4.3	0.04	5.1	0.7	2.6
Cerapachyinae	0	0	0.1	0	0	0	0	0
Myrmeciinae	0	6.3	0.1	0	0	0	0	0
Pseudomyrmecinae	0	0	0.2	0	0.04	0	0	0
Myrmicinae	1.9	2.1	1.7	0.5	4.8	51.3	21.9	40.8
Armaniidae	36.9	0	0	0	0	0	0	0
Sphecomyrminae	47.2	0	0	0	0	0	0	0
Brownimeciinae	3.8	0	0	0	0	0	0	0
Specimens studied	53	96	10919	630	5592	39	137	76

**Text-figure 88** Restored general appearance of female ants most common in the Bembridge Marls: (A) *Oecophylla atavina* Cockerell; (B) *Leucotaphus gurnetensis* (Cockerell); (C) *Emplastus britannicus* (Cockerell).

Eocene European ambers, which certainly formed in forested areas. Of all Ponerinae found (except for *Bradoponera meieri* Mayr, 1867), 94% are winged sexual forms, which implies that their workers do not normally visit tree trunks and kept to the forest floor, unlike sexual forms which probably started their nuptial flight from trees and other high points.

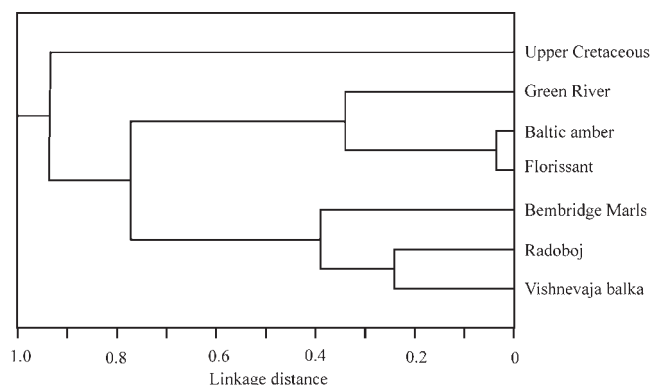
Hence, at least two thirds of the Bembridge ant assemblage consists of forest ants. Of the remaining species, the second most abundant, *Leucotaphus gurnetensis*, most probably resembles *Lasius*, which is a eurybiotic ant genus commonly found in forests, meadows and steppe. In the late Eocene European ambers, *Lasius schiefferdeckeri* Mayr, 1867, is the second most abundant ant species (after *Ctenobethylus goepperti* (Mayr, 1867)).

The Bembridge ants indicate a warm climate. Extant *Oecophylla* is characteristic of tropics and southern subtropics. One

of the authors (GMD) had a chance to observe *O. smaragdina* introduced from China to North Caucasus (Lazarevskoye Insectarium, All-Union Institute of Plant Protection). After a successful winter (1963/64) spent in a greenhouse, the ants did well during the summer in the open air in a garden, including raising larvae, but they became inactive when the temperature dropped below 18°C. Unfortunately, they were not returned to the greenhouse in time, so they died out when the temperature dropped below zero. Since *Oecophylla* makes its nest only from living leaves, they can only populate forests with evergreen angiosperm trees. We have no reason to hypothesise different habits in the past, except that the ants could have existed in a somewhat less hot climate than now: they are found in the late Miocene of Oeningen in Germany, where the climate is reconstructed as warm temperate rather than subtropical (Mai 1995).

The above results enable a possible comparison of the Bembridge assemblage with some other past ant assemblages. In this comparison, we rely on the following sources: for Late Cretaceous – Dlussky (1975, 1983, 1987, 1996, 1999a,b), Dlussky *et al.* (2004), Grimaldi *et al.* (1997), Grimaldi & Agosti (2000), Engel & Grimaldi (2005); for Green River (Middle Eocene of western North America) – Dlussky & Rasnitsyn (2002), for Baltic amber (Russia and Poland, Late Eocene) – Wheeler (1915) and our unpublished observations; for Florissant (latest Eocene, Colorado, USA) – Carpenter (1930); for Sicilian amber (Upper Oligocene of Italy) – Emery (1891); for Radoboj (Lower Miocene of Croatia) – Mayr (1867) with our additions; for Vishnevaja Balka (Middle Miocene of Stavropol, Russia) – Dlussky (1981a).

The comparison traditionally refers to composition of past assemblages at the subfamily level (Table 3). It has been shown (Dlussky 1983, 1987; Dlussky & Rasnitsyn 2002) that the Late Cretaceous assemblages were dominated by extinct Armaniidae and Sphecomyrminae, with extant subfamilies Aneuretinae, Dolichoderinae, Formicinae, Ponerinae and Myrmicinae known from rare (usually unique) plesiomorphic fossils. Since the end of the Late Cretaceous, Armaniidae and Sphecomyrminae are no longer found in the fossil record; which starts containing only extant subfamilies, with the sole exception of the short-living (Early and Middle Eocene only) monotypical giant Formicinae, and subfamilies became the highest-level changes in composition of the ant assemblages. The Palaeocene (Sakhalin amber), Eocene (Green River and European ambers) and possibly in Oligocene (except the zoogeographically different – Afrotropical – Sicilian amber, and



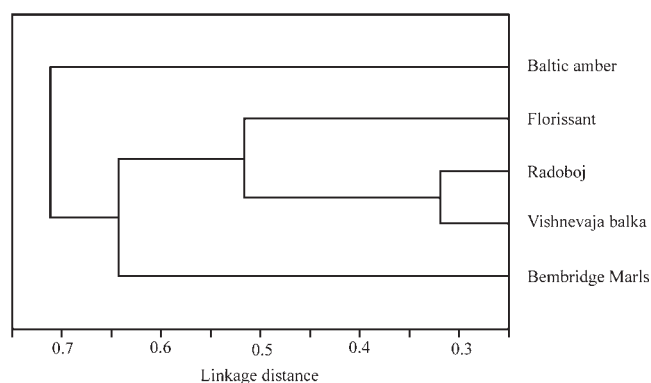
**Text-figure 89** Dendrogram of similarity of the subfamily level composition of the best known ant rock fossil assemblages in the unweighted pair-group average.

supposing the Florissant assemblage to be of Oligocene rather than uppermost Eocene in age) are characterised by dominating Dolichoderinae and rare Myrmicinae. It is only in the Miocene (or in the Late Oligocene, if Sicilian amber is taken into consideration) when the subfamily level composition of the ant assemblages approached the contemporary pattern, with dominating Formicinae and Myrmicinae and far subordinated Dolichoderinae.

Table 3 shows that the Bembridge assemblage differs somewhat from the above sequence in that it is strictly dominated by Formicinae (81.3%, the highest percentage ever observed throughout the known ant history), Dolichoderinae far subordinate (13.8%), and Myrmicinae (0.5%) being the rarest. The dendrogram based on cluster analysis shows this pattern particularly clearly (Text-fig. 89). Cretaceous assemblages are naturally the most distinct, and the Cenozoic ones form two clusters; one composed of the Baltic amber and Florissant, and a little more distant Green River, and the other cluster embracing the Bembridge and Miocene assemblages. The particular similarity of the Baltic amber and Florissant is based on their virtually identical shares of Formicinae and Dolichoderinae; that of the second cluster is the result their high share of Formicinae, and the still higher similarity of Vishnevaya and Radoboj depends on the high participation of Myrmicinae. Hence, the geochronologically similar Bembridge and Florissant faunas reveal quite different relationships – the former closer to Miocene, and the latter to Eocene assemblages.

However, the pattern does not seem convincing, particularly when considering the generic level composition of the assemblages. Indeed, Florissant is dominated by *Protazteca* (28.2%) which has no analogue in Baltic amber, and *Liometopum* (30.4%) known in Baltic assemblage from two specimens of the same species. Unlike Florissant, the Baltic assemblage is dominated by *Ctenobethylus* (47.0%) unknown at Florissant. This makes it important to make a wider generic level comparison; which is difficult, however, because the Bembridge assemblage is rich in morphogenera not directly comparable to genera in other assemblages.

The problem might be resolved to some extent by relying partly on intuition, comparing the similarity of morphogenera to particular ant orthogenera. In particular, we associate *Emplastus* with *Iridomyrmex* s.l. (*Iridomyrmex* + *Anonychomyrma* + *Ctenobethylus*), *Leucotaphus gurnetensis* with *Lasius*, *Leucotaphus donisthorpei* with *Prenolepis*, and *Ponerites* + *Taphopone* with *Ponera* + *Hypoconera* + small-sized *Pachycondyla*. The Cretaceous and Green River assemblages are excluded from consideration because these genera are absent there, and Green River is dominated by the morphogenus



**Text-figure 90** Dendrogram of similarity of the genus level composition of the best known ant rock fossil assemblages in the unweighted pair-group average.

*Eoformica*, which is impossible at present to associate with orthogenera because of poor preservation.

The resulting dendrogram (Text-fig. 90) shows the Baltic amber assemblage to be isolated while Florissant is closer to the Miocene assemblages than Bembridge is; apparently because of a higher share of *Lasius* (25.0% in Florissant, 33.3% in Radoboj and 34.2% in Vishnevaya, as opposed to 13.9% of *Leucotaphus gurnetensis* in Bembridge).

The isolated position of the Baltic amber assemblage is apparently taphonomically based (Dlussky & Rasnitsyn 2002). The ants trapped in resin are mostly workers running about on tree trunks, with those of smaller size having more chance to be trapped there. As a result, larger ants and soil and litter dwellers have less chance to become amber inclusions. Unlike them, to become impression fossils, an ant has to be buried in the bottom sediments of an ancient lake. The allochthonous insects (those living outside the target lake) find their way to future burial mostly from the air (Zherikhin 1980). That is why the chance to become an impression fossil is higher for winged ants, and particularly for those in aerial swarms (mating flight with copulation starting in air, as opposed to the ground swarm where males and females copulate only after the dispersal flight is over (Kannowski 1963)). Aerial swarms often involve numerous ant sexual forms, travelling far from their start area (swarms can be found tens of kilometres beyond the ants' northern limits as recorded in tundra (Dlussky 1967)). In contrast, ground swarming female ants (for example, *Formica* L.) execute a comparatively short flight, while males are flying low over the ground to seek females by their pheromones. Thus their potential to become impression fossils is lower.

Hypothesising the burial of ants inhabiting the source environments of the late Eocene European ambers (Baltic and others) in water bodies whose deposits would become fossiliferous rocks, we would expect the resulting ant assemblages to be much different than those in the ambers. Firstly, they would probably have a higher proportion of *Oecophylla*. Baltic amber has yielded two species of the genus, jointly producing less than 1% of the assemblage. This is not surprising, as these ants are large and long-legged, and more likely to escape resin traps compared to other ants. Still more importantly, they are tightly connected with angiosperm trees and their workers would have had little reason to visit the amber source trees, which were conifers. Secondly, the proportion of *Ctenobethylus* would be much lower. Alates of this genus are noticeably rare in amber – 0.3% (n = 5685; figures in this paragraph summarise those by Mayr 1868, Wheeler 1915, and our unpublished observations). Other ants, both supposed dendrobiotic and herpetobiotic (forager on the surface of soil), show a higher share of

their sexual forms: herpetobiotic *Lasius schiefferdeckeri* Mayr – 3.7% (n = 1719), *Formica* spp. 9.9% (n = 1448), *Prenolepis henschei* Mayr – 10.3% (n = 715); herpetobiotic or dendrobic *Iridomyrmex geinitzi* (Mayr) – 2.6% (n = 1260); specialised dendrobic *Tetraponera* spp. – 3.3% (n = 61); and *Dolichoderus* spp. – 4.6% (n = 653). As to geo- and stratiobiotic ants (foraging within soil and leaf litter respectively), their amber inclusions are almost invariably alates: *Paratrechina pygmaea* (Mayr) – 98% (n = 107), *Hypoponera atavia* (Mayr) – 94% (n = 52). This is quite natural, because their workers rarely leave the soil and leaf litter, unlike sexual forms that climb use trees to start their nuptial flight.

The exceedingly high proportion of worker inclusions in the case of *Ctenobethylus* has only one explanation: this ant nested in old conifer trees, had huge families and made very active trails towards colonies of symbiotic aphids. This inference is confirmed by the common syninclusions of these ants with *Germaraphis* aphids (Perkovsky 2006) and the high proportion of multiple inclusions of *Ctenobethylus* in a piece of amber. Our observations produced a figure of 9.2% (56 out of 608), with more than one worker per piece (up to 11); and Wheeler (1915) noticed numerous such cases, including seven pieces with more than ten workers each (up to 50). As already mentioned, *Ctenobethylus* is apparently a close analogue (the closest relative) of *Emplastus britannicus*, the fourth most common ant in the Bembridge assemblage, with a share of 10.1%.

The third most likely feature of the fossil ant assemblage expected in rocks resulting from an ant community populating the amber forest is a lower proportion of *Formica*. This inference can be deduced from the observation that these ants do not form nuptial swarms and thus have a comparatively low chance of being buried in a target water body. *Formica* is absent from the Vishnevays Balka, although there is little doubt that these ants lived near the water body of burial. The Bembridge Marls apparently lack ant fossils referable to that genus (there are no large *Leucotaphus* there comparable to the *Formica* sexual forms). In contrast, 10% of the Baltic assemblage are *Formica* ants, mainly *Formica flori* Mayr, 1868 and *Formica gustawi* Dlussky, 1967.

Thus, the expected rock fossil assemblage of ants from the Late Eocene amber forest would show considerable similarity to the Bembridge assemblage. This would then only highlight differences between the European (Baltic and Bembridge) and American (Florissant) assemblages, as follows:

1. *Protazteca* is the most abundant ant group in Florissant (28.3%), but neither the genus nor its analogues are known in the present and past in Europe.
2. In contrast, *Oecophylla* is common in the European Eocene, Oligocene and Miocene, as well as in the contemporary tropics of the Old World, but has never been recorded in the past and present Americas.
3. The Florissant assemblage lacks small Ponerinae dwelling within the leaf litter (*Ponera*, *Hypoponera*, *Ponerites*), which are well represented in the Cenozoic assemblages of Europe.

In contrast, the Bembridge ant assemblage shows significant similarity to those in the European Miocene. As already mentioned, the second most abundant Bembridge species, *Leucotaphus gurnetensis*, is probably similar to *Lasius*; the Vishnevaya Balka assemblage has *Lasius vetulus* Dlussky, 1981, equally the second most common ant there, which probably deserves to be transferred to *Leucotaphus*. The Bembridge Marls contain ant wings attributable to *Paraphaenogaster*, which were previously known only from the Miocene of Europe (described from Vishnevays and also identified by GMD in the collections from Oeningan and Parschlung kept at the NHMUK) and China. *Aphaenogaster sommerfeldti*

Mayr, 1868 from Baltic amber and *A. mayri* Carpenter, 1930 from Florissant are not attributable to *Paraphaenogaster* because they have cell rm closed. It is quite possible that the wing described as *Camponotites macropterus* Dlussky, 1981 in fact represents *Oecophylla*, otherwise known in the European Miocene of Radoboj and Oeningen (*O. obesa* (Heer), Mayr, 1867). *Dolichoderus vectensis* of the Bembridge Marls belongs to the *D. quadripunctatus* group equally known in Baltic amber (Dlussky 2002) and the European Miocene (*D. haueri* (Mayr, 1867) from Radoboj, *D. tauricus* Dlussky, 1981 from Kerch, Ukraine) and not in the present and past Americas.

The above pattern indicates that the widely traceable generic level similarity of the Cenozoic (late Eocene through to Miocene) ant faunas shows its geographic orientation as more distinct than the temporal one. Within the above time interval, the Old World ant faunas are more similar to each other than to the synchronous American assemblages. For insects at least, this is a rather uncommon example of a geographically constrained evolution which inherits features of taxonomic composition of regional faunas for a long time, despite their intense evolution. There are further examples of this sort; e.g., the Myrmicinae ants whose evolution was apparently mostly confined to Africa (Dlussky *et al.* 2004), or honey bees which are confined to the Old World throughout all their rather long history (Engel 2006). This may indicate that evolution of highly social insects might be particularly geographically constrained compared to other insects. However, more detailed testing of this hypothesis lies far beyond the scope of the present project.

## 4. General discussion

### 4.1. Composition of the Bembridge Marls hymenopteran assemblage

1460 fossils were studied, which yielded a minimum of 118 species, counting those identified to any level, but only those that are evidently different from others identified to the same level. These 118 species represent 20 families and at least 78 genera.

In Ichneumonidae, 32 fossils were studied, including the types of 11 species described by Cockerell (1921a). One new genus and eight new species are described: *Exeristes gurnetor* Khalaim, sp. nov.; *Scambus fossilis* Khalaim, sp. nov.; *Eusterinx humalai* Khalaim, sp. nov.; *Hemiteles dirus* Khalaim, sp. nov.; *Hemiteles protervus* Khalaim, sp. nov., *Paxylobembra kozlovi* Khalaim, gen. et sp. nov.; *Marjorietta gigantea* Khalaim, sp. nov.; and *Lithapechtis salmacidus* Khalaim, sp. nov. The genus *Acourtia* Cockerell, 1921 is transferred to the subfamily Townesitinae. The following new combinations are established: *Eusterinx arcuatus* (Cockerell, 1921); *Hemiteles acurti* (Cockerell, 1921); and *Marjorietta disrupta* (Cockerell, 1921). The most abundant subfamilies are Cryptinae (five specimens), Pimplinae (five specimens), Townesitinae (three specimens) and Orthocentrinae (three specimens), and the genus *Lithapechtis* Cockerell, 1921 (three specimens) of uncertain subfamily assignment. The subfamilies Metopiinae and Paxyloommatinae are known from a single specimen each. Fossil Townesitinae and Paxyloommatinae are known from the Bembridge Marls and from Baltic amber only. Townesitinae are quite numerous and are very similar morphologically in both deposits. The ichneumonid fauna of the Bembridge Marls is close to that of the Baltic amber, and these two are apparently older than that of Florissant.

The Braconidae are represented by 13 subfamilies (Agathidinae, Brachistinae, Braconinae, Cheloninae, Doryctinae, Euphorinae, Exothecinae, Helconinae, Homolobinae, Microgasterinae, Opiinae, Orgilinae and Rhyssalinae) and 23 genera and 36 species are recorded. Two new genera and 26 new

species are described: *Bassus magnareola* Belokobylskij, sp. nov.; *Eubazus* ? *brodiei* Belokobylskij, sp. nov.; *E. flavistigma* Belokobylskij, sp. nov.; *E. ? grandareola* Belokobylskij, sp. nov.; *E. ? hooleyi* Belokobylskij, sp. nov.; *E. nanus* Belokobylskij, sp. nov.; *Bembracon acourtsmithi* Belokobylskij, gen. sp. nov.; *B. medialis* Belokobylskij, sp. nov.; *Bracon* ? *antefurcalis* Belokobylskij, sp. nov.; *B. brodiei* Belokobylskij, sp. nov.; *Ascogaster* (*Ascogaster*) *brodiei* Belokobylskij, sp. nov.; *A. (A.) yulei* Belokobylskij, sp. nov.; *A. (A.) pygmaea* Belokobylskij, sp. nov.; *Ontsira cenozoica* Belokobylskij, sp. nov.; *Meteorus applanatus* Belokobylskij, sp. nov.; *M. crassitergum* Belokobylskij, sp. nov.; *Chremylus infuscatus* Belokobylskij, sp. nov.; *Aspicolpus temporalis* Belokobylskij, sp. nov.; *Diospilus curtithorax* Belokobylskij, sp. nov.; *Hellenius* ? *kozlovi* Belokobylskij, sp. nov.; *Homolobus rasnitsyni* Belokobylskij, sp. nov.; *Palaeomicrogaster oculatus* Belokobylskij, gen. sp. nov.; *Semionis wightensis* Belokobylskij, sp. nov.; *Dolopsidea* ? *intermedia* Belokobylskij, sp. nov.; *Oncophanes andrewrossi* Belokobylskij, sp. nov.; and *Rhyssalus bruesi* Belokobylskij, sp. nov. The taxonomic status of *Syntaphus* Donisthorpe, 1920 is reduced to subgeneric level into *Ascogaster* Wesmael, 1835 (syn. nov.). The genus *Diospiloides* Cockerell, 1921 is a junior synonym of *Microtypus* Ratzeburg, 1848 (syn. nov.). The following new combinations are suggested: *Bassus quadrangularis* (Brues, 1933), *Bracon micrarche* (Cockerell, 1921), *Cyanopterus vectensis* (Cockerell, 1921), *Ascogaster wheeleri* (Donisthorpe, 1920), *Ontsira coleutti* (Cockerell, 1921), *O. distracta* (Cockerell, 1921), *Taphaeus cervicalis* (Cockerell, 1921), *Biosteres peritus* (Cockerell, 1921), and *Microtypus hooleyi* (Cockerell, 1921). The genus *Calypitoides* Cockerell is transferred to the subfamily Helconinae and the genus *Dacnusites* Cockerell to Microgastrinae. A discussion about contents, validity, generic and species combinations in the aforementioned subfamilies is provided.

In Aculeata s.str., other than ants, two new monotypic genera *Palaeopolybia* Antropov, gen. nov. for *P. anglica* (Cockerell, 1921), and *Protopolistes* Antropov, gen. nov. for *O. oblitus* (Cockerell, 1921) are established in Polistinae, and *Sphaerancistrocerus petiolatus* Antropov, gen. sp. nov. is described in Eumeninae (Vespidae). Also described are *Dryophia oculata* Antropov, gen. et sp. nov. (Dryophiinae, Tiphidae) and *Palaeoscolia relictata* Antropov, gen. sp. nov. (Palaeoscolinae Antropov, subfam. nov., Scoliidae), and *Oligobombus cuspidatus* Antropov, gen. et sp. nov. is described in Apinae (Apidae). Protosceliphriini Antropov, trib. nov. and *Protosceliphron* Antropov, gen. nov. are described for *Sceliphron brevior* Cockerell, 1921 in the family Sphecidae. *Tytthopsen nanus* Antropov, gen. et sp. nov. and *Plisomena gigantea* Antropov, gen. et sp. nov. are described in Pemphredoninae (Crabronidae).

The ant (Formicidae) assemblage studied includes 20 species, ten genera, and five subfamilies, of which nine species and two genera are described as new: *Emplastus kozlovi* Dlussky & Perfilieva, sp. nov.; *Leucotaphus donisthorpei* Dlussky & Perfilieva, sp. nov.; *Ponerites hooleyi* Dlussky & Perfilieva, sp. nov.; *P. antropovi* Dlussky & Perfilieva, sp. nov.; *Taphopone macroptera* Dlussky & Perfilieva, sp. nov.; *T. microptera* Dlussky & Perfilieva, sp. nov.; *T. petrosa* Dlussky & Perfilieva, sp. nov.; *T. aberrans* Dlussky & Perfilieva, sp. nov.; *Paraphaenogaster hooleyana* Dlussky & Perfilieva, sp. nov.; *Britaneuretus* Dlussky & Perfilieva, gen. nov.; and *Taphopone* Dlussky & Perfilieva, gen. nov. Nine new combinations are proposed: *Britaneuretus anglicus* (Cockerell, 1915); *Emplastus britannicus* (Cockerell, 1915); *E. hypolithus* (Cockerell, 1915); *E. gurnetensis* (Donisthorpe, 1920); *Campnotus cockerelli* (Donisthorpe, 1920); *Leucotaphus gurnetensis* (Cockerell, 1915); *Ponerites crawleyi* (Donisthorpe, 1920); *Taphopone karaganensis* (Dlussky, 1981); and *Taphopone stauropolitana* (Dlussky, 1981). Ants are the most common hyme-

nopteran family in the Bembridge Marls, differing from other fossil ant assemblages because of the overwhelming dominance of *Oecophylla* weaver ants. Composition of the ant assemblage indicates a warm climate, with temperature permanently above 0°C, and a forested landscape with abundant broad-leaved trees. At a generic level, past ant assemblages have more in common geographically than geochronologically. Also described is *Oxyserphus kozlovi* Kolyada, sp. nov. (Proctotrupidae).

Thus, the assemblage appears to be one of the richest among the rock fossil hymenopteran assemblages, even if it is second to the Florissant assemblage, which has always attracted much more attention, comprising 291 species, 161 genera and 37 families described but largely not yet revised (Meyer 2003). Out of 52 orthotaxonomical genera named in the Bembridge assemblage 18 are extinct (35%). Also extinct are all species, two tribes and three subfamilies. All species but one, two tribes and two subfamilies are endemic to the Bembridge Marls hymenopteran fauna, with one species and one tribe endemic jointly to the Bembridge Marls and the Baltic amber. Aculeate wasps and bees are highly endemic, with all genera and several higher taxa known from no other fossil site, nor from any contemporary fauna. Other groups are less specific in distribution. The assemblage is overwhelmingly dominated by winged ants (84%), an ordinary feature for the Cenozoic since the Eocene (Dlussky & Rasnitsyn 2002). These are followed by Braconidae (5.1%), Ichneumonidae (2.3%), Diapriidae (1.6%) and Scelionidae (0.8%), with the remaining families less than 0.5% each. The braconid dominance over Ichneumonidae is unusual: this is characteristic of assemblages of amber inclusions, primarily because of the generally smaller size of braconids that are more likely to be stuck in resin traps, as compared with the water surface trap that produces rock fossils. In the Cenozoic rock fossil assemblages, ichneumonid wasps are normally either much more common than the braconid ones (in the Paleocene of Tadushi and in the Eocene/Oligocene of Biamo, the Asian analogue of Florissant, both in the Russian far east; and in the Miocene of Stavropol in Ciscaucasian, personal observation by APR), or of roughly similar proportion (in the Eocene of Green River, western N. America, personal observation by APR; and near the the Oligocene/Miocene boundary of Rott in Germany; Statz 1936).

## 4.2. Palaeoenvironmental inferences

**4.2.1. Climate.** Palaeoclimatic inferences from hymenopteran fossils are somewhat contradictory, a usual case for the Paleogene (Archibald & Farrell 2003). Undoubtedly, the climate was warmer than it is now in southern England, as the absence of sawflies suggests, amongst many other indications. However, when looking at the details, we found some fossils of both tropical and warm subtropical affinities (Agaonidae fig pollinators, *Oecophylla* ants, Polybiini wasps), and those implying temperate conditions (at most a warm temperate climate), particularly the majority of Braconidae and several abundant ant taxa (noticeably *Leucotaphus gurnetensis*, which is similar to the genus *Lasius*). This contradiction can be resolved by employing the concept of an equable climate; that is, weakly seasonal even if not very warm, and so suiting both high and low thermophilous taxa (Archibald & Farrell 2003). This reveals another problem, however: the paradigmatic example of equable climate is that exemplified by the Baltic amber biota, which nevertheless includes an appreciable share of animals depending on a seasonable climate, particularly the aphids. The Bembridge Marls biota shows few aphids and not their specific parasites (Aphidiidae) and so better fits the concept of equability.

**4.2.2. Landscape.** Landscape signals from the Bembridge hymenopterans are also contradictory. The contradiction arises from the observation that some groups indicate a heavily forested territory, and the others imply extensive open meadows (A.J. Ross pers. comm., 2010). The former are primarily ants which are dendrobiotic, forming 60% of the fossil specimens, including those nesting in tree crowns and in tree trunks. 5% live in the forest floor leaf litter, with the remaining ants being mainly eurytopic. Additionally, fig-pollinating Agaonidae indicate the presence of fig trees, and there are several xylobiotic parasitic and aculeate wasps found there (Gasteruptionidae, some Braconidae and Crabronidae). Ichneumonidae recorded are predominantly forest dwellers, and Orthocentrinae are particularly indicative of humid forests. Diapriidae being more abundant than Scelionidae also suggest a forested environment. In contrast, the majority of Braconidae indicate dominance of open meadows, and about a half of aculeate wasps and bees are ground- and not wood-nesting.

**4.2.3. Biotic indication.** As already mentioned, Agaonidae wasps infer the presence of fig trees. *Oecophylla* ants are closely connected to broadleaf trees, and some Braconidae indicate this type of vegetation. Braconid and, to a lesser extent, ichneumonid wasps imply an abundance of moths, both macro- and particularly microlepidopterans, including leaf miners. This confirms the hypothesis that the rarity of the lepidopterans in the Cenozoic rock fossil assemblages is taphonomically caused, and does not necessarily imply their rarity in the source biocenoses. However, Jarzembowski (1980) described 21 moth taxa from the Bembridge Marls. Parasitic wasps infer the presence of various other insect hosts. Noteworthy is the comparative rarity of xylobiotic hosts and an absence of the aphidiid parasites of aphids, and of Alysiinae, the specialised parasites of higher (calyptrate) flies.

**4.2.4. Geography.** The majority of the fossils are either non-indicative in respect of large-scale biogeography, or otherwise fit the hypothesis of the predominantly Eurasian (Palaeartic + Oriental) character of the past fauna. This holds particularly true of ants that share more taxa with the Eurasian assemblages of different age (e.g. Miocene ones) than with the Florissant assemblage of similar age in N. America. Alternative indications are uncommon, including Ambositrinae (Diapriidae) which are mainly Australasian and Neotropical now, with one species Nearctic, one Afrotropical and three recorded in the Baltic amber, one being very close to the Afrotropical one. *Oxyserphys* (Proctotrupidae) is also mainly Australian, but known in the Oriental region up to Japan and in Central America as well. *Semionis* (Braconidae) is now restricted to S. Africa. As in many other cases, the “Gondwanan” distribution of the above taxa results most probably from contraction of the wider past distribution (Eskov 2002). As to the unexpectedly high level of endemism of aculeate wasps and bees in the Bembridge assemblage, which concerns all their genera and a few tribes and subfamilies, this effect shows no parallel in other hymenopteran taxa and so should be considered as characteristic of these aculeates themselves, rather than an indication of the isolated position of the source territory. Indeed, there are many genera there in common with the Baltic amber and Florissant, and one species (*Bassus quadrangularis*, Braconidae) and one subfamily (Townesitinae, Ichneumonidae) only occur in the Bembridge and Baltic assemblages.

**4.2.5. Geochronology.** Hymenopteran evolution is comparatively slow during the Cenozoic, with few higher rank taxa going extinct in that interval, and well dated cases of first appearances are uncommon. That is why the geological time correlation signals from the Cenozoic hymenopterans are rarely conclusive. Yet we have some signals of that sort; first of all, the presence of Townesitinae and the absence of Ichneu-

moninae (Ichneumonidae) from the Bembridge assemblage. Townesitinae is an extinct subfamily characteristic of Baltic amber and the Bembridge fauna and not recorded in younger strata, including the Florissant beds. Alternatively, Ichneumoninae are recorded neither in the Bembridge Marls, nor in the Baltic amber or in any older fossil sites, but they are common in the Florissant beds and also recorded from the Miocene. The same might also hold true of Alysiinae (Braconidae): this group is recorded in number from the Florissant and Miocene deposits, unlike the Bembridge fauna, the Baltic amber and all the older insect sites. However, all records of Alysiinae badly need revision and so should be used carefully. Also, the higher (calyptrate) Diptera, the alysiine hosts, have a generally similar time range. Taken altogether, these observations suggest that the Bembridge Marls might be older than the Florissant.

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